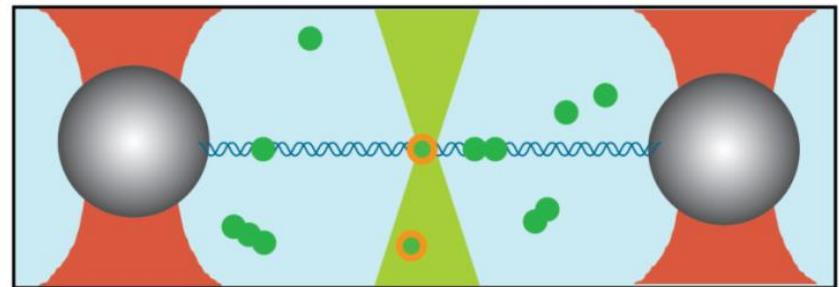
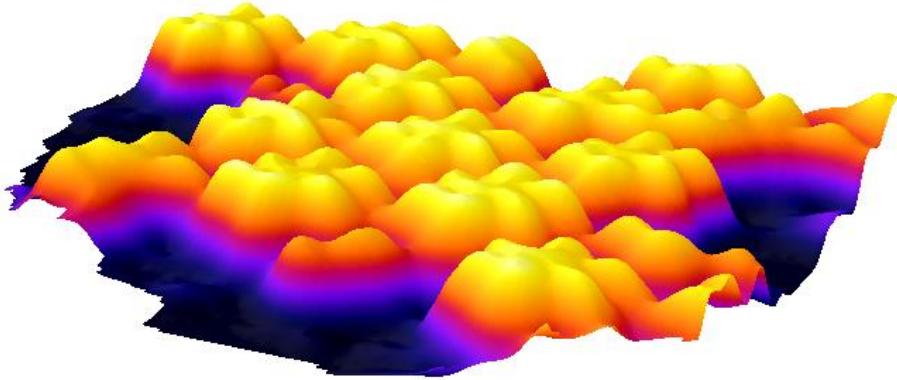
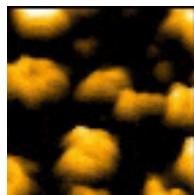


Single-molecule approaches to study viral self-assembly in real-time



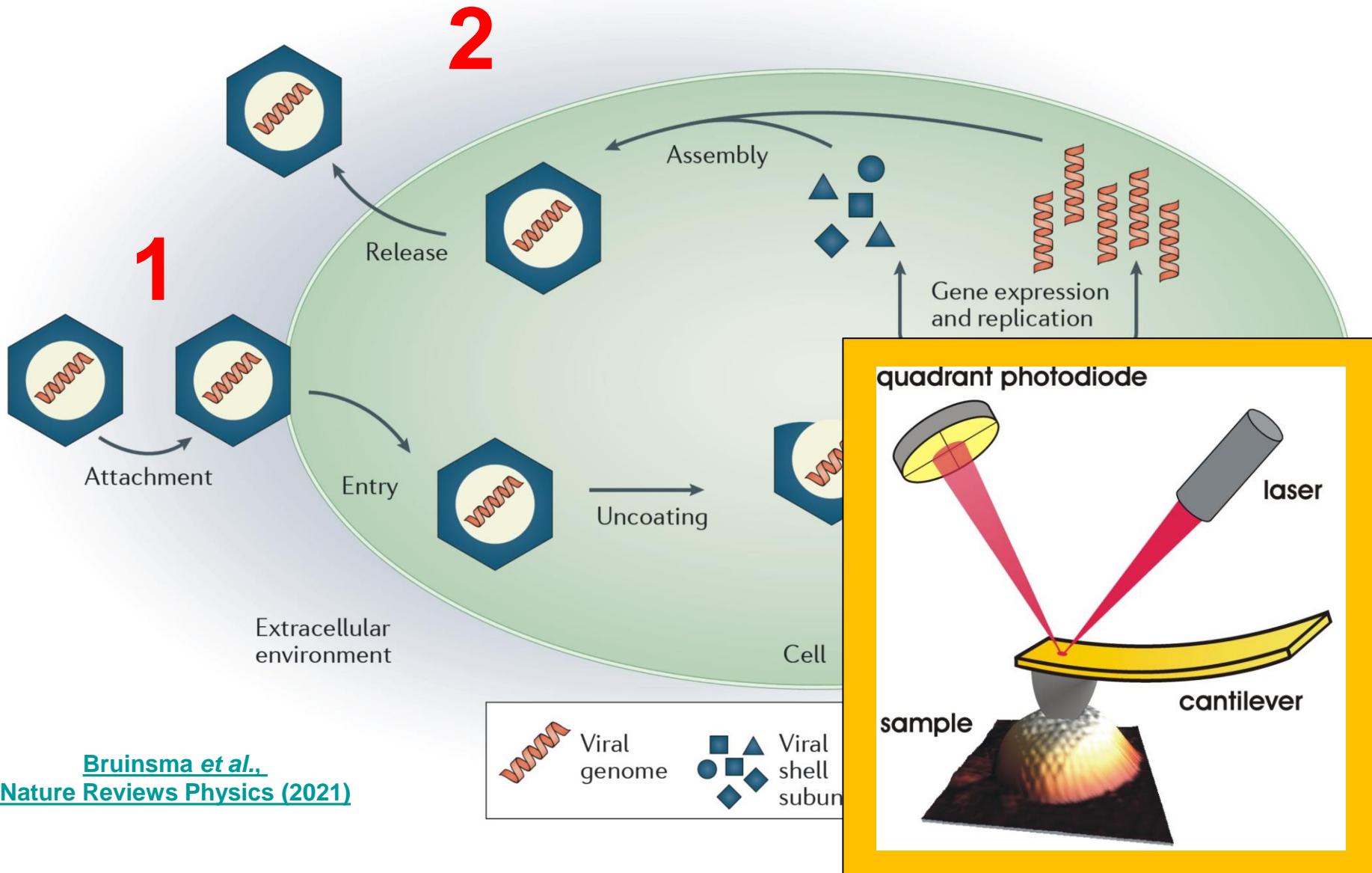
Wouter Roos



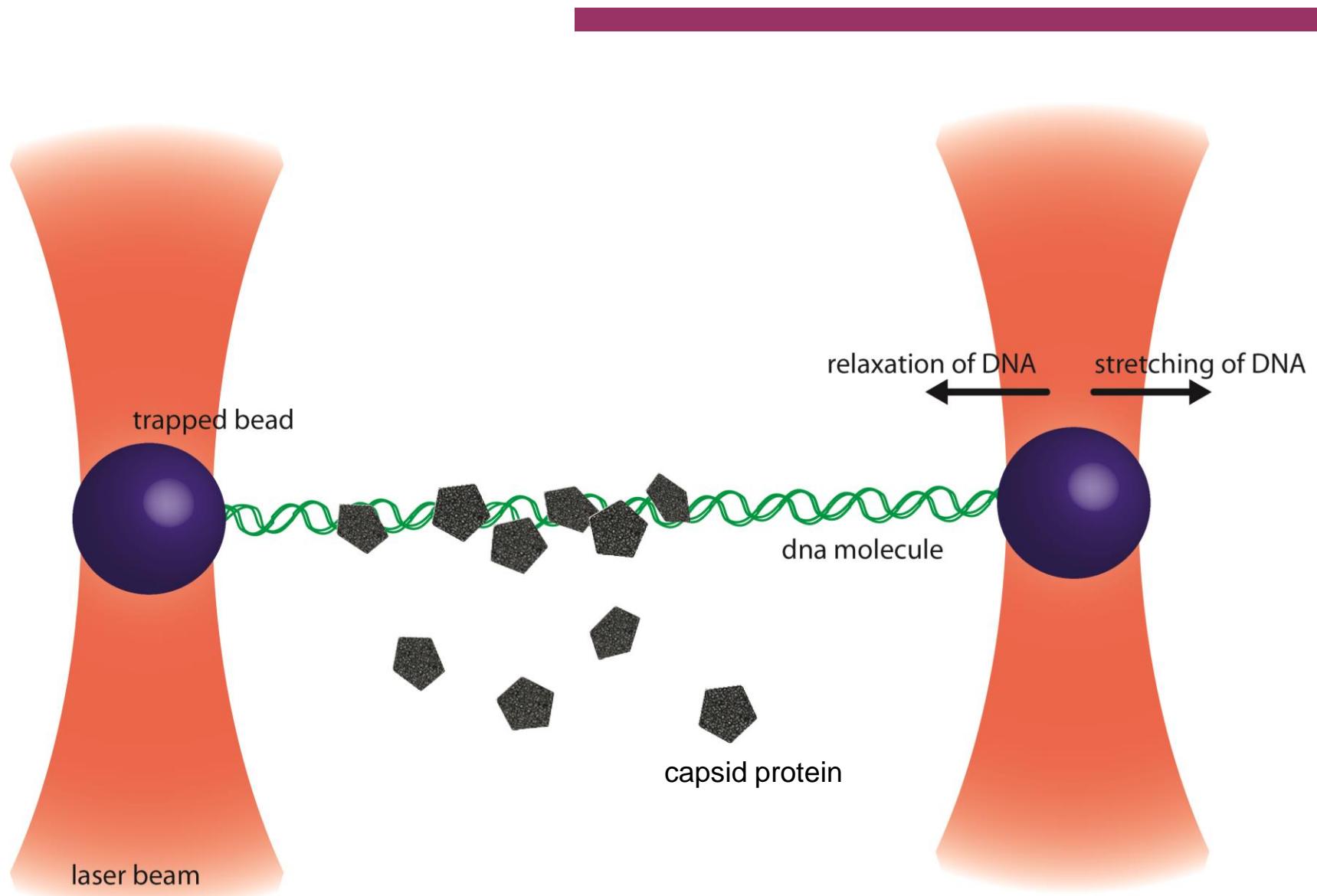
rijksuniversiteit
groningen

zernike institute for
advanced materials

Viral life cycle



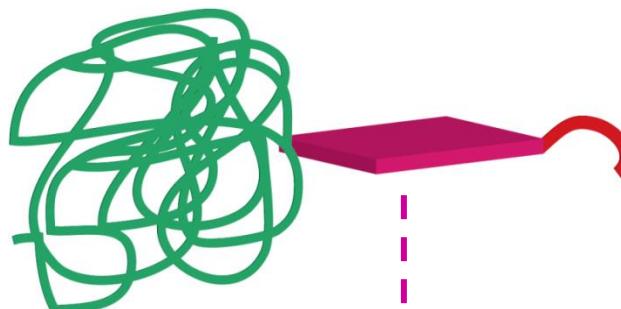
Following virus assembly by optical tweezers



Synthetic Virus-Like-Particles

Artificial
virus-like particles are
controlled systems

Our goal:
unravel the assembly process of a
rod-shaped artificial VLP



**Biosynthetic coat
protein: polypeptide**
44.8 kDa

Prevent aggregation of the assembled VLPs.
High fraction of prolines and hydrophilic
(mostly uncharged) amino acids:
~400-amino-acid- hydrophilic random-coil C

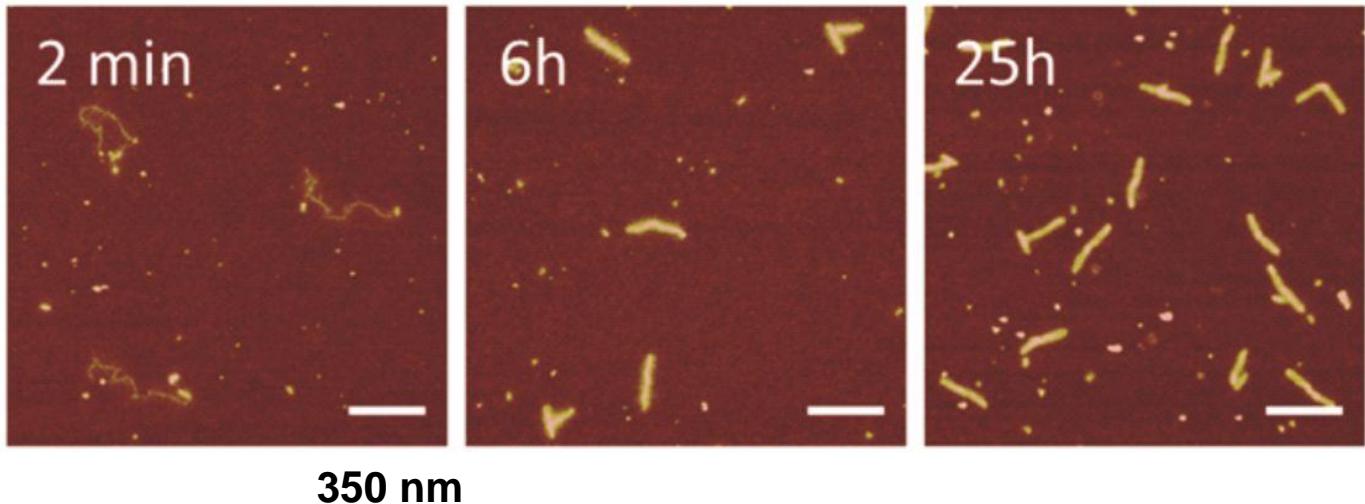
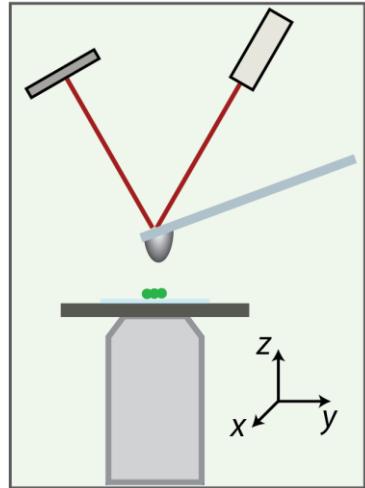
Folds and stacks in solution into stiff
filamentous structures.
Precisely tunes cooperativity:
Silk-like sequence $S_{10} = (GAGAGAGQ)_{10}$

