

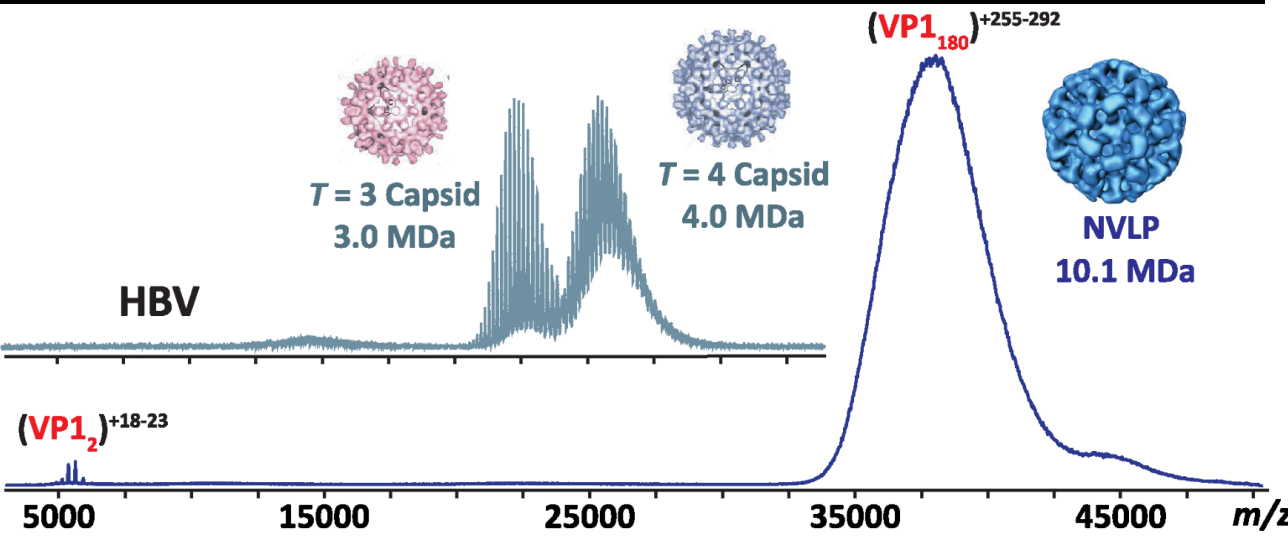
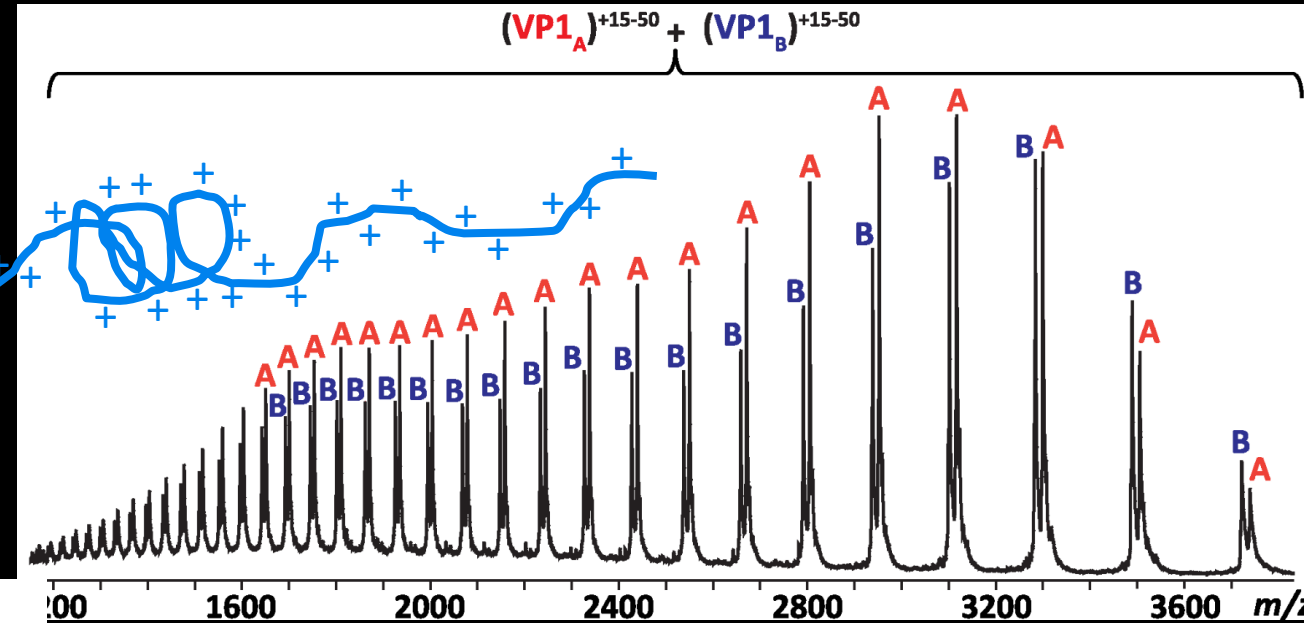
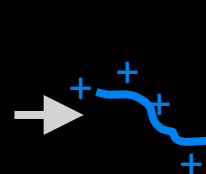
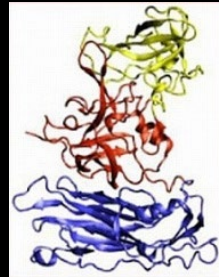
Flying viruses – mass spectrometry meets X-rays