Binds DNA non-sequence-specifically through electrostatic
interactions:
Oligolysine block B



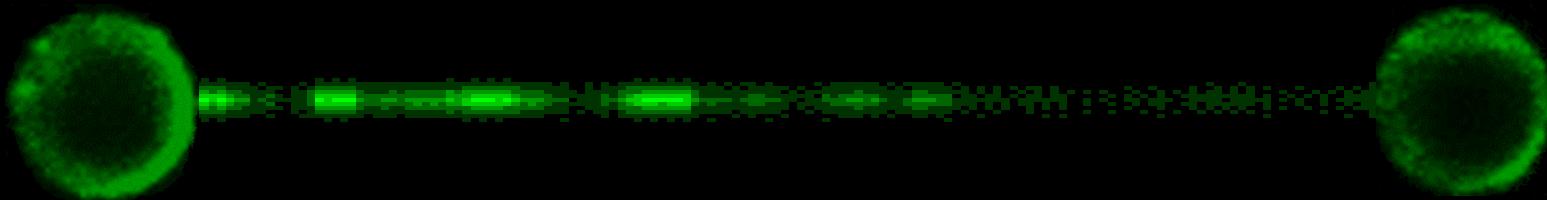
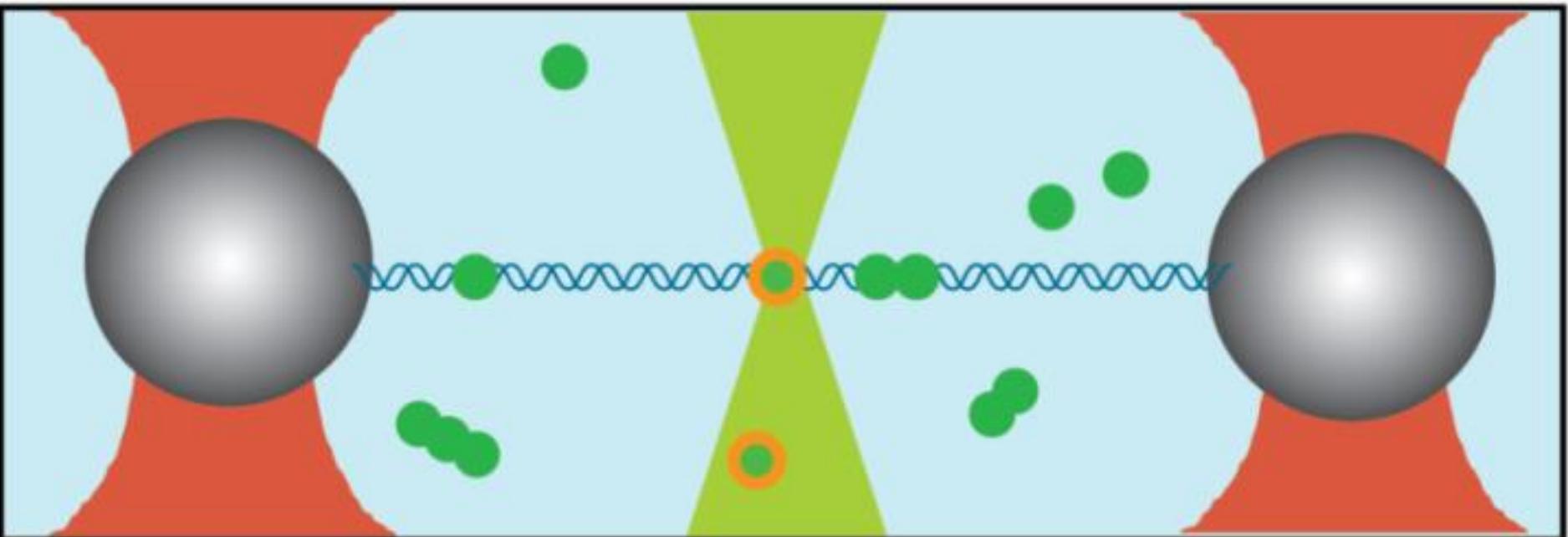
De Vries Lab:
A. H. Garcia et al., *Nature Nanotechnology* (2014)
B. Zhao et al., *Soft Matter* (2016)

Particle visualisation by AFM

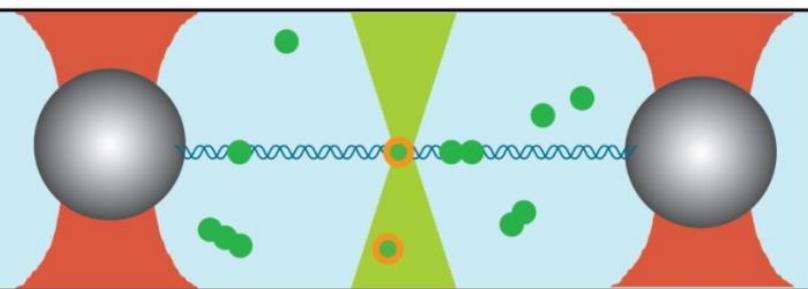


- Stiff rod-like appearance of the VLPs (300nm)
- DNA compacted 1/3 its original length
- Cooperative self-assembly.

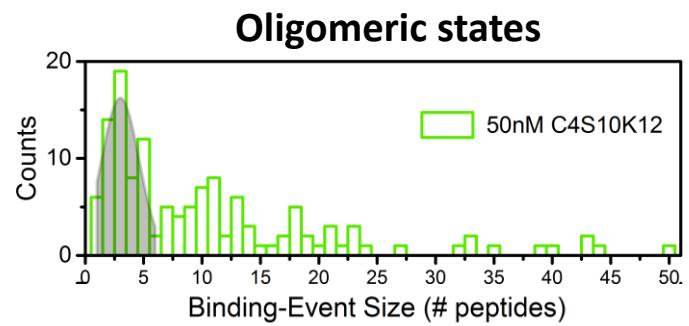
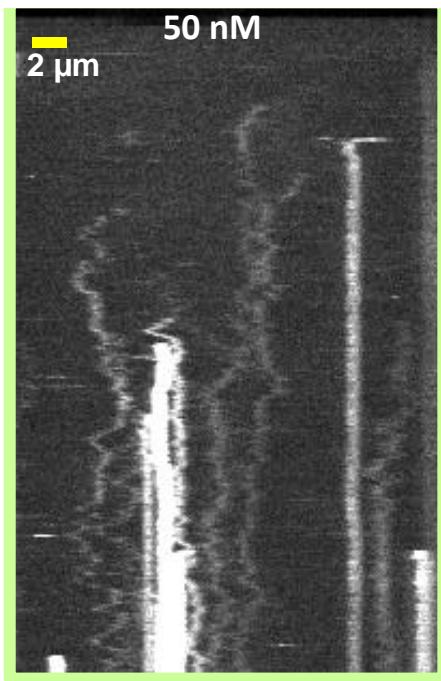
Protein attachment, in real-time



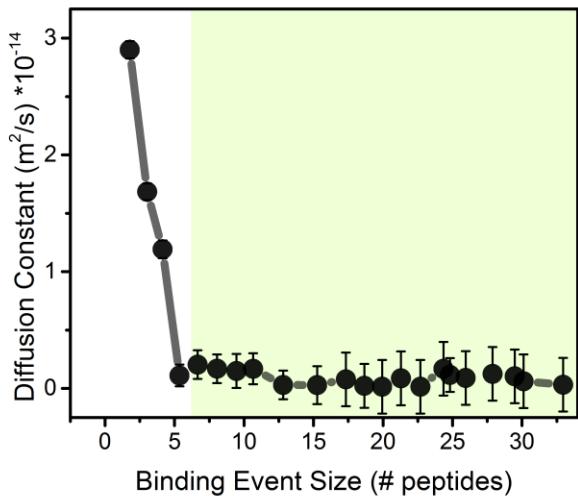
Protein attachment, in real-time



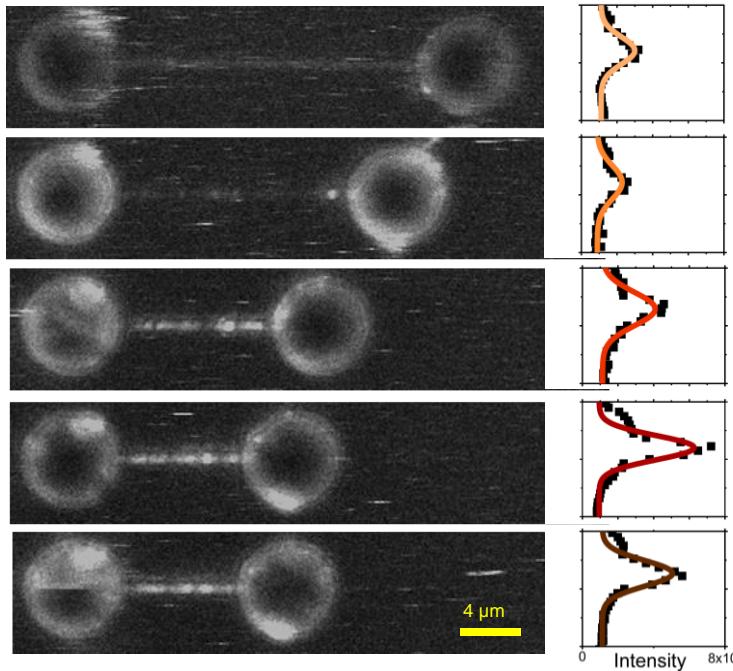
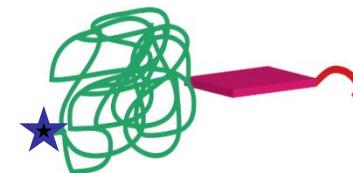
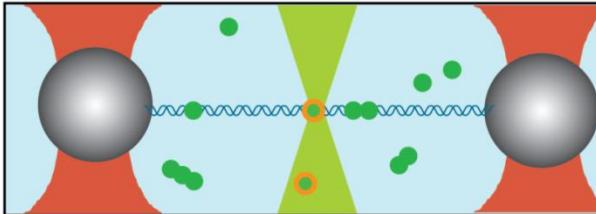
Time
↓



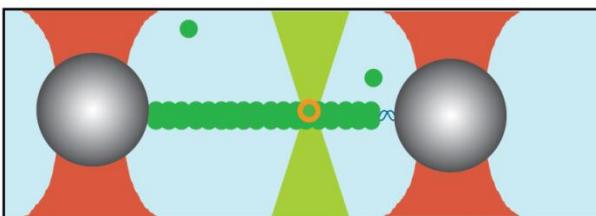
Diffusive events vs. Static Nuclei



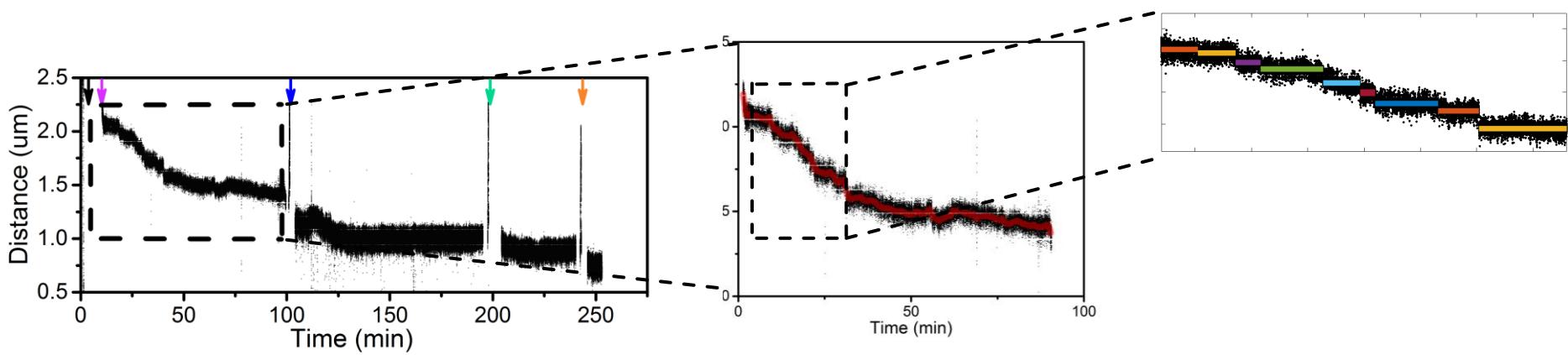
DNA compaction by rod shaped VLP



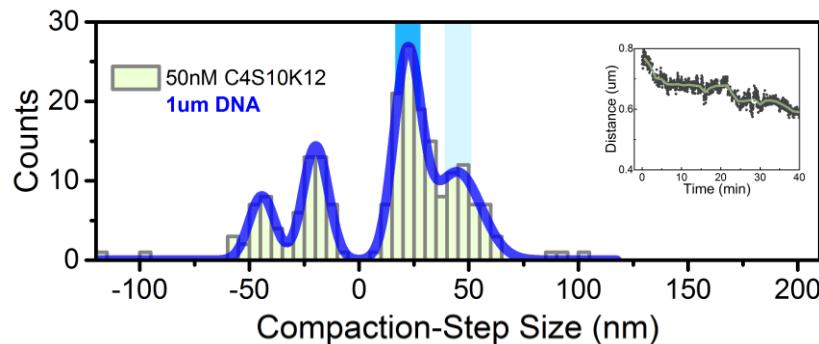
[Marchetti et al.](#)
[Nano Letters \(2019\)](#)



DNA compaction in real-time

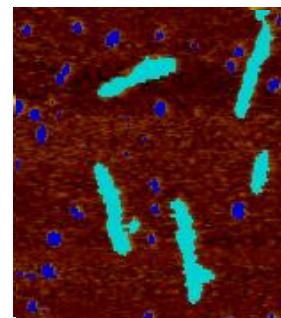
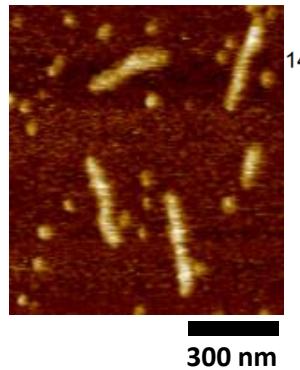


Regular compaction steps for assembly model

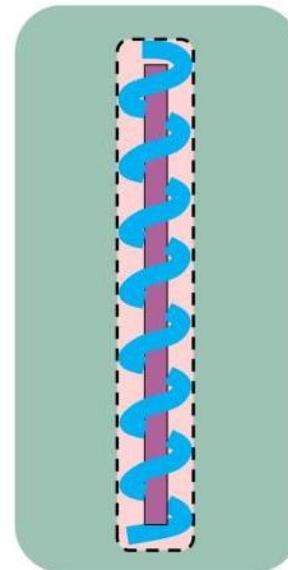


Average
Compaction-Step:

~ 30 nm

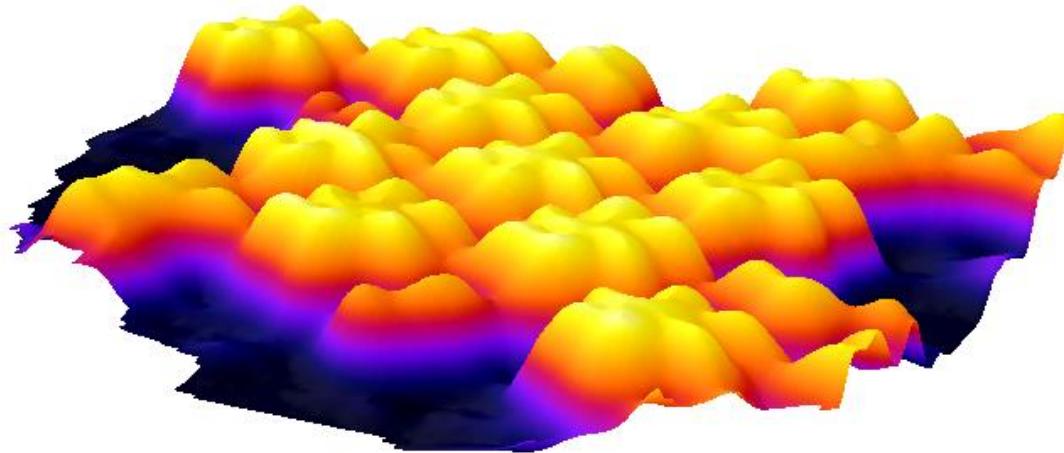


Mean particle
HEIGHT:
 9.1 ± 0.5 nm



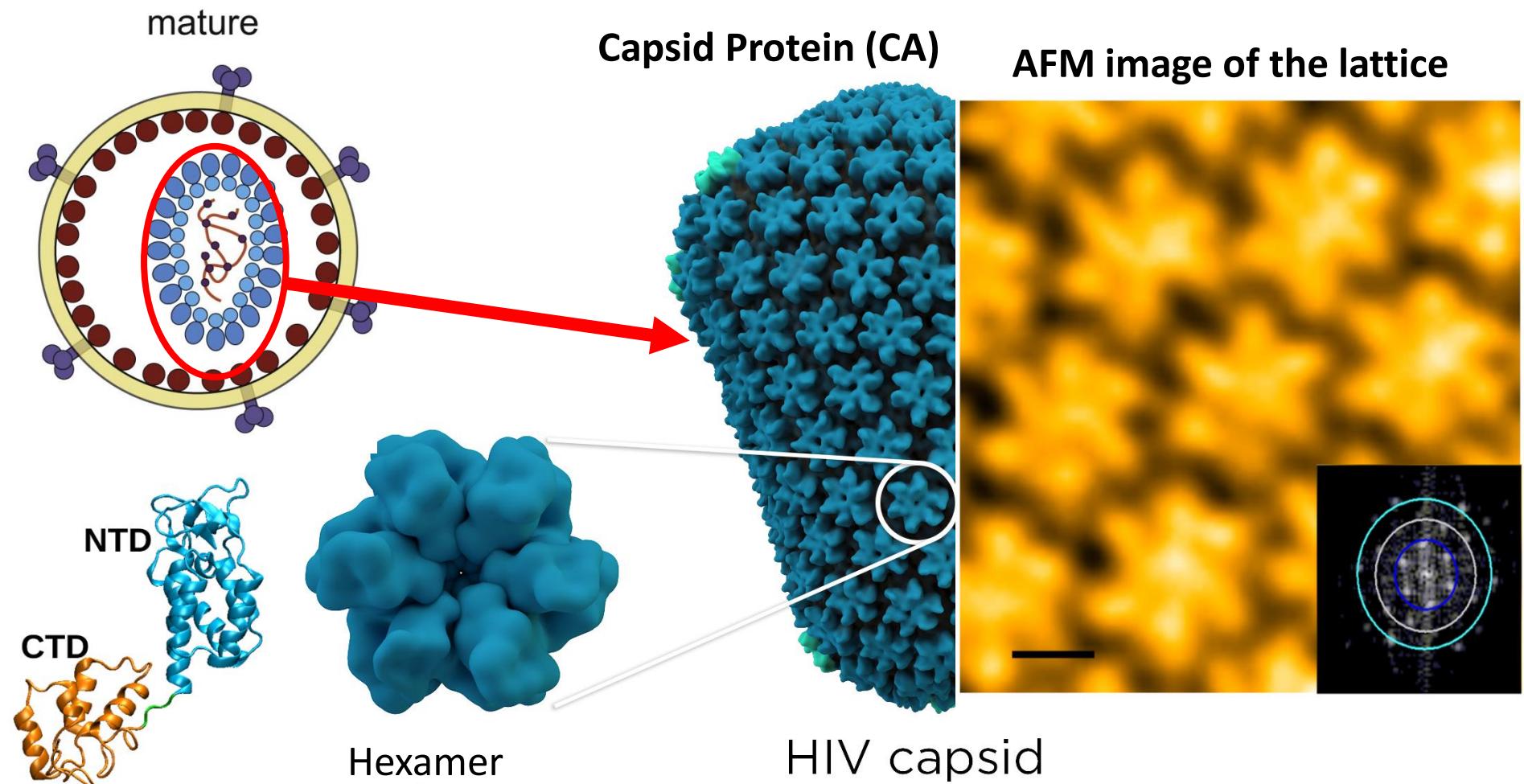
DNA
corona
silk core
region of DNA condensation

High Speed AFM studies of dynamics

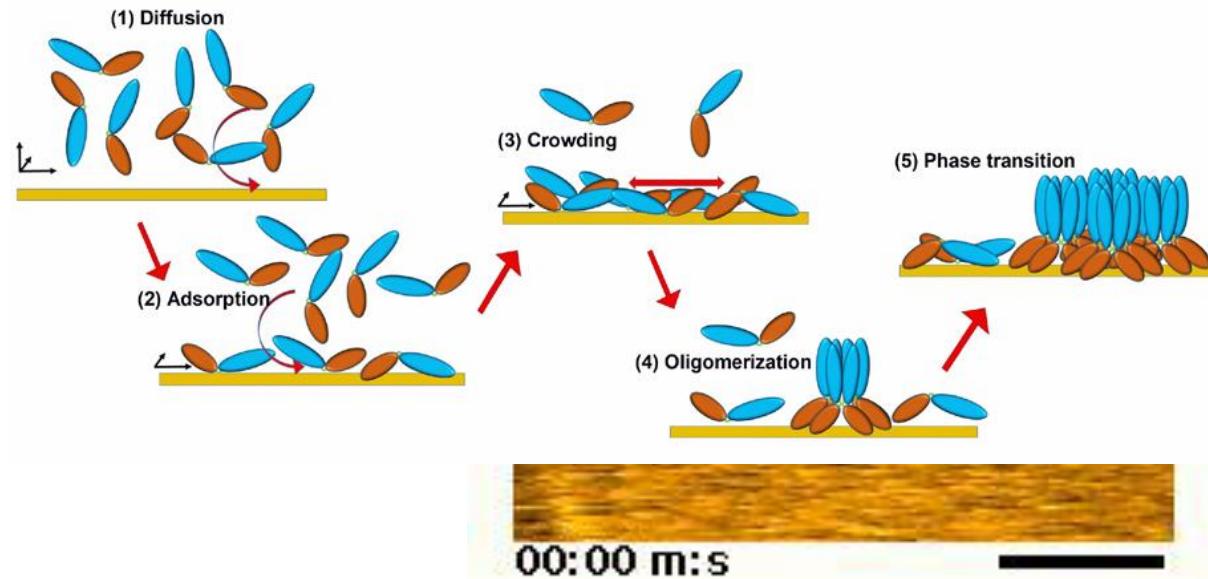
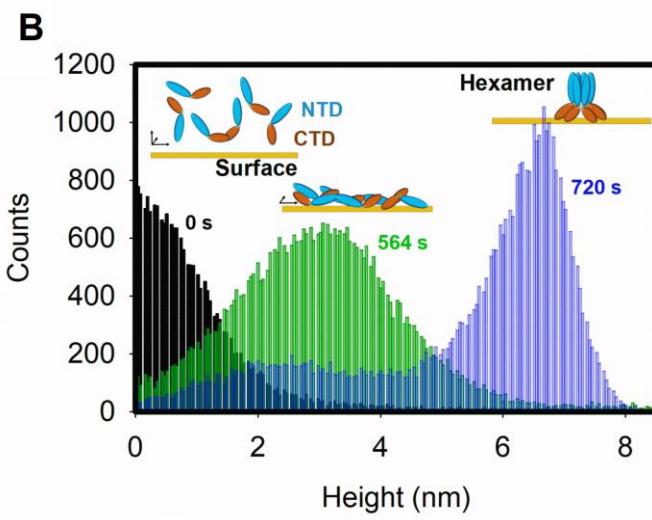
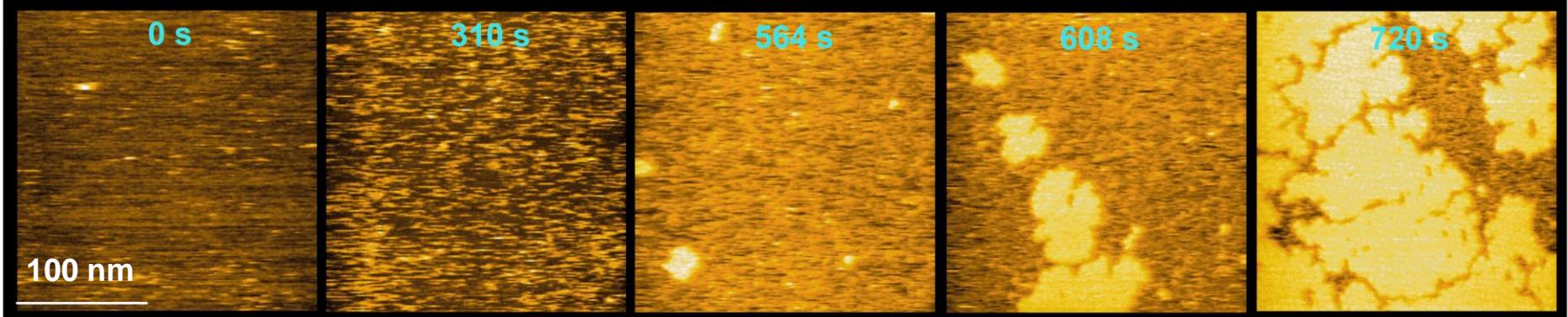


Are we able to follow assembly in real-time?