Charlotte Uetrecht



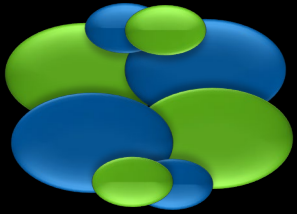
Native MS – up to viruses

- Denatured VP1



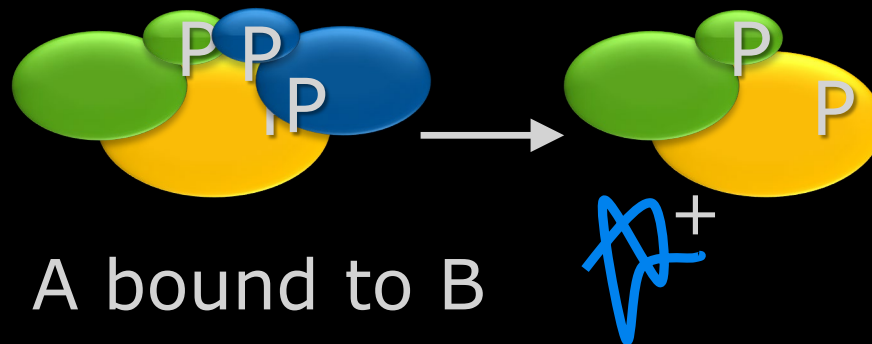
- Buffered solution
→ Intact VLP

Stoichiometry

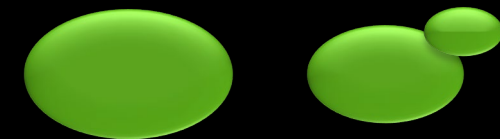


$$M = xA + yB$$

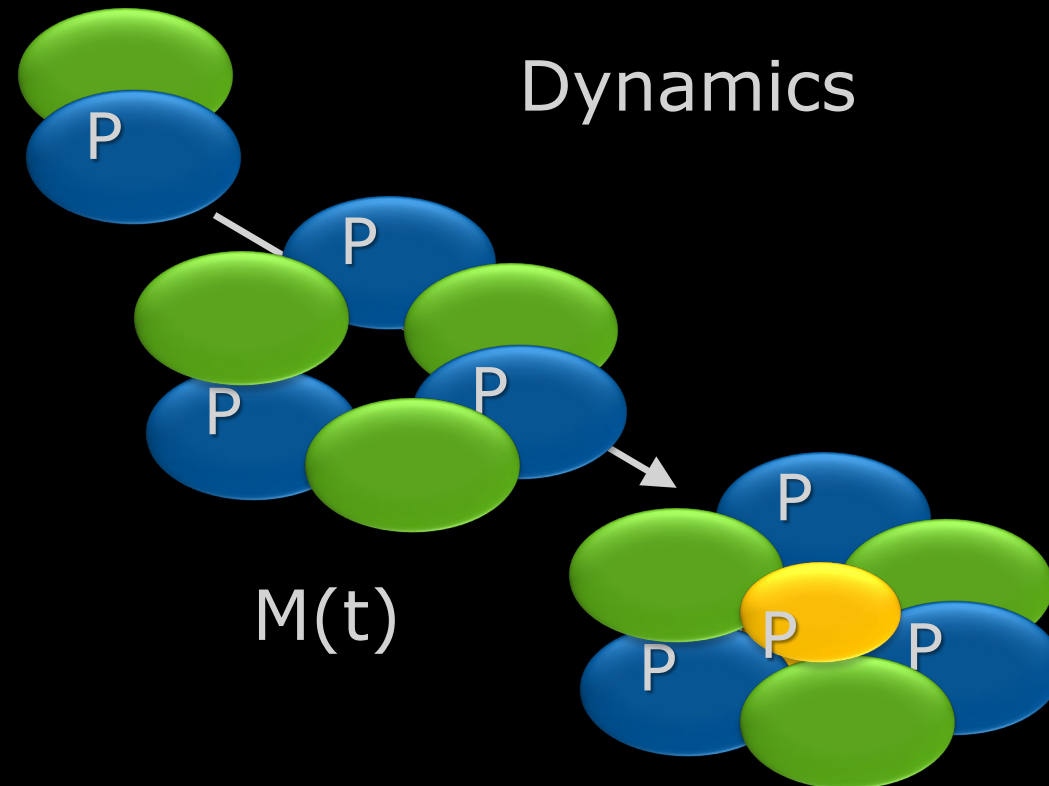
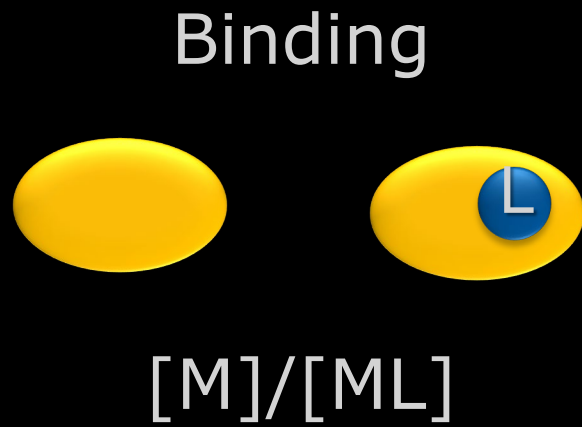
Topology