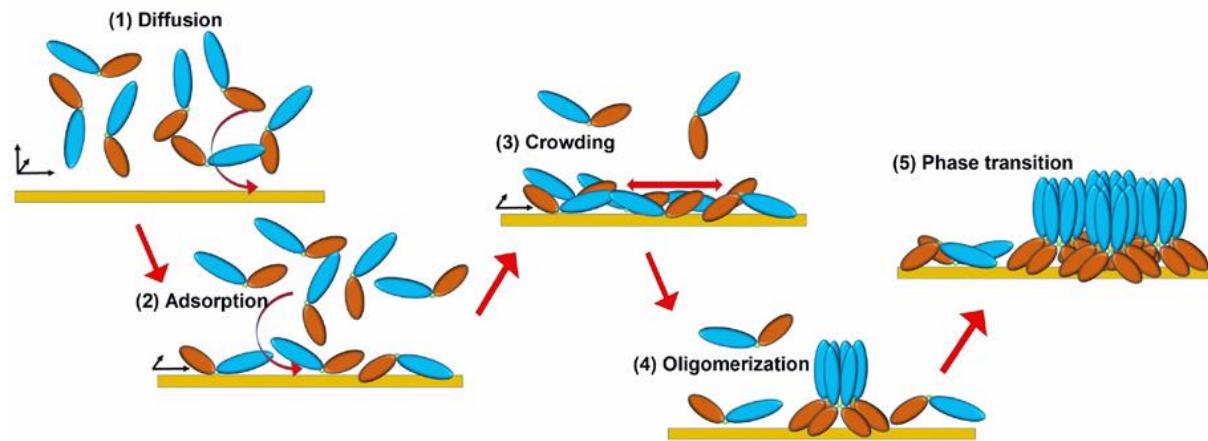
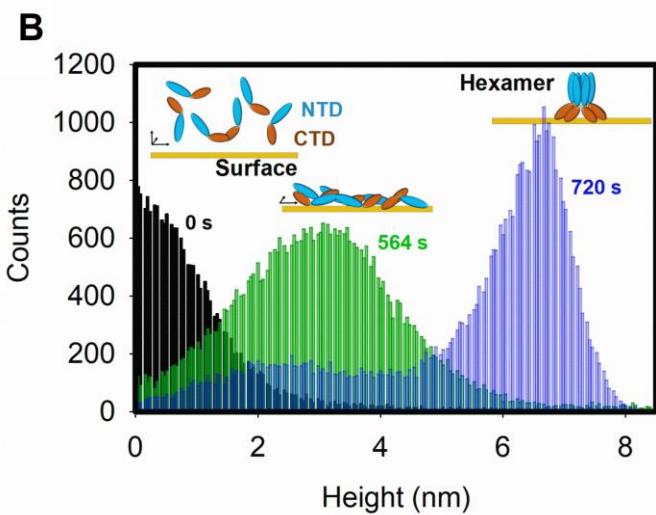
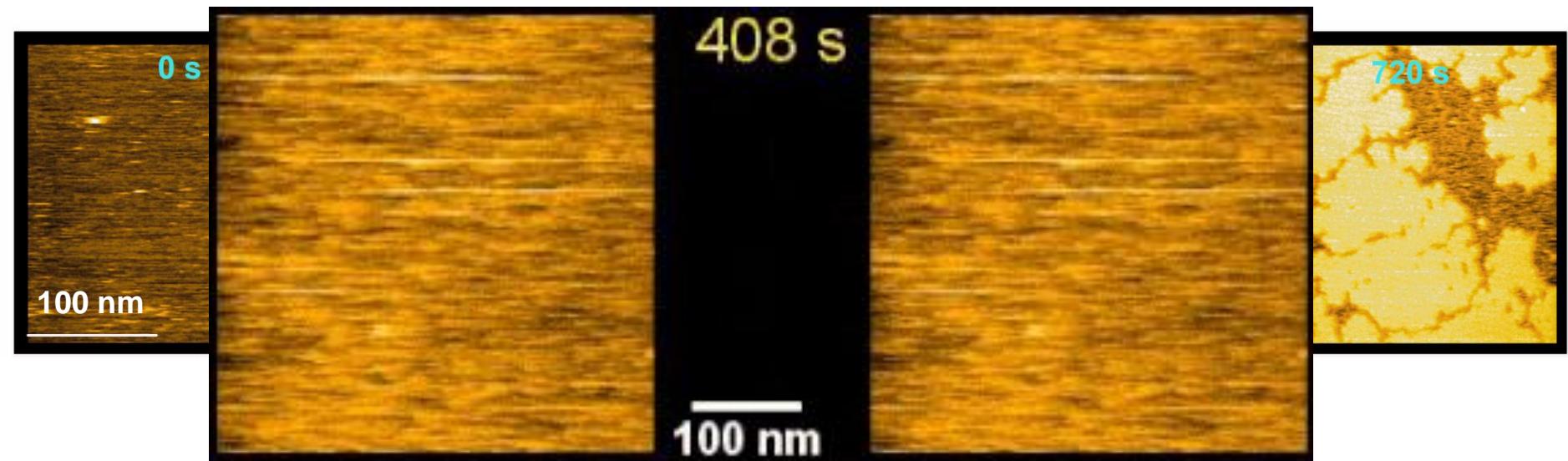
High Speed AFM studies of 2D HIV assembly



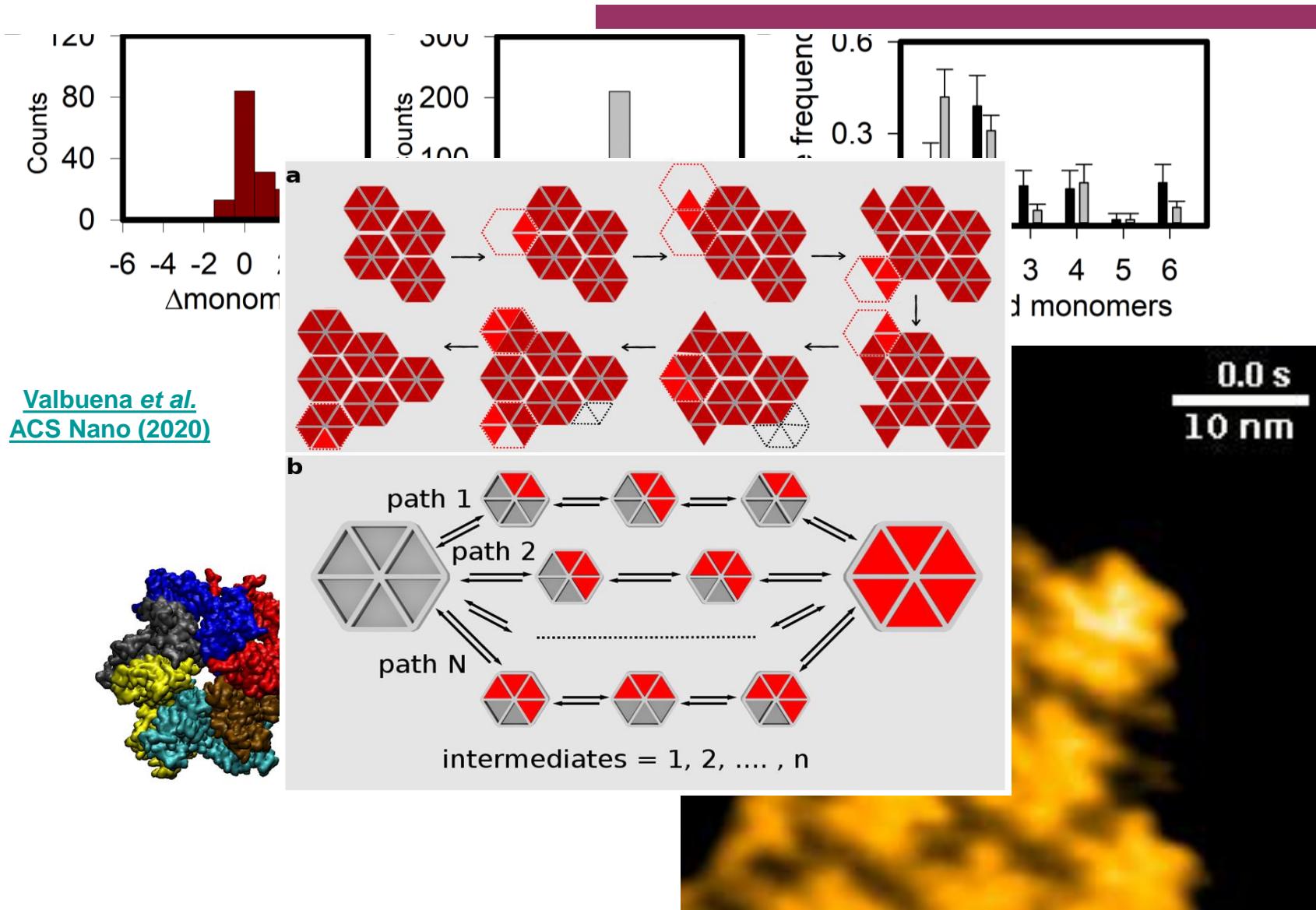
High Speed AFM studies of 2D HIV assembly



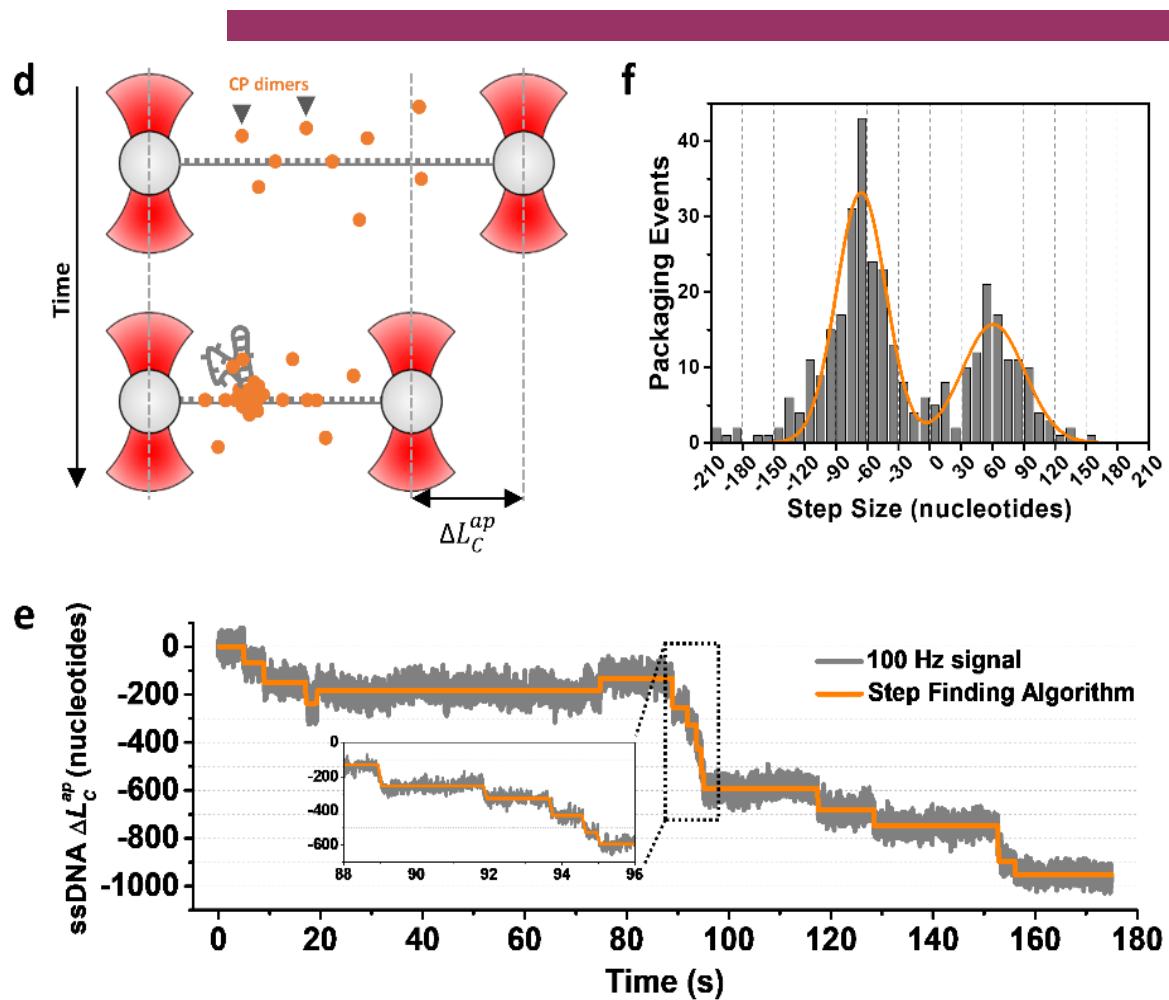
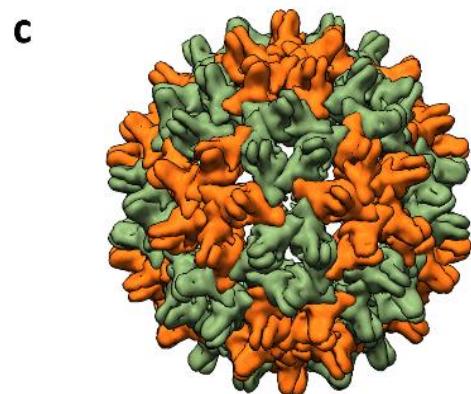
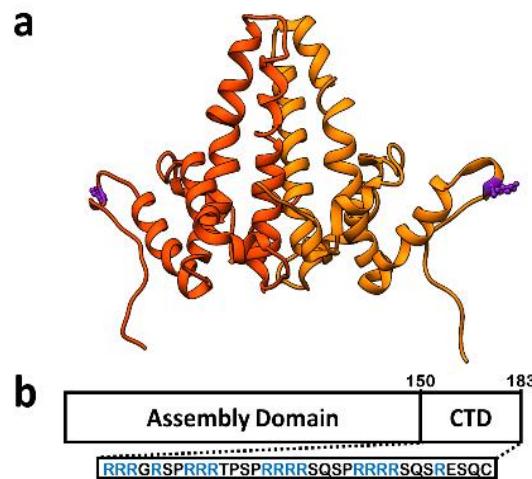
High Speed AFM studies of 2D HIV assembly



High Speed AFM studies of 2D HIV assembly



Hepatitis B Virus assembly



Buzon et al. Science Advances 2021

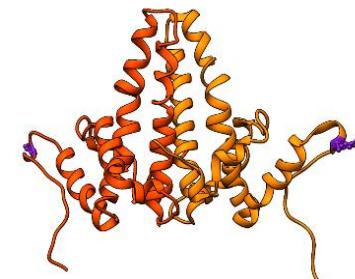
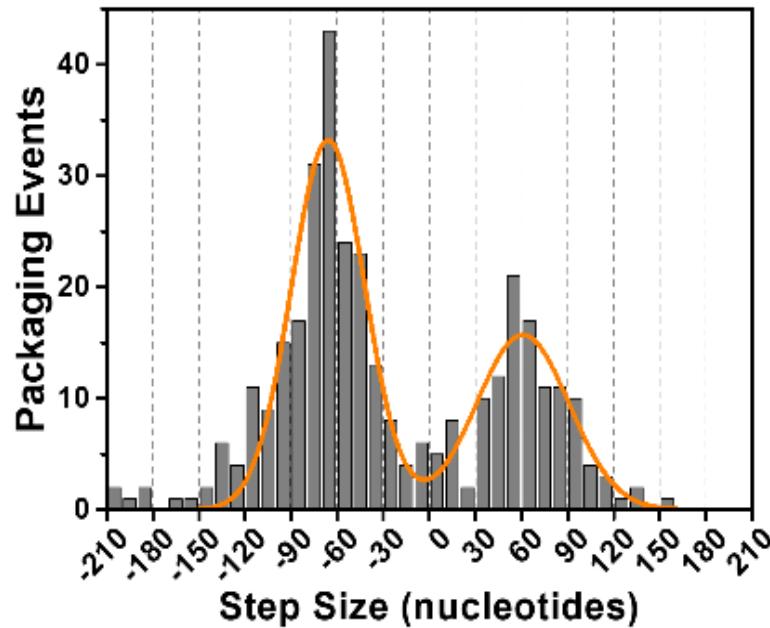
Hepatitis B Virus assembly