Shape/ Conformational change

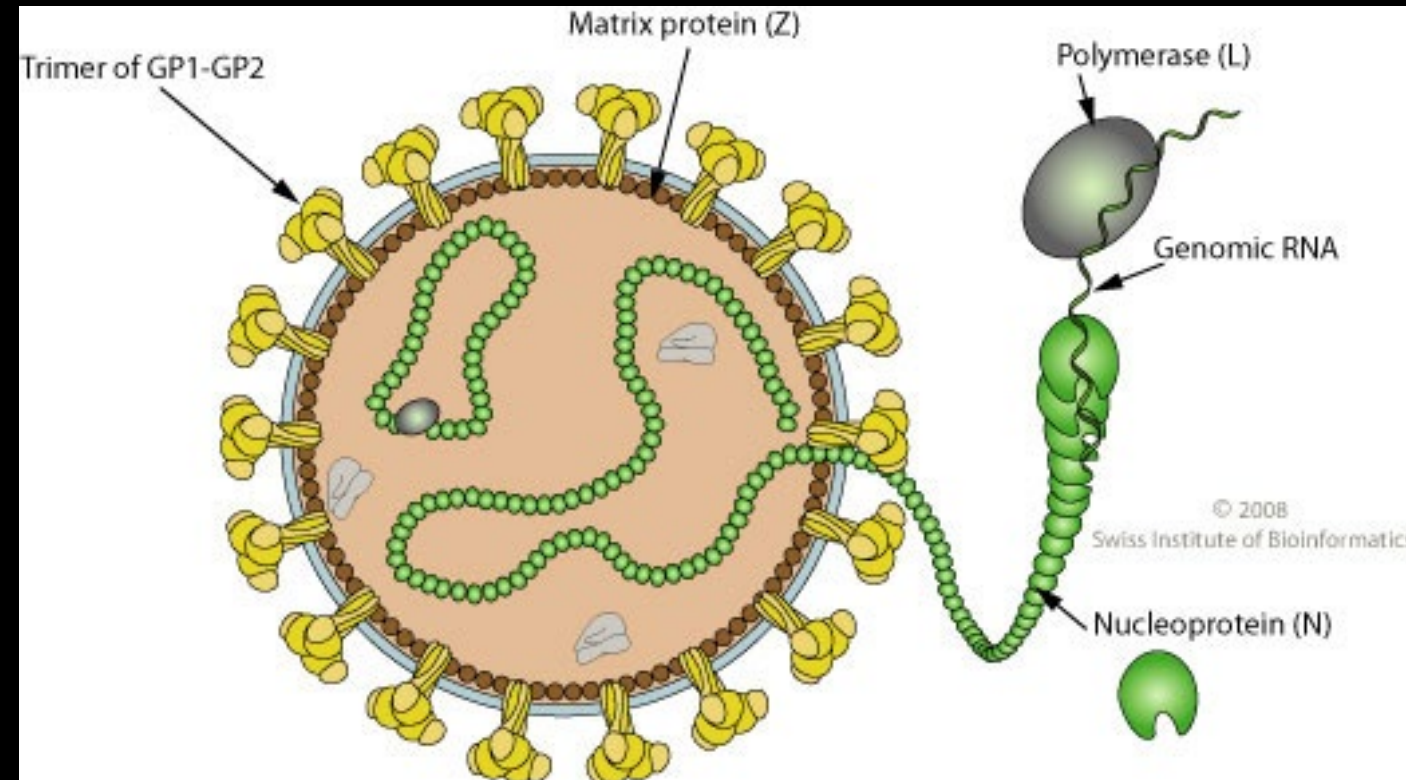


Amount A or B

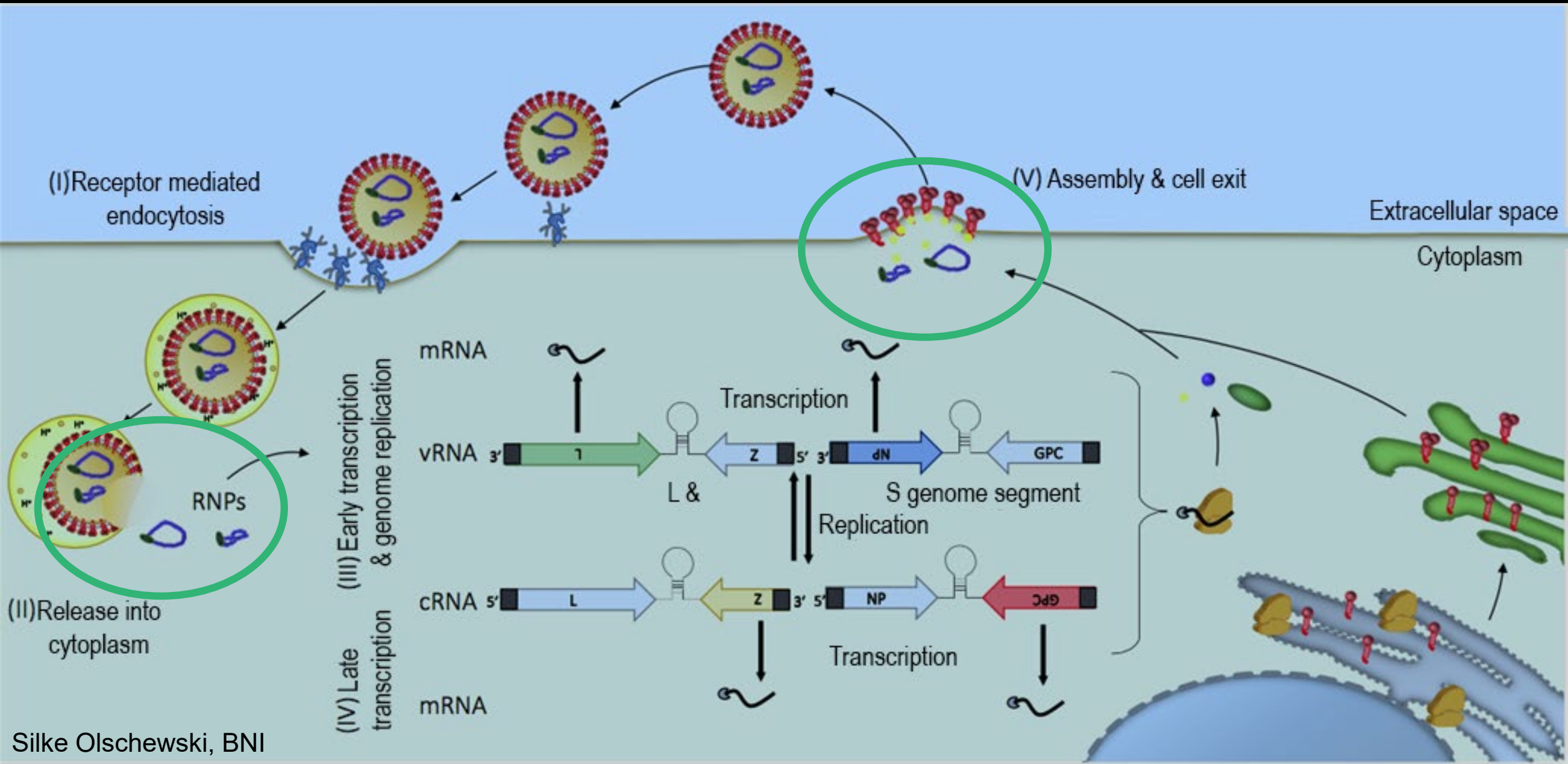


Lassa Virus

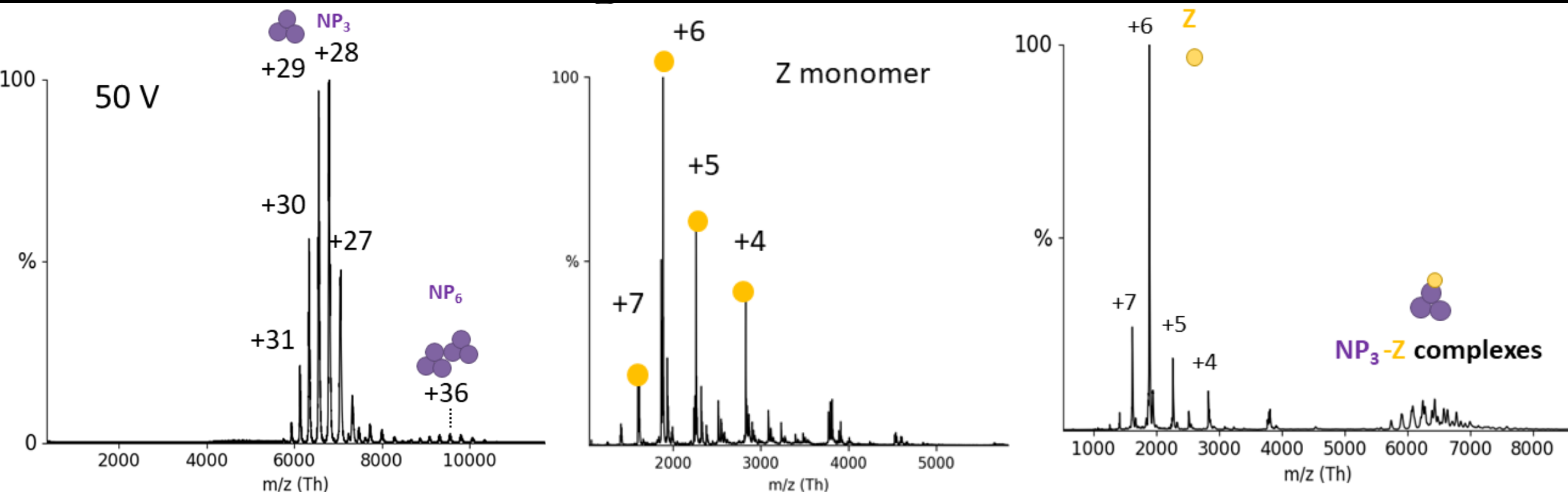
- *Bunyavirales, Arenaviridae*
- Endemic in West Africa,
reservoir: rodents
- Lassa hemorrhagic fever in
humans (~5000 fatalities / a)
- - ssRNA, L & S segment
- 4 proteins
 - (pre)GP
 - L (253 kDa)
 - NP (63 kDa)
 - Z (11 kDa)



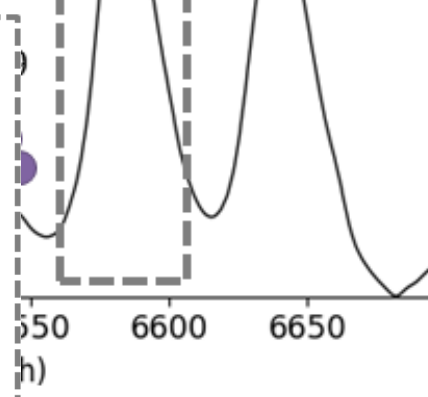
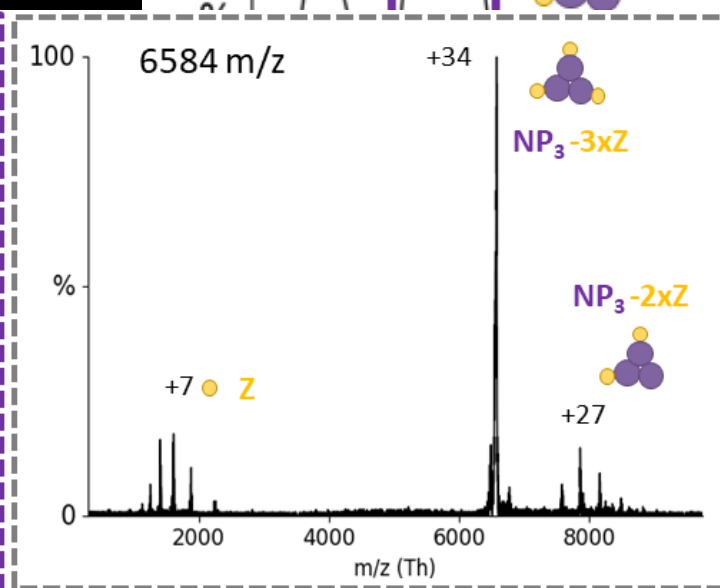
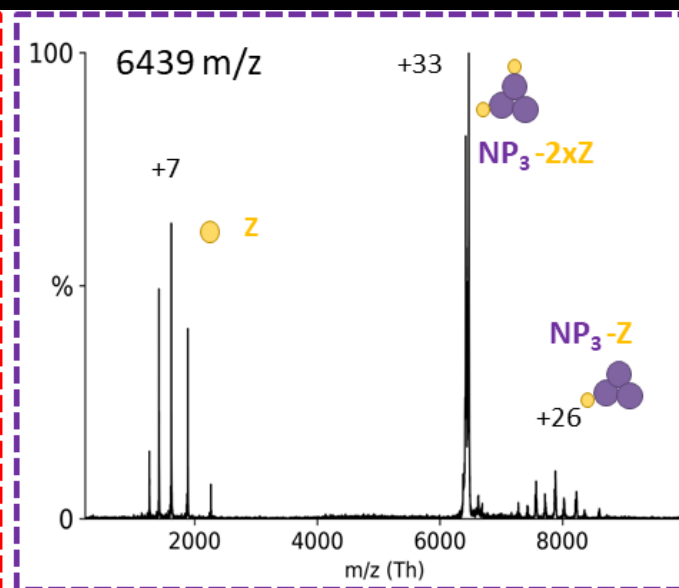
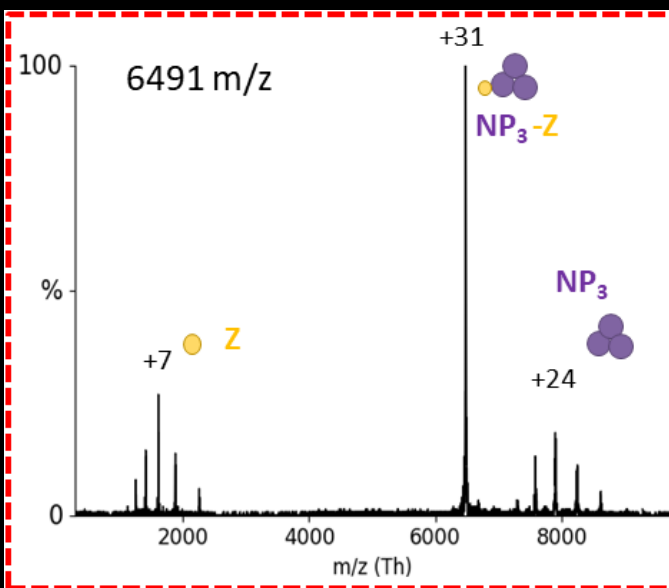
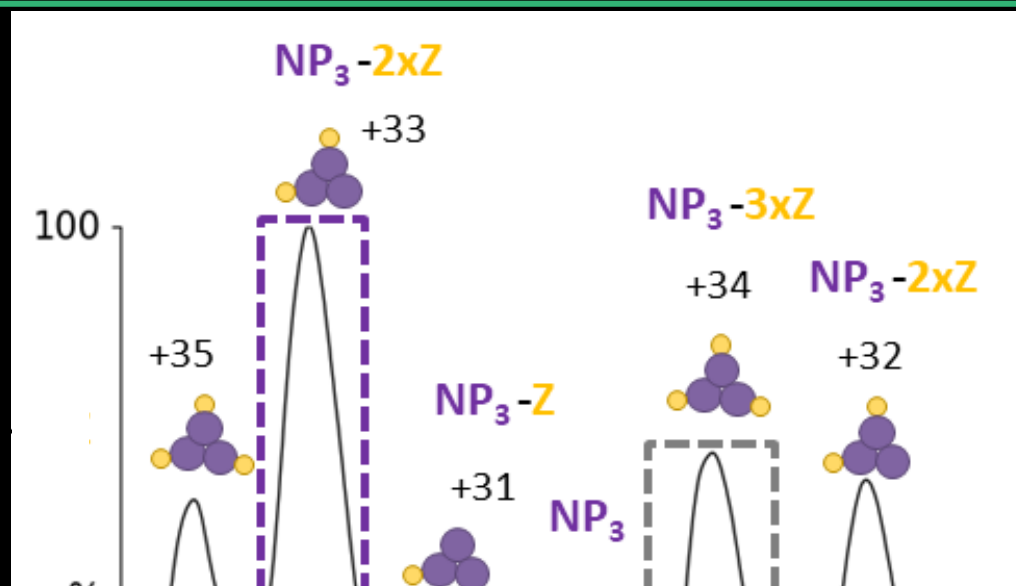
Lassa Virus