Cp condenses nucleic acids,
even under tension ($F = 11$ pN)

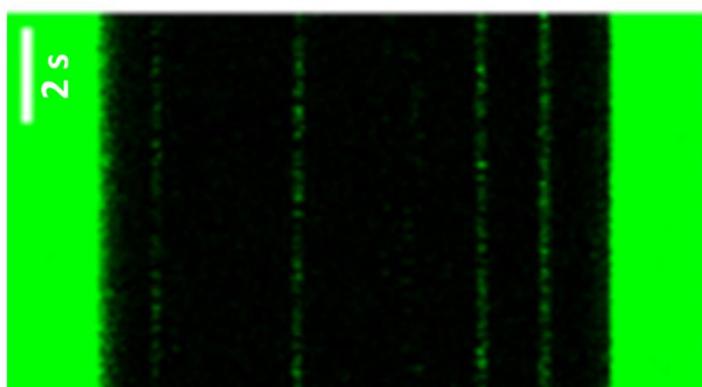
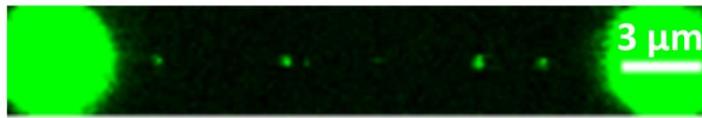
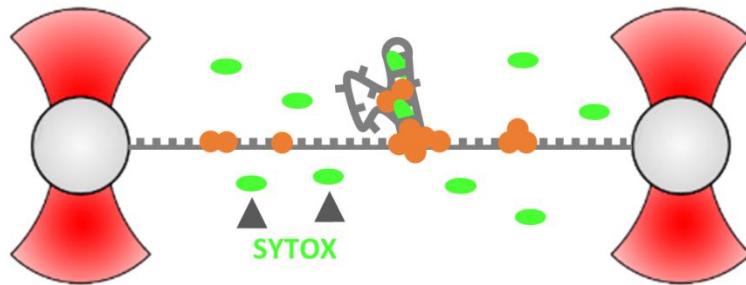
Assembly footprint: ~70 nt

Work per condensation step:
 F^* step size = ~ 100 $k_B T$

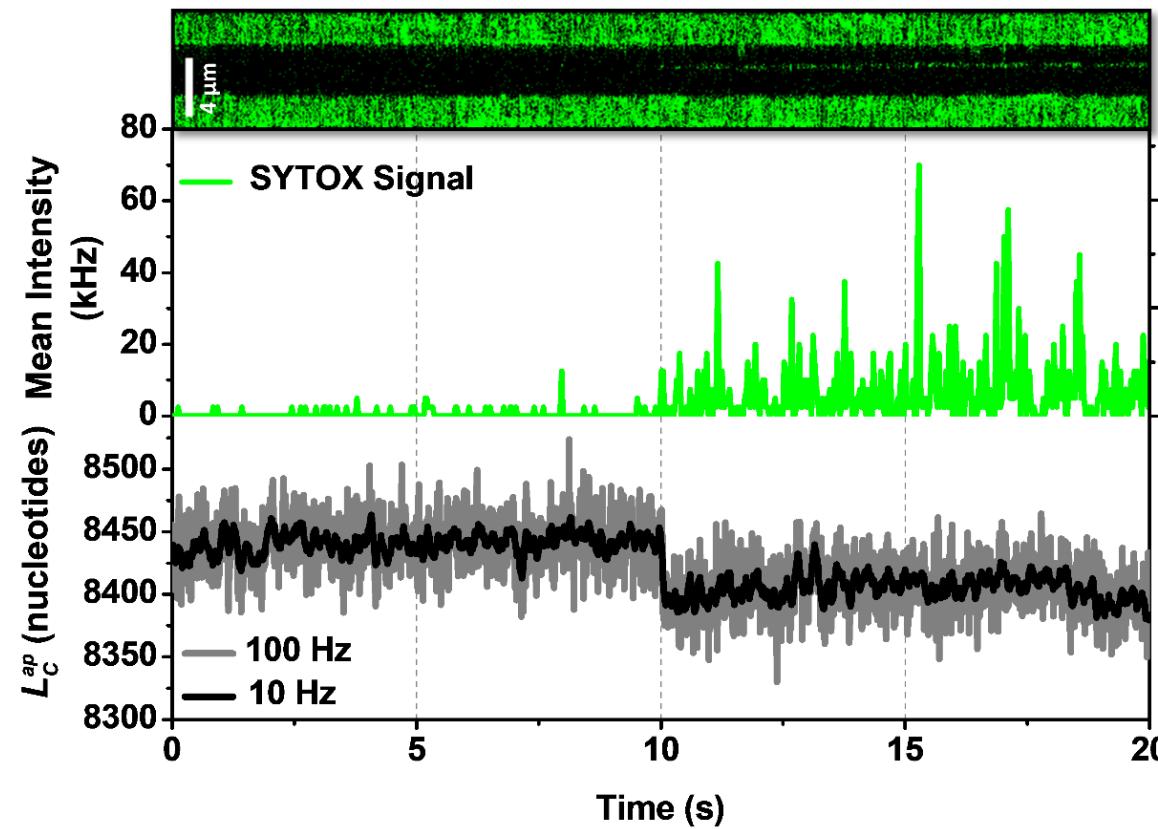
This corresponds to ~1.4 $k_B T/nt$
(compare to ~5 $k_B T/bp$ for ATP driven
packaging motor of $\Phi29$)



Hepatitis B Virus assembly

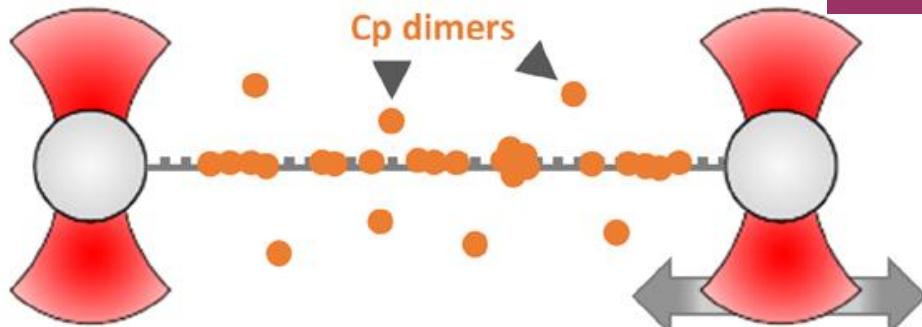


Cp has the capability to chaperone the formation of ds genome patches

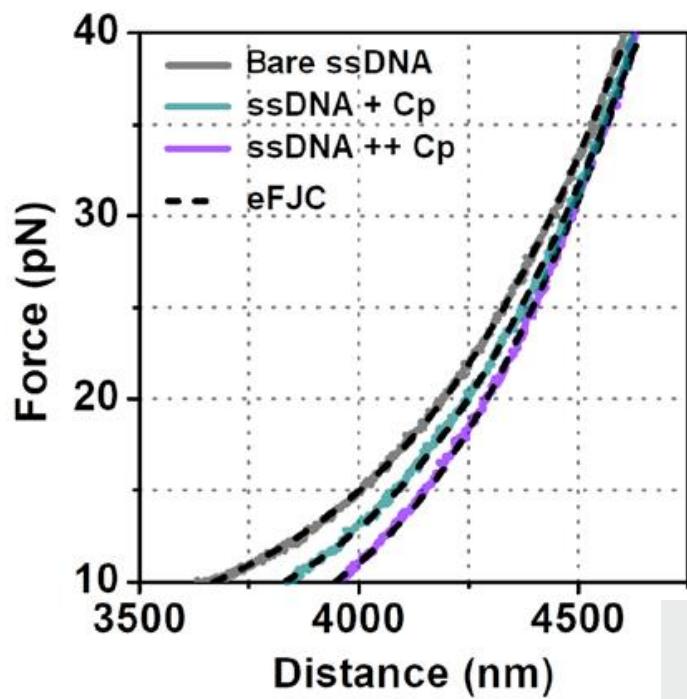


Hepatitis B Virus assembly

A



B



Force-extension curves for varying Cp concentrations

extensible Freely Jointed Chain fitting yields L_p as function of [Cp]

$$\Delta G_{\text{prot-DNA}}^0 (k_B T)$$

$$-10 \pm 2$$

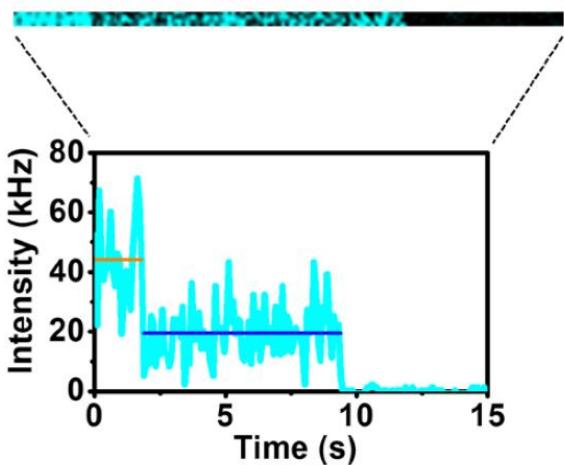
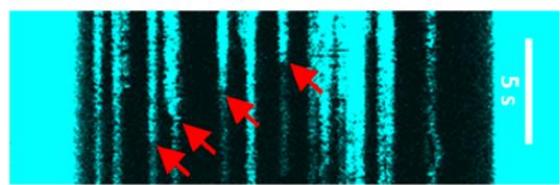
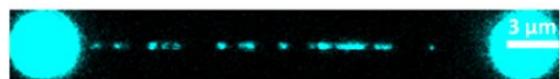
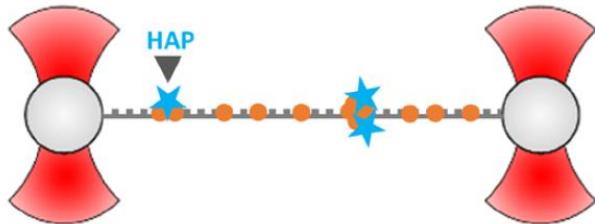
$$\Delta G_{\text{prot-prot}}^0 (k_B T)$$