- Trimeric NP
- Monomeric Z

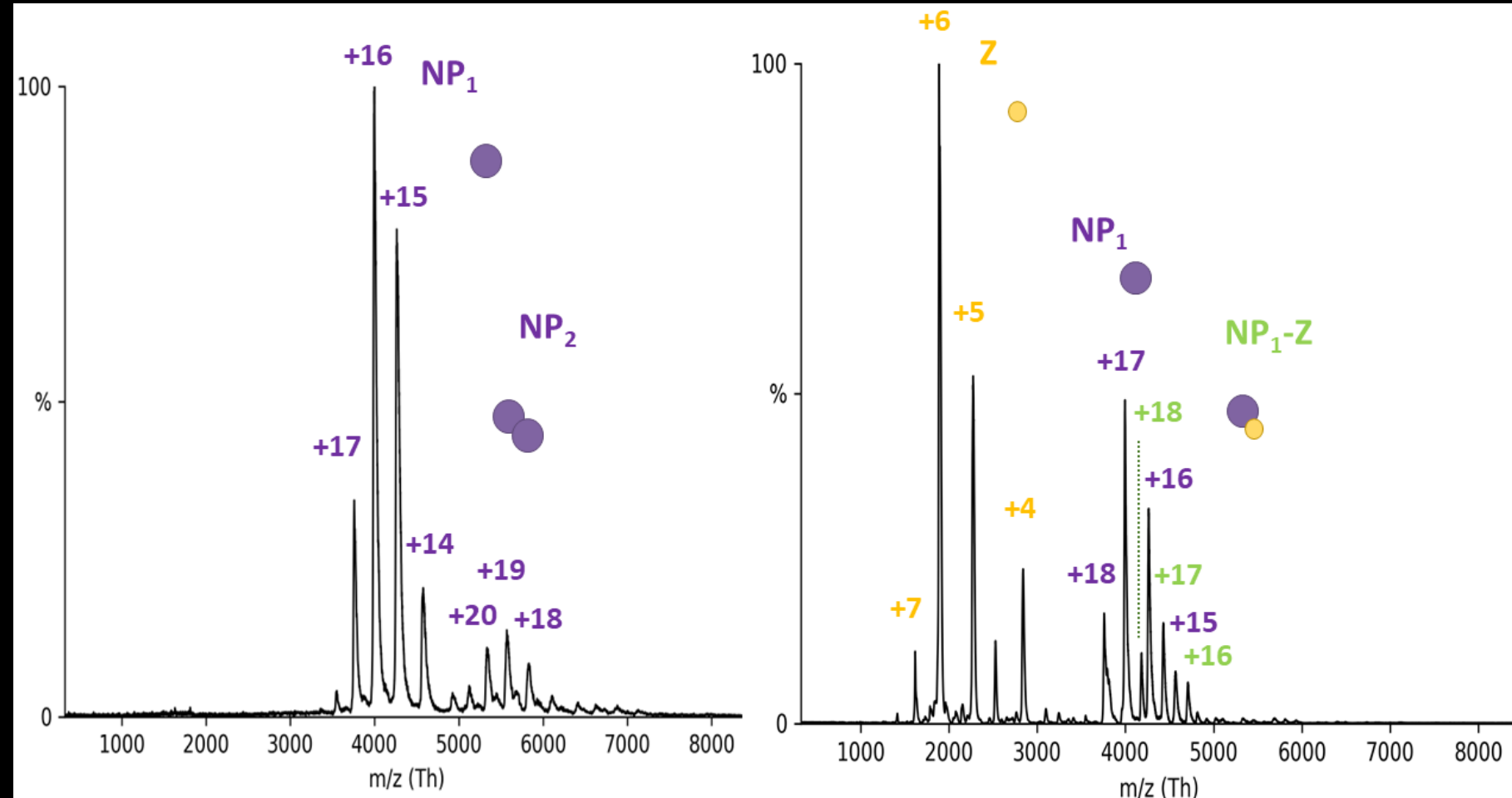


- Up to 3 Z per NP₃
- K_D ~ 110 ± 10 μM

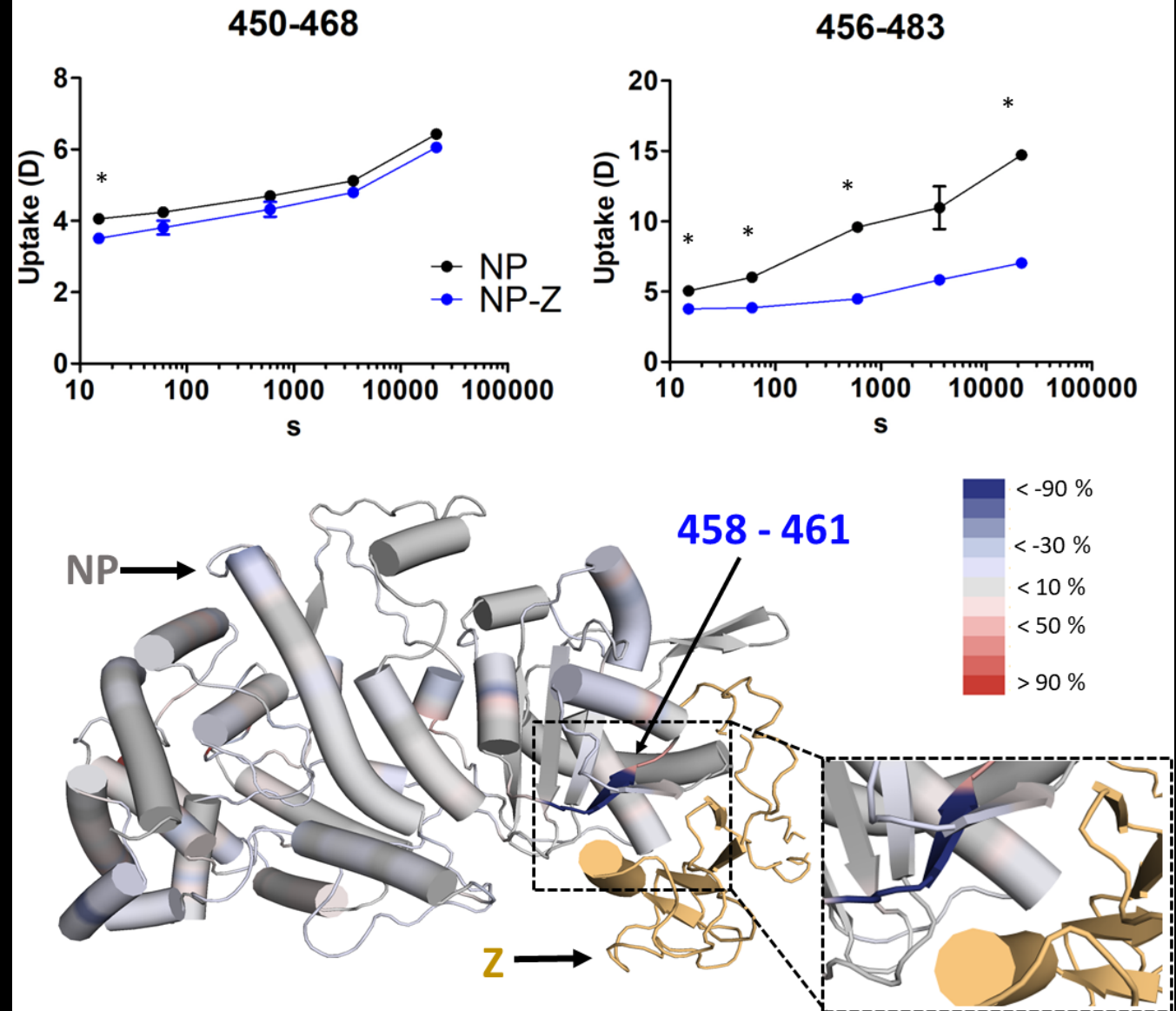
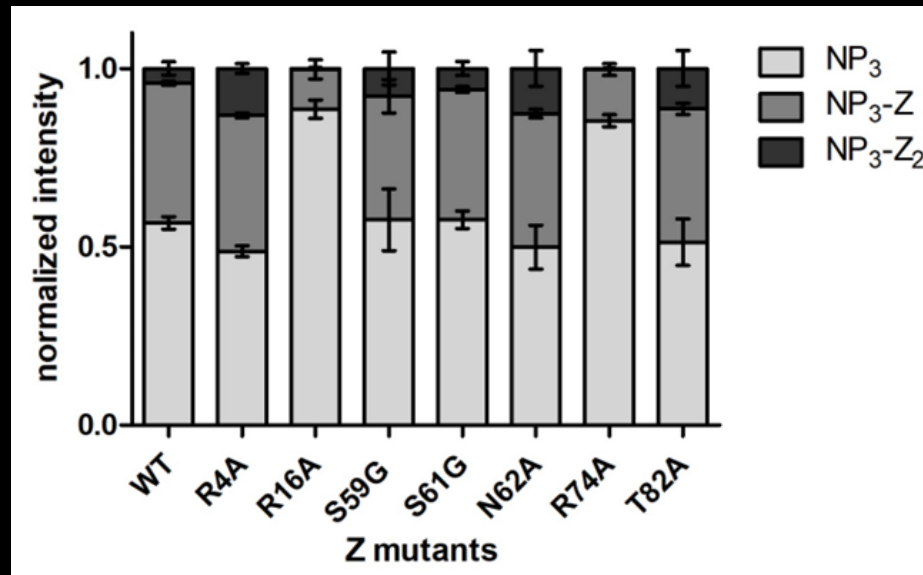