$$-7 \pm 1$$

$$\Delta G_{\text{bind}}^0 (k_B T)$$

$$-16 \pm 2$$

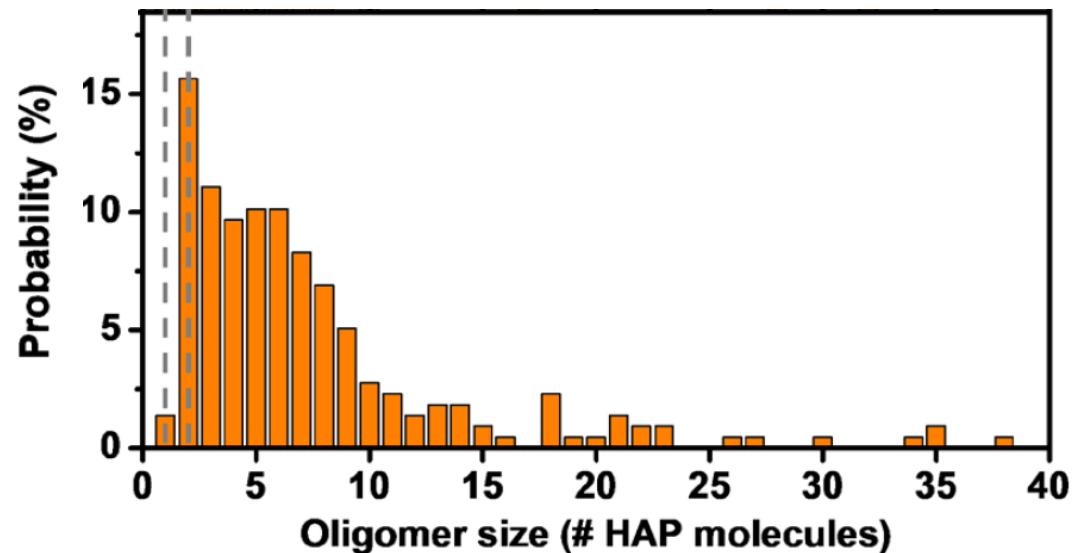
Hepatitis B Virus assembly



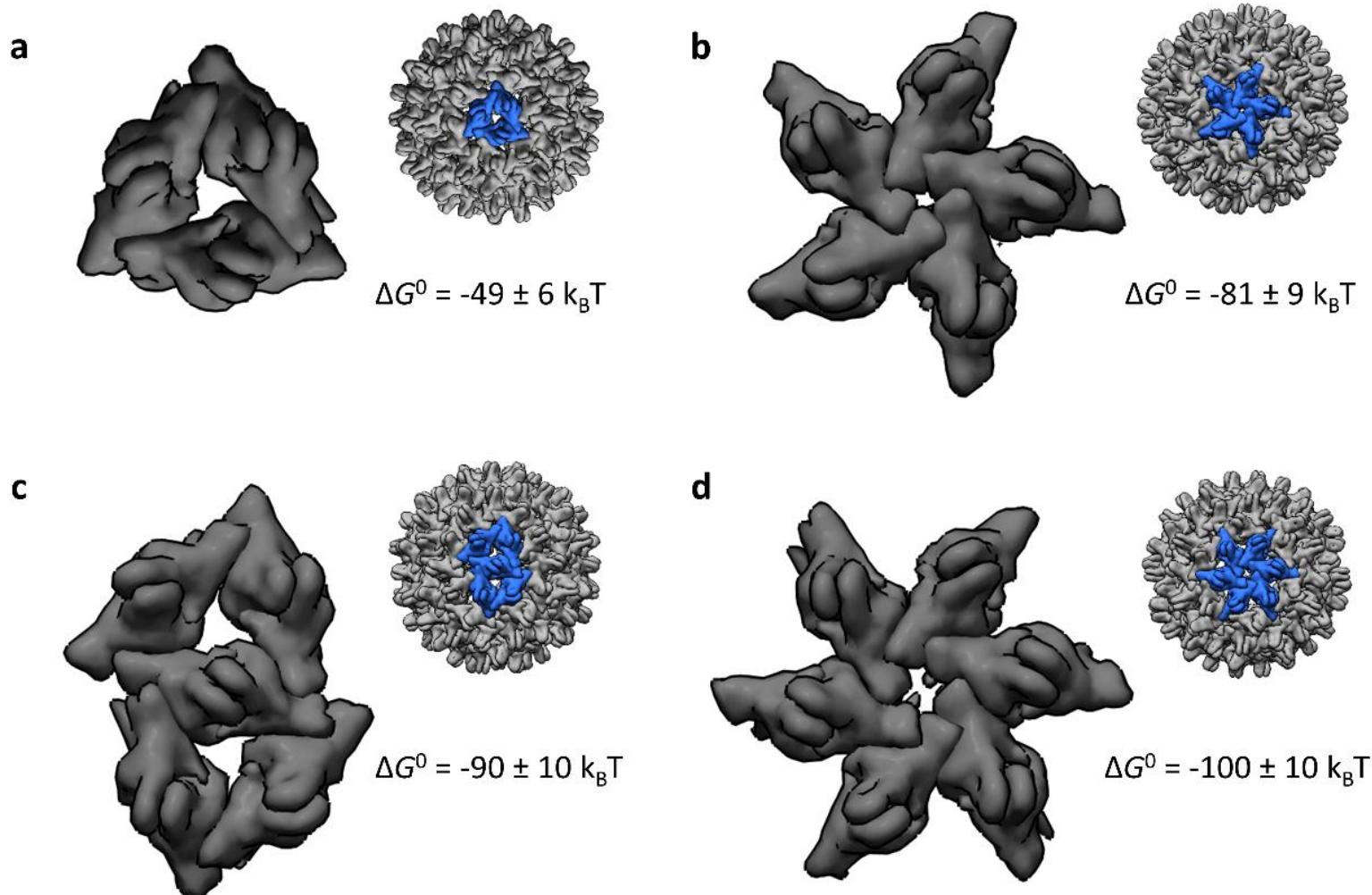
Assembly nucleus?

HAP only binds dimer-dimer interfaces

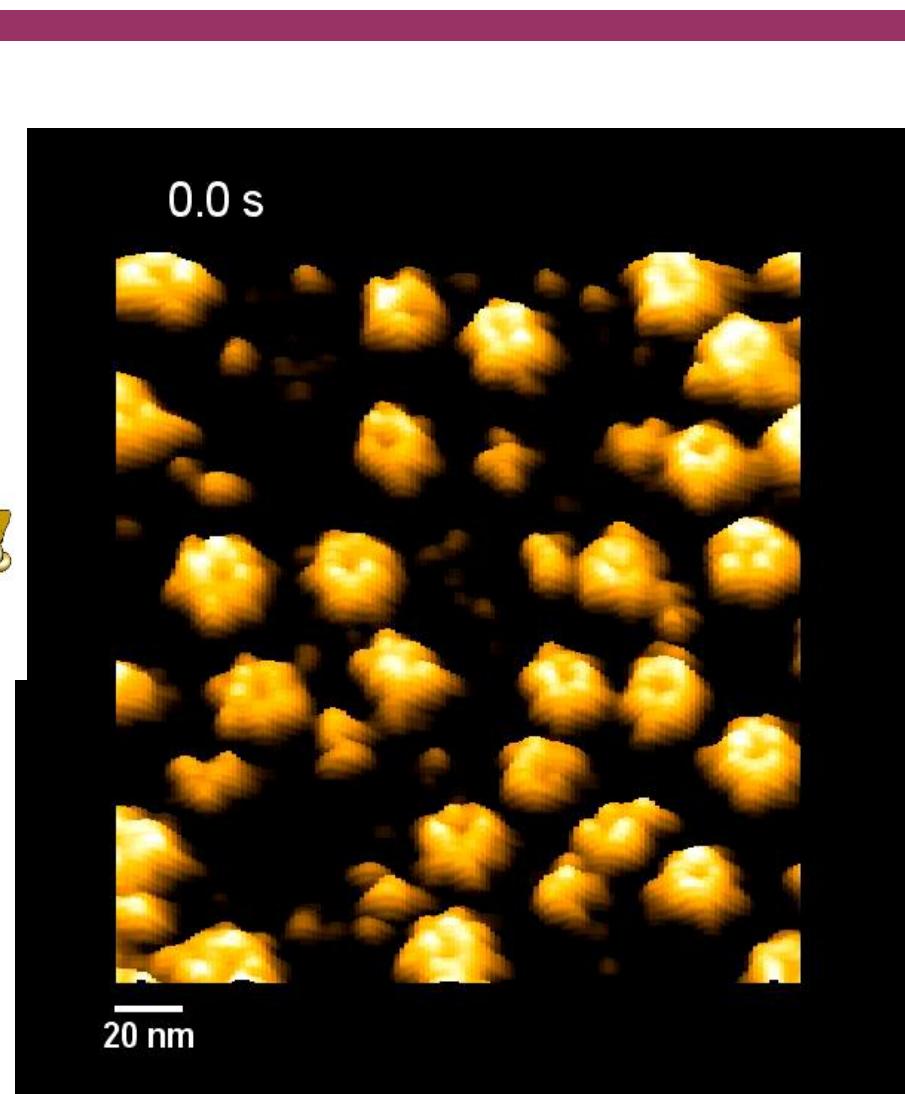
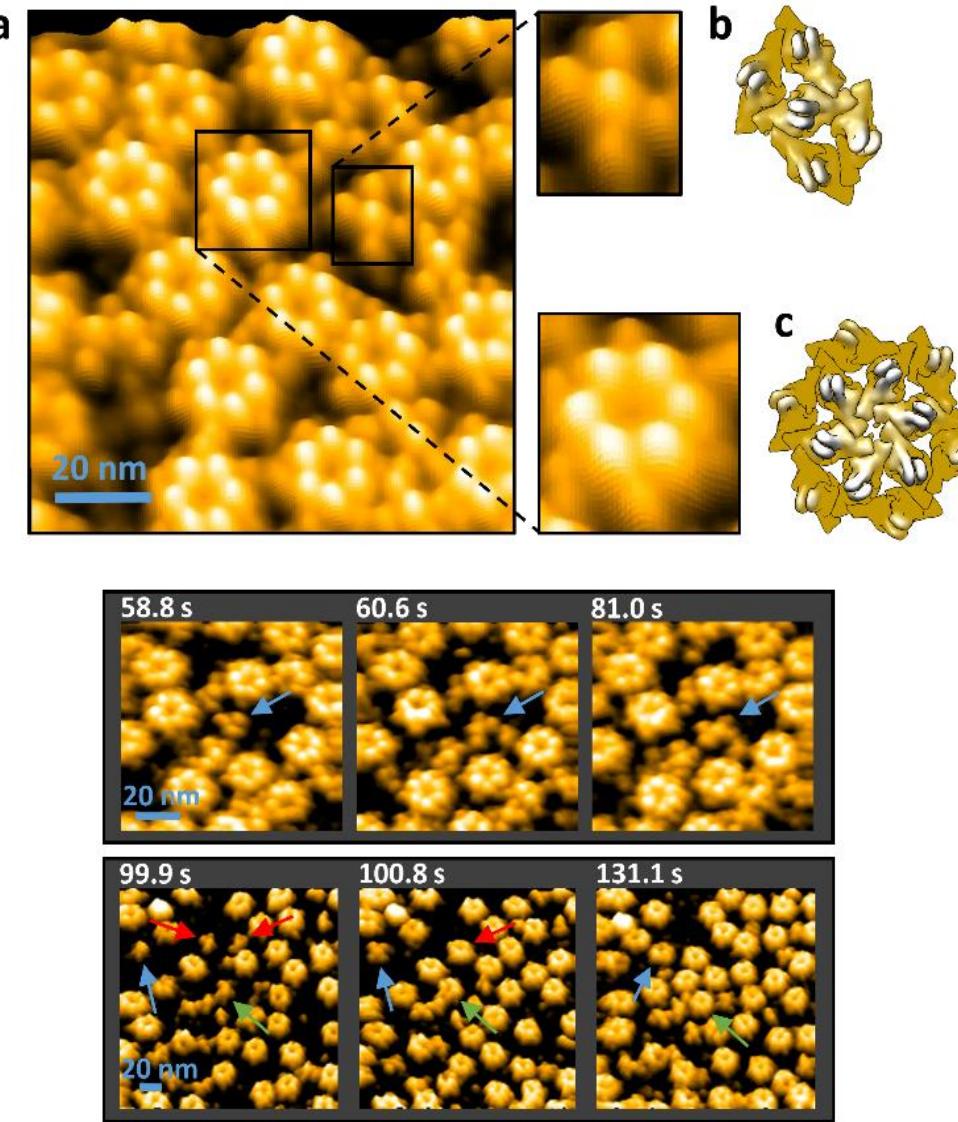
Discrete bleaching steps yields # HAP



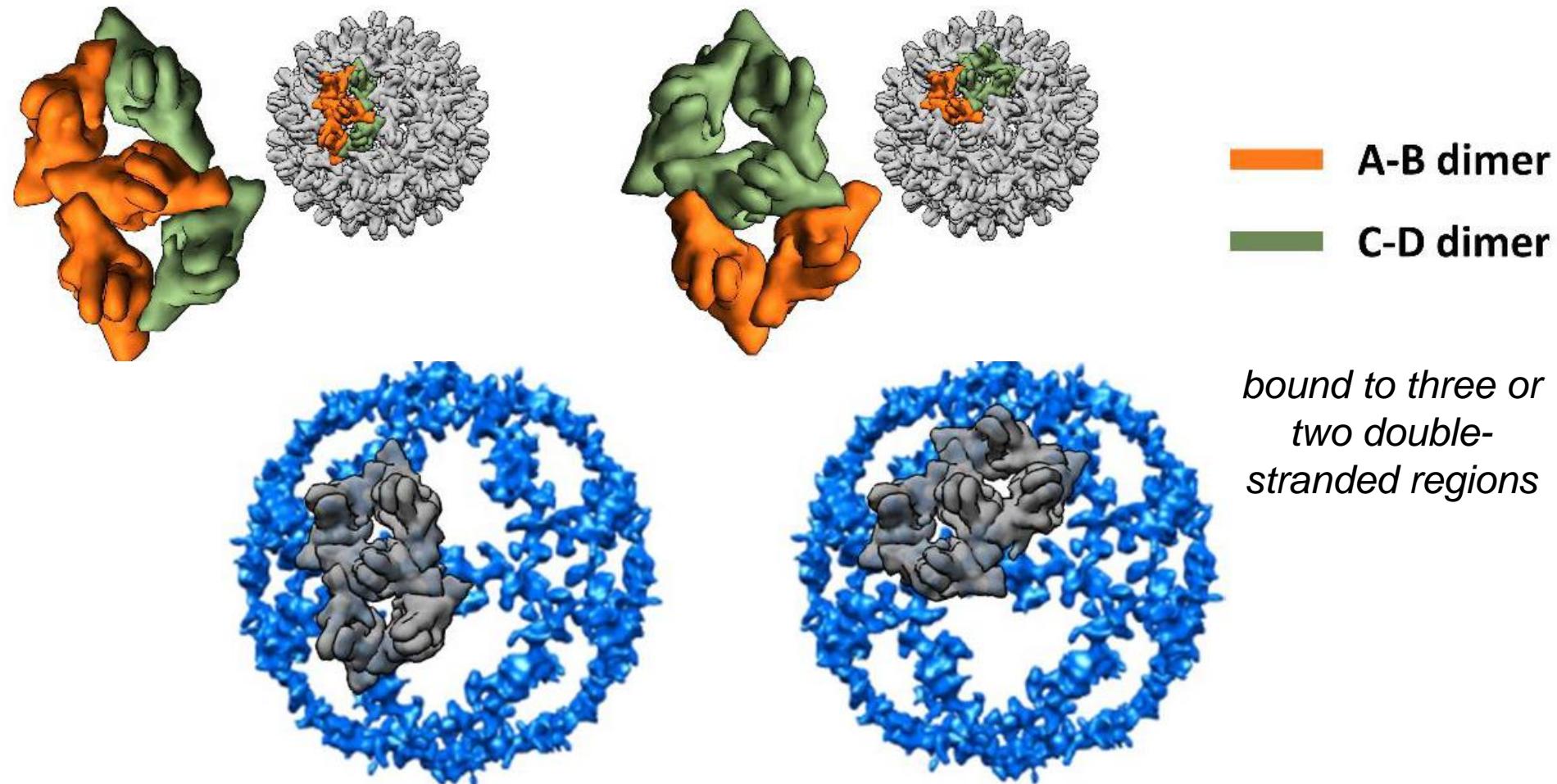
Hepatitis B Virus assembly



Hepatitis B Virus assembly



Hepatitis B Virus assembly



HBV assembly pathway proceeds by the formation of diamond pentamers and dodecamers, instead of through the fivefold symmetric pentamer

Hepatitis B Virus assembly

Index	Structure	Single contacts	Total contacts
-------	-----------	-----------------	----------------

Diamond pentamer represents local minimum of energy as the smallest configuration of dimers that has more interfacial contacts than subunits.

1.1		4.1		4
-----	---	-----	---	---

2.1  1 1

4.2  3 3

Assembly proceeds by reversible reactions, where subunits forming part of contact-rich structures are likely to persist.