- R52A NP trimerization mutant

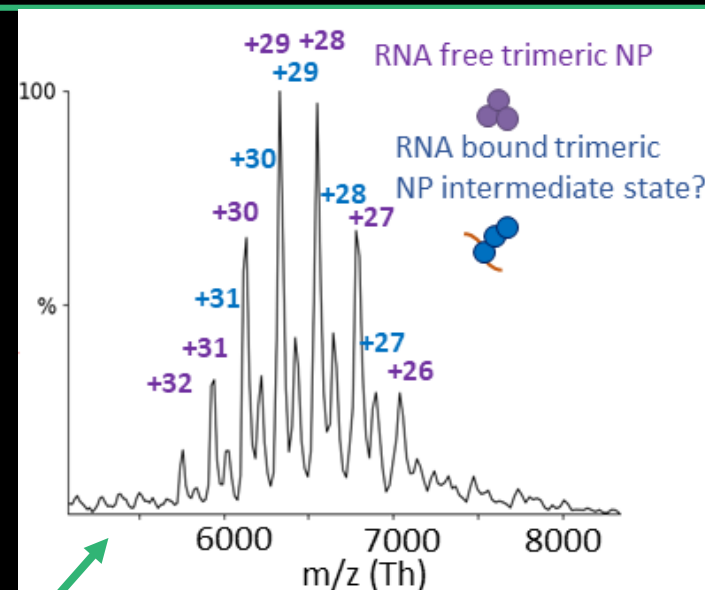
→ $K_D \sim 33 \pm 2 \mu\text{M}$



- AlphaPulldown
- Z via NP C-terminal domain
- HDX-MS NP₃ vs. NP₃-Z
- Z binding mutants

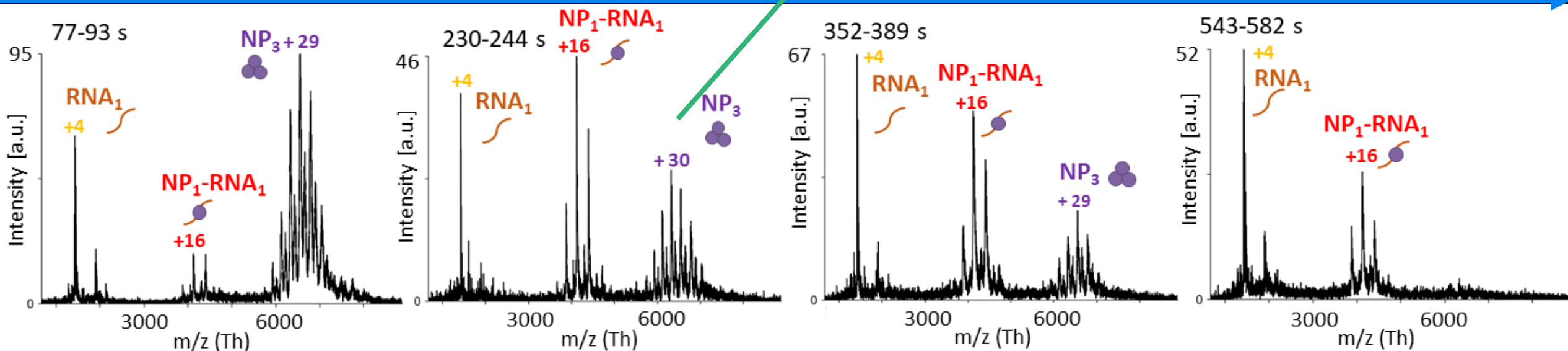


- 9 nt RNA dissociates NP₃
- NP₃-RNA as intermediate

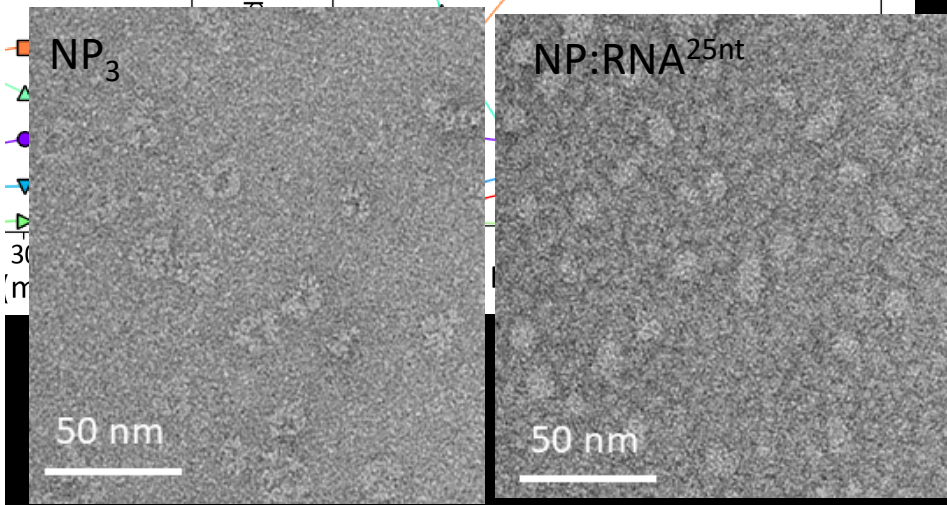
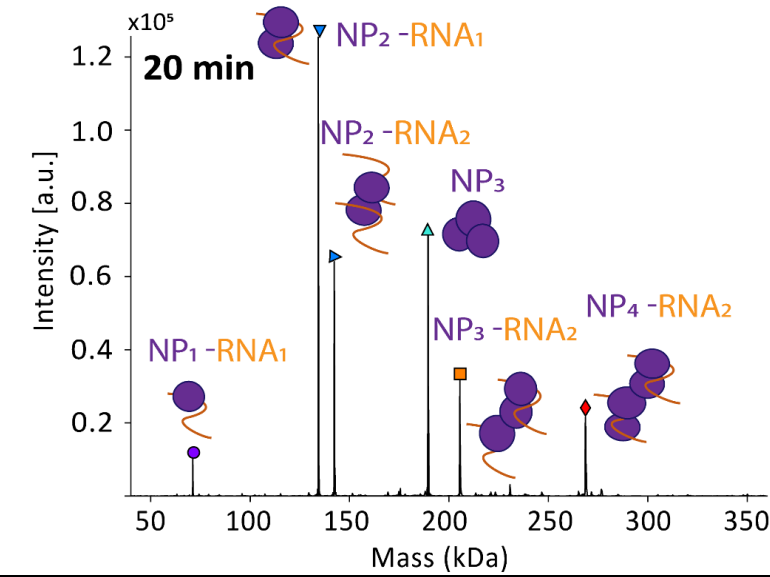
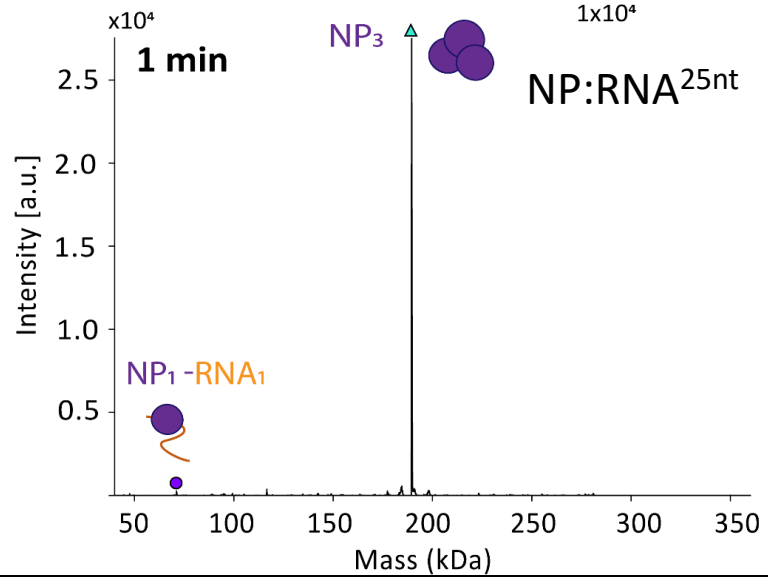
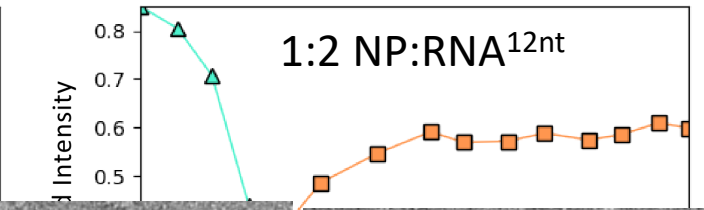
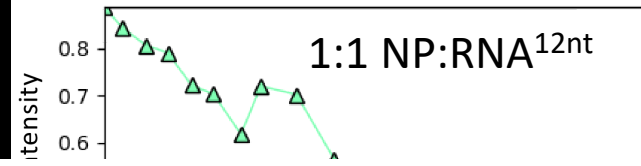
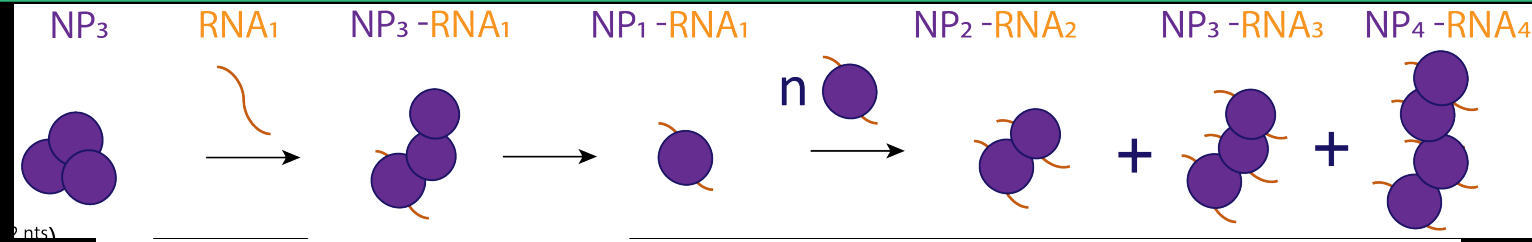