3.2  2 2

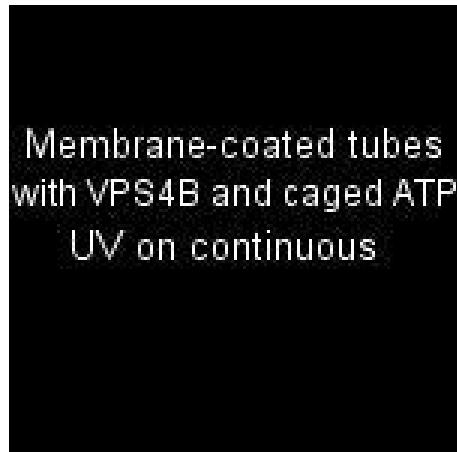
5.1  0 6

Virus self-assembly proceeds through contact-rich energy minima

3.3  5

4.3		5
-----	---	---

For those not only interested in viruses

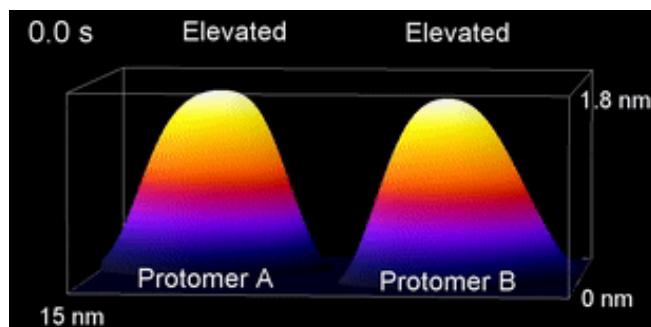


ESCRT:

[Maity et al. Science Advances \(2019\)](#)

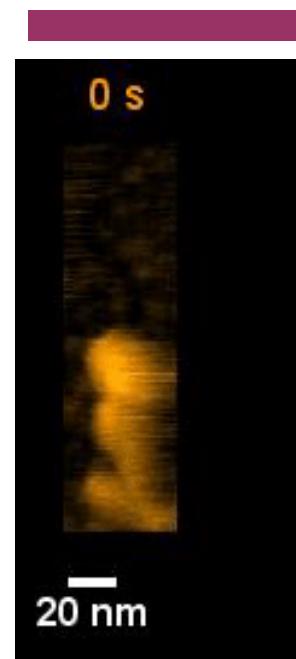
[Bertin et al. Nat. Comm. \(2020\)](#)

[Azad et al. Nat. Struct. Mol. Biol. \(2023\)](#)



Bacterial transporters:

[Maity et al. PNAS \(2022\)](#)



Assembly of synthetic systems:

[Maity et al. JACS \(2020\)](#)

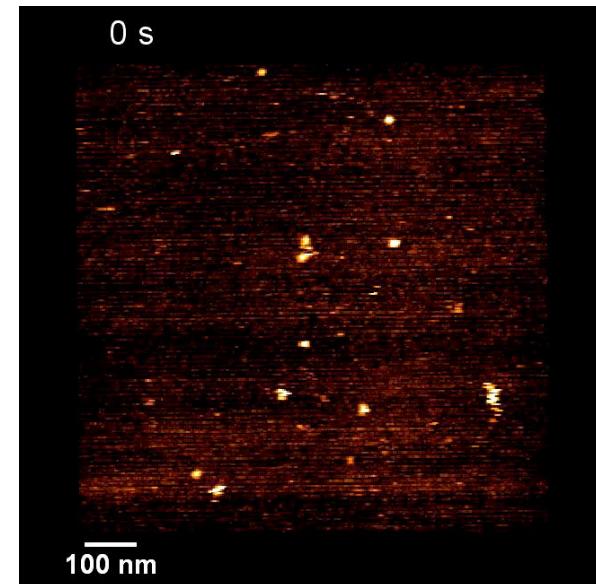
[Liu et al. Nat. Chem. \(2023\)](#)

Assembly of antibiotics:

[Shukla et al. Nature \(2022\)](#)

[Melcrova et al. Nat. Comm. \(2023\)](#)

[Shukla et al. Cell \(2023\)](#)



High Speed AFM studies of dynamics

Antimicrobial peptides in action

January 20, 2022

Articles

THE LANCET

Global burden of bacterial antimicrobial resistance in 2019: a systematic analysis

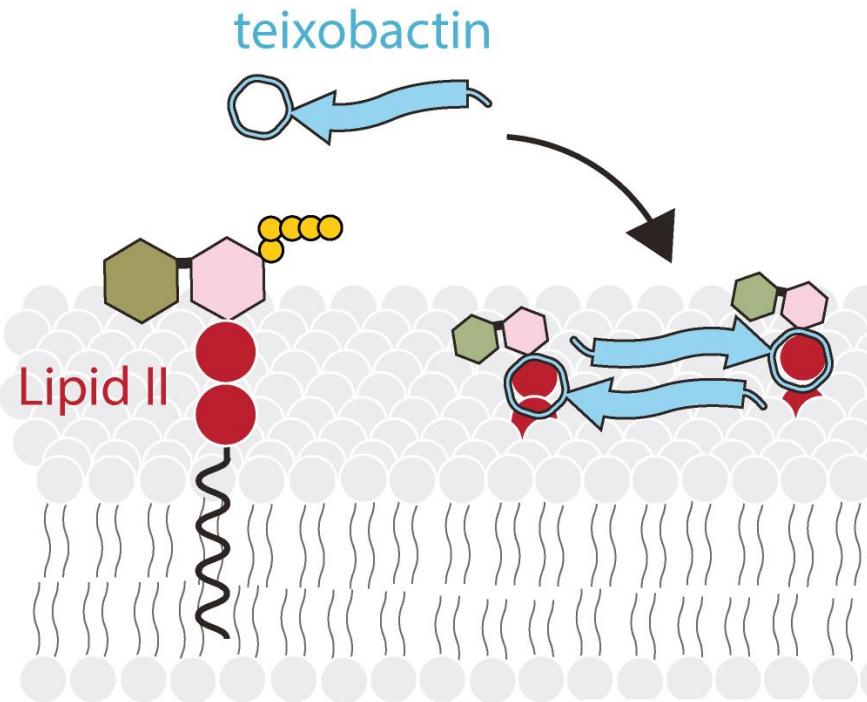
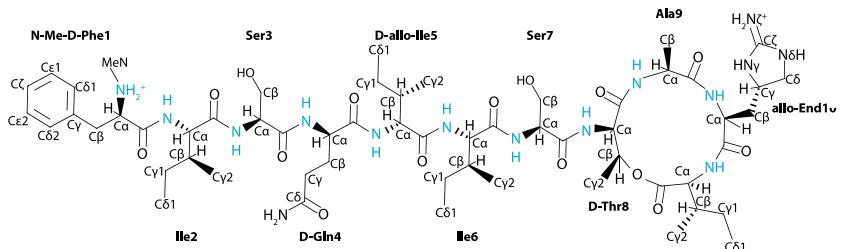
Antimicrobial Resistance Collaborators*



- *First comprehensive analysis of global impact of antimicrobial resistance (AMR) estimates resistance itself caused 1.27 million deaths in 2019, and that antimicrobial-resistant infections played a role in 4.95 million deaths*

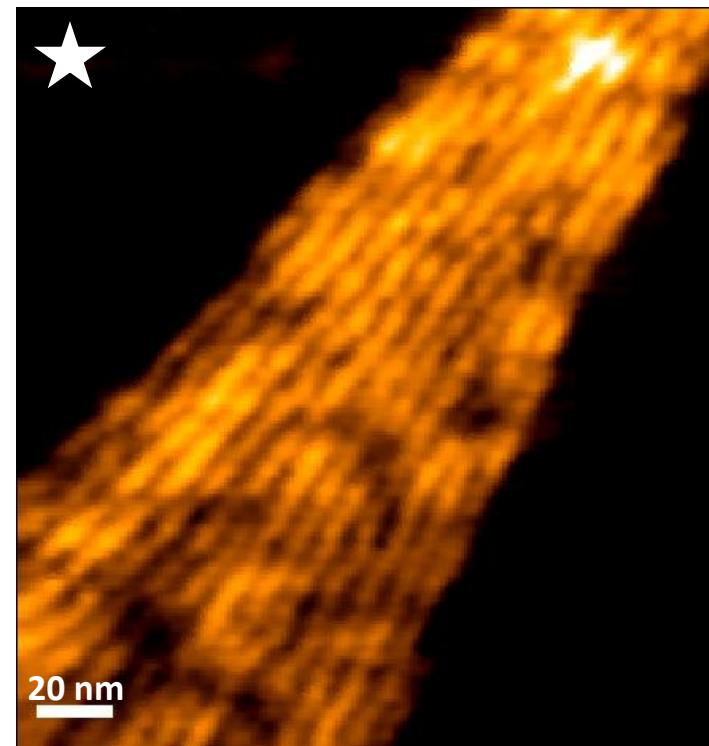
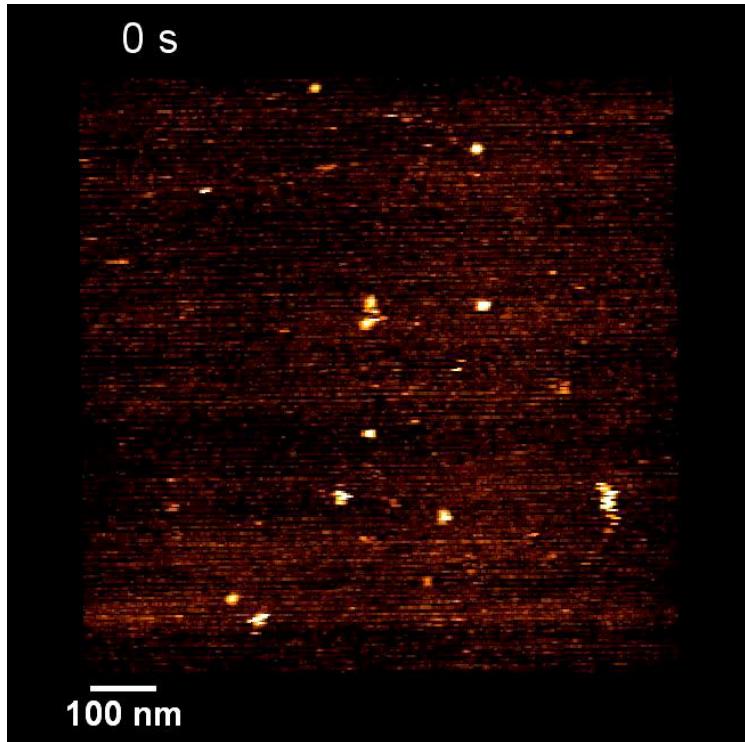
Molecular Mechanism of
Antimicrobials...

Antimicrobial peptides in action: Teixobactin

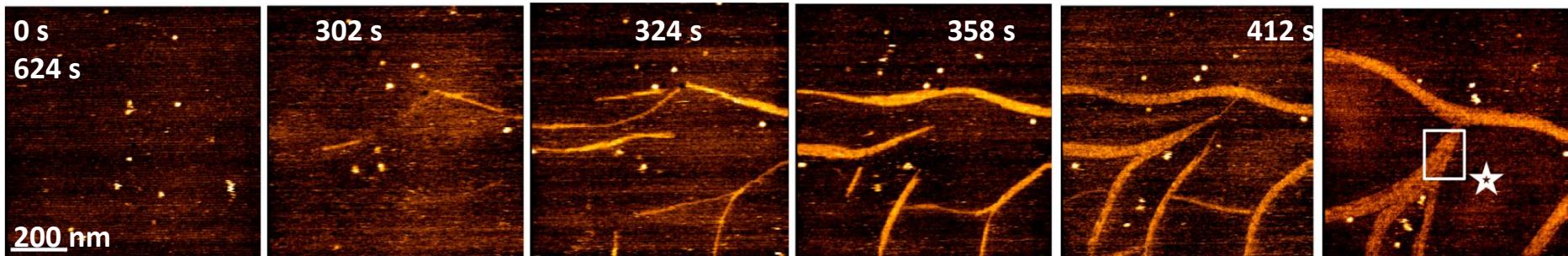


- Active against methicillin-resistant *Staphylococcus aureus* and *Mycobacterium tuberculosis*.
- Teixobactin targets Lipid II (precursors for peptidoglycan synthesis).

Antimicrobial peptides in action: Teixobactin

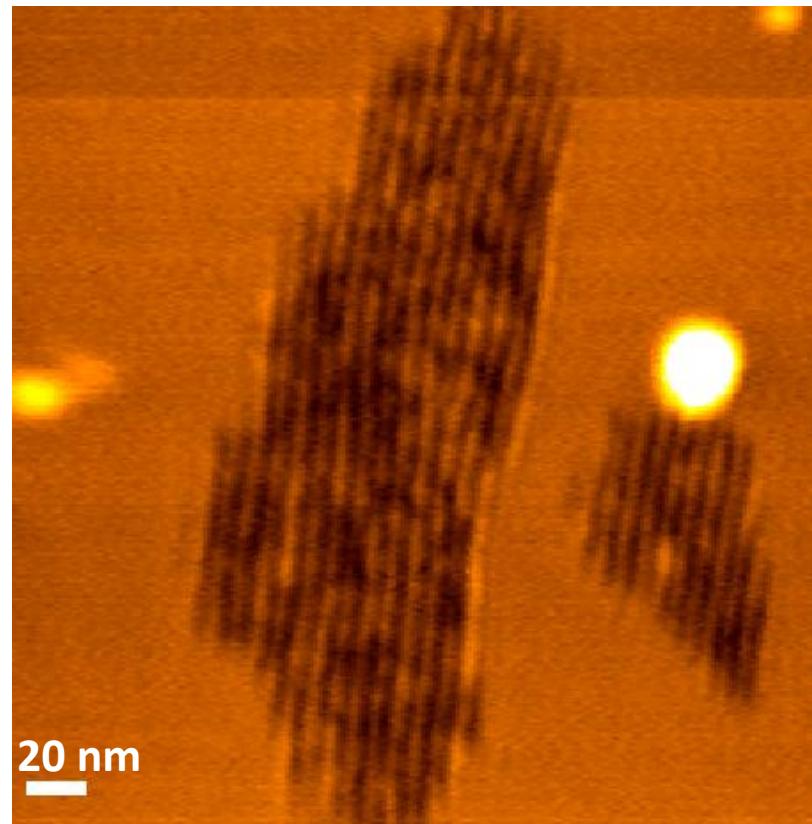
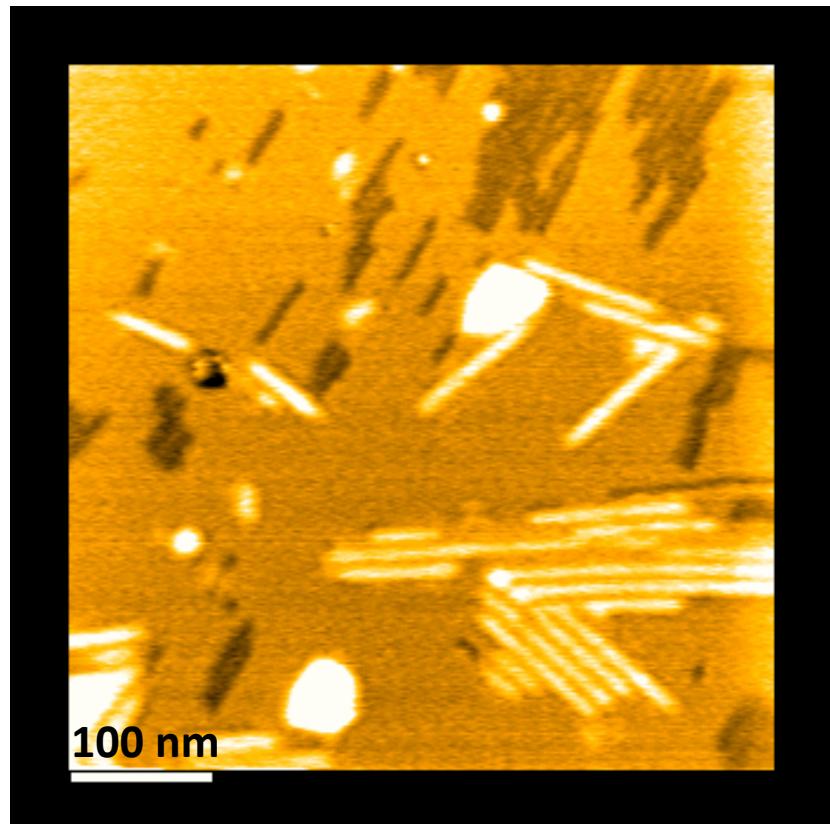


Shukla et al. Nature (2022)

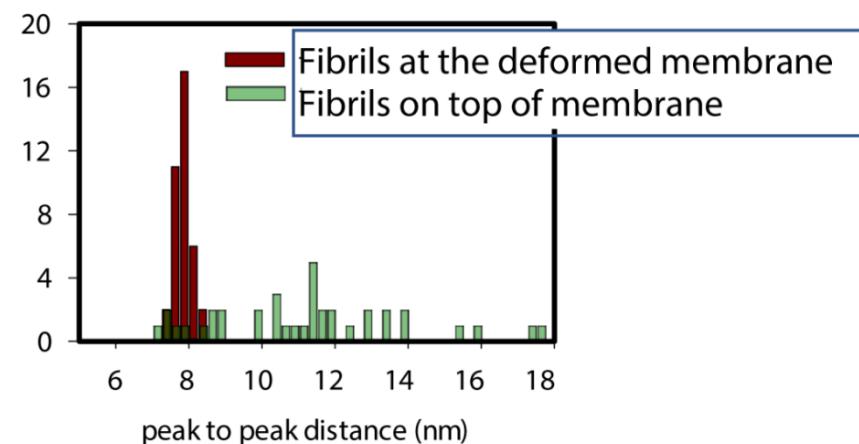
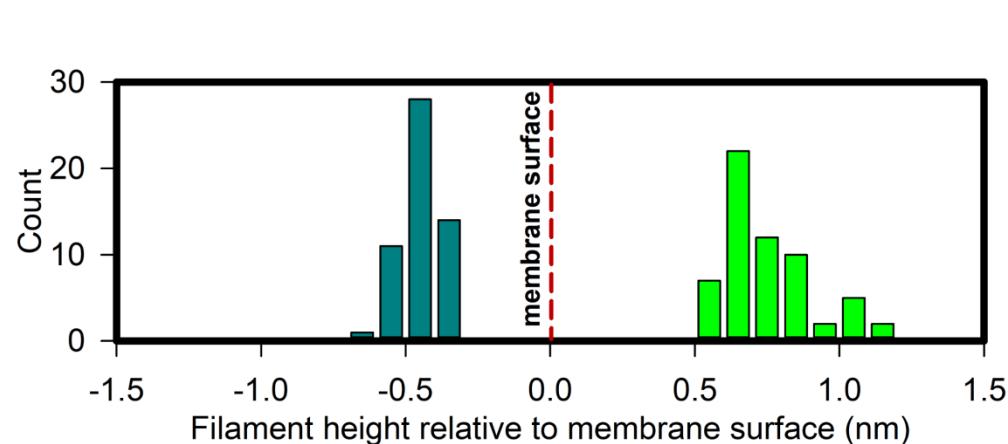
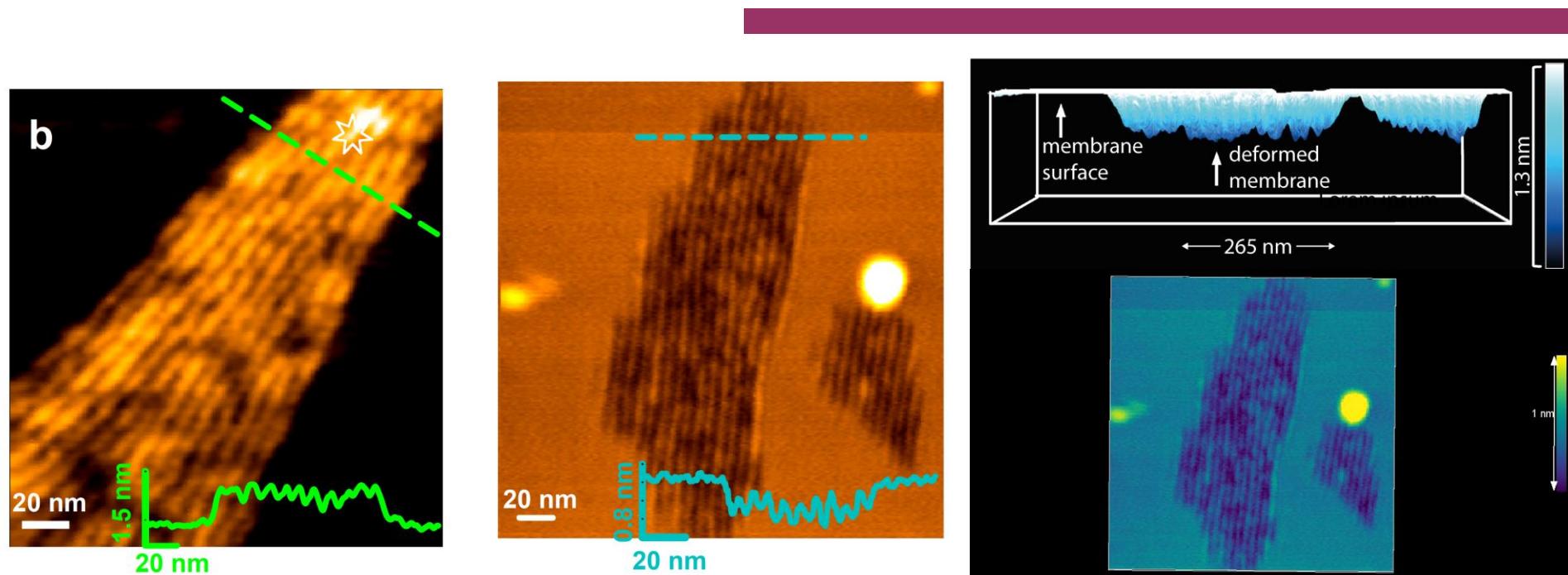


Antimicrobial peptides in action: Teixobactin

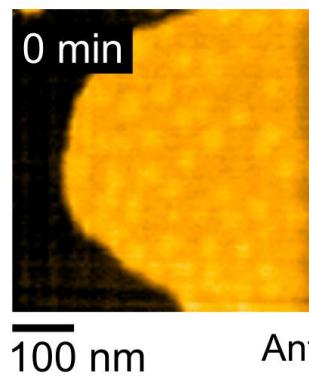
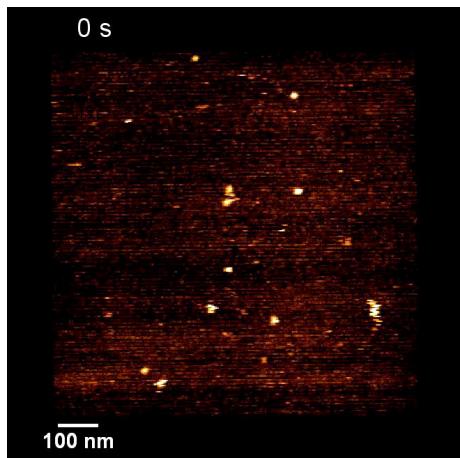
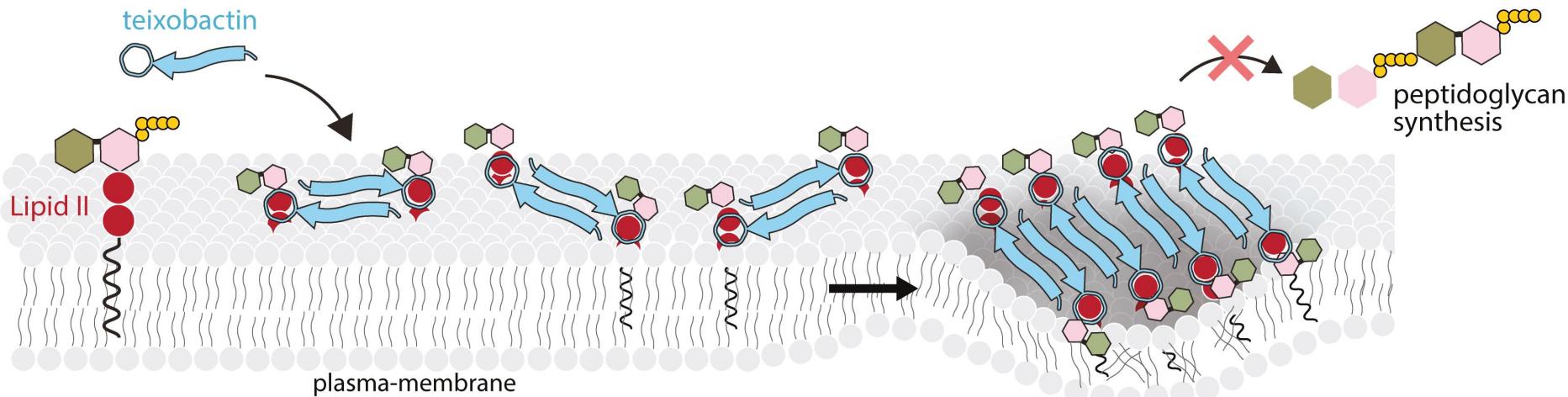
After 30 minutes....



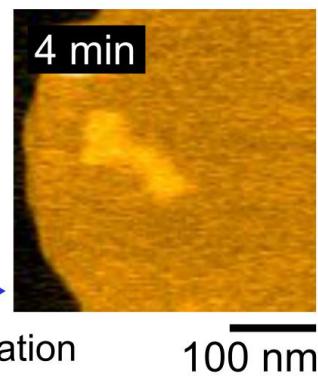
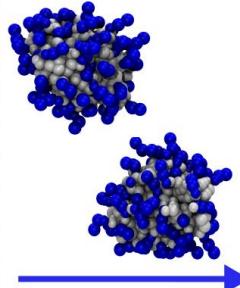
Antimicrobial peptides in action: Teixobactin



Antimicrobial peptides in action



AMC-109



100 nm

Antibiotic incorporation
into bacterial membrane

100 nm

Teixobactin: Shukla et al. *Nature* (2022)
Clovibactin: Shukla et al. *Cell* (2023)

Melcrova et al. *Nature Communications* (2023)

Acknowledgments



Roos Lab

G. v/d Borg, S. Maity, Y. Feng, A. Melcrova, C. Richards, Y. Gong, M. Middelkamp,
C. v. Ewijk, S. Sasidharan, S. de Weerd, L. Presutti, Y. Knelissen



R. Sorkin, D. Denning, M. Baclayon, J. Snijder, R. Lira
D. Vorselen, M. v Rosmalen, M. Marchetti, P. Buzón, M. Piontek



Collaborators

S. Otto, S. Marrink, D. Slotboom (**RUG**)
G. Wuite, F. MacKintosh (**VU A'dam**)
R. Schiffelers, A. Heck,
M. Weingarth (**U Utrecht**)
M. Mateu (**CSIC**), G. Nemerow (**Scripps**)
A. Zlotnick (**IU Bloomington**)
N. Kodera, K. Ngo, T. Ando (**Kanazawa**)
C. Utrecht (**CSSB**), R. de Vries (**WUR**)
A. Garcia (**UNAM**), P. Bassereau (**Curie**)
P. vd Schoot (**TUe**), W. Weissenhorn (**IBS**)
M. Schelhaas (**U Münster**)