Start

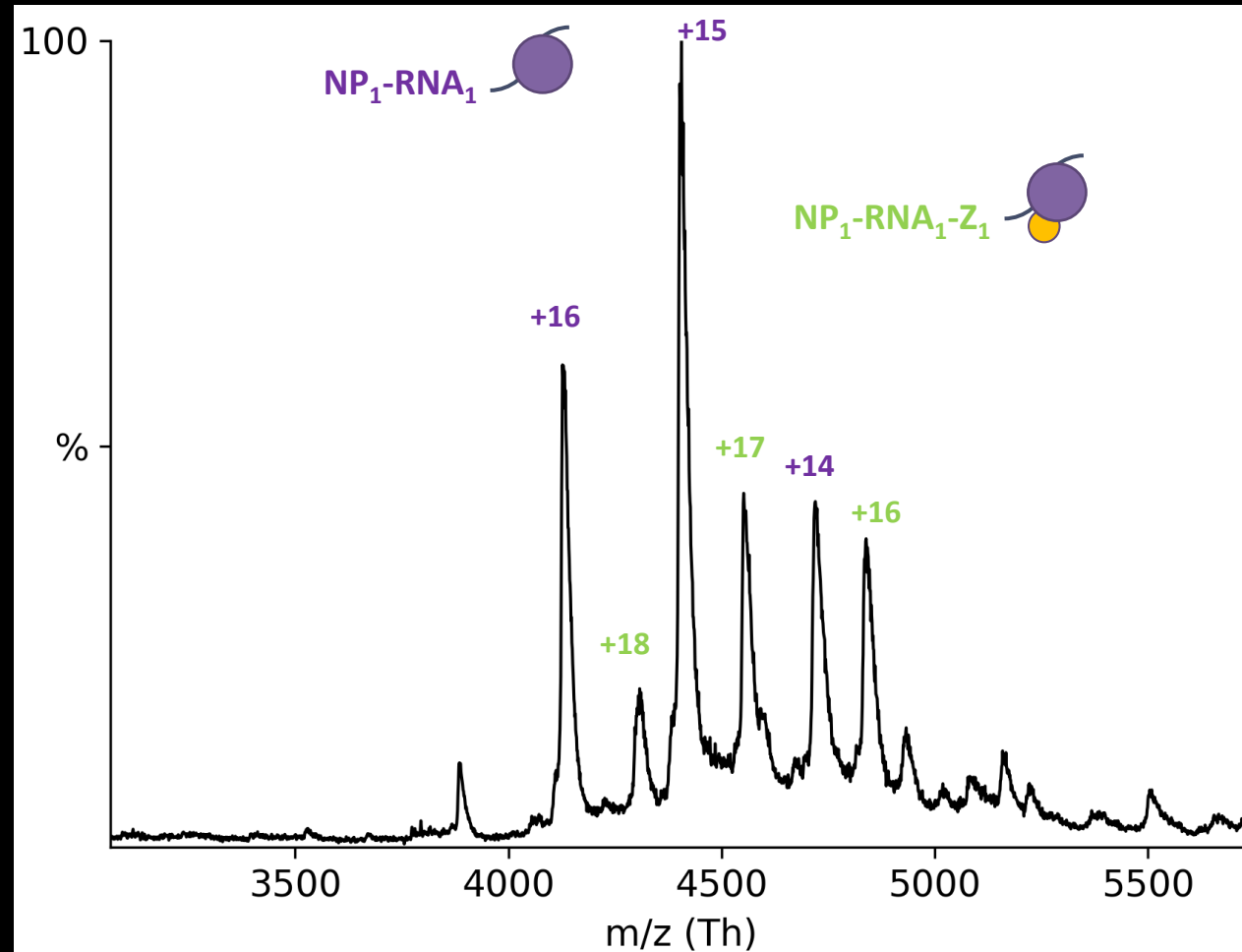
End



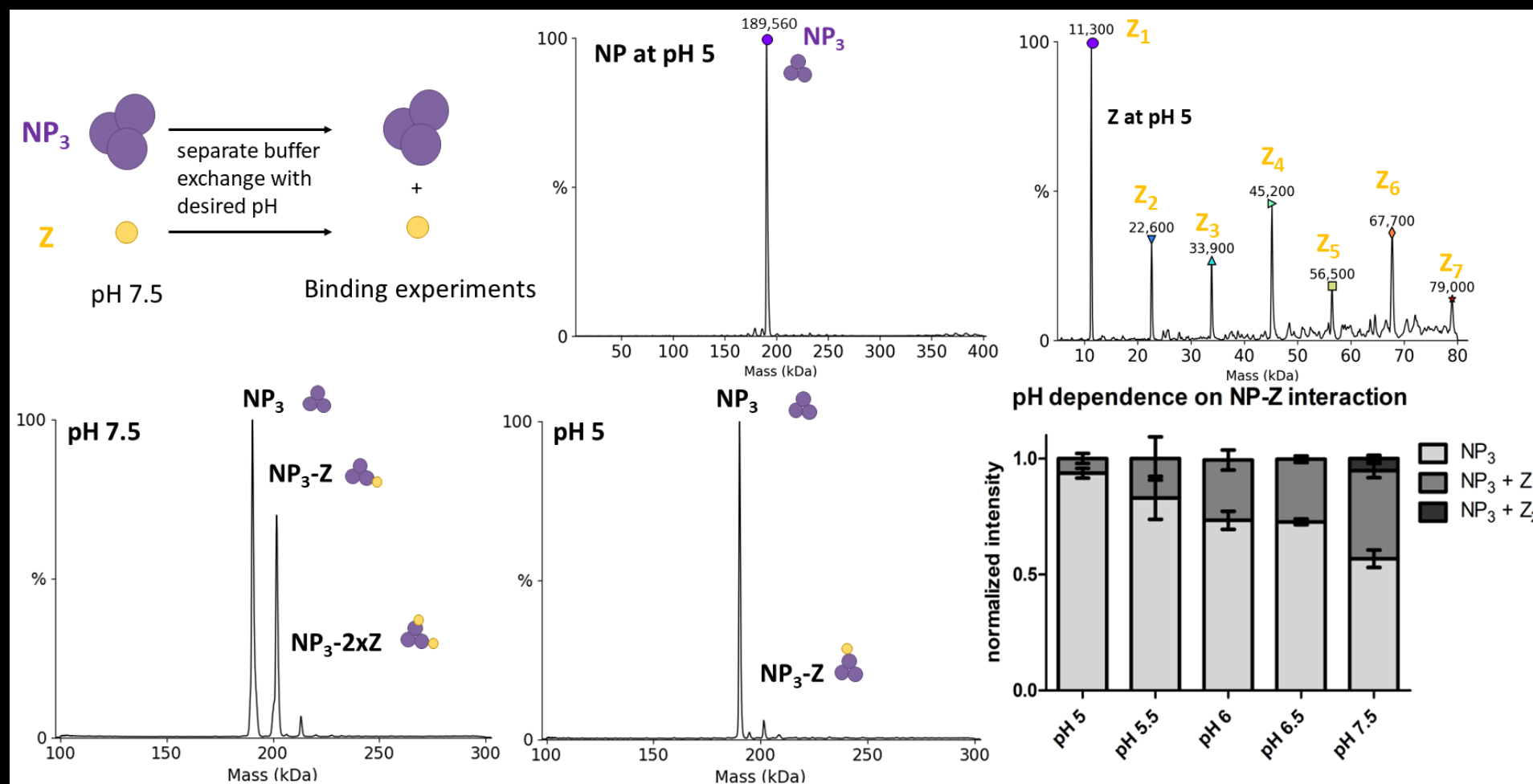
- RNP assembly
 - ≥ 18 nt \rightarrow 2 NP / RNA



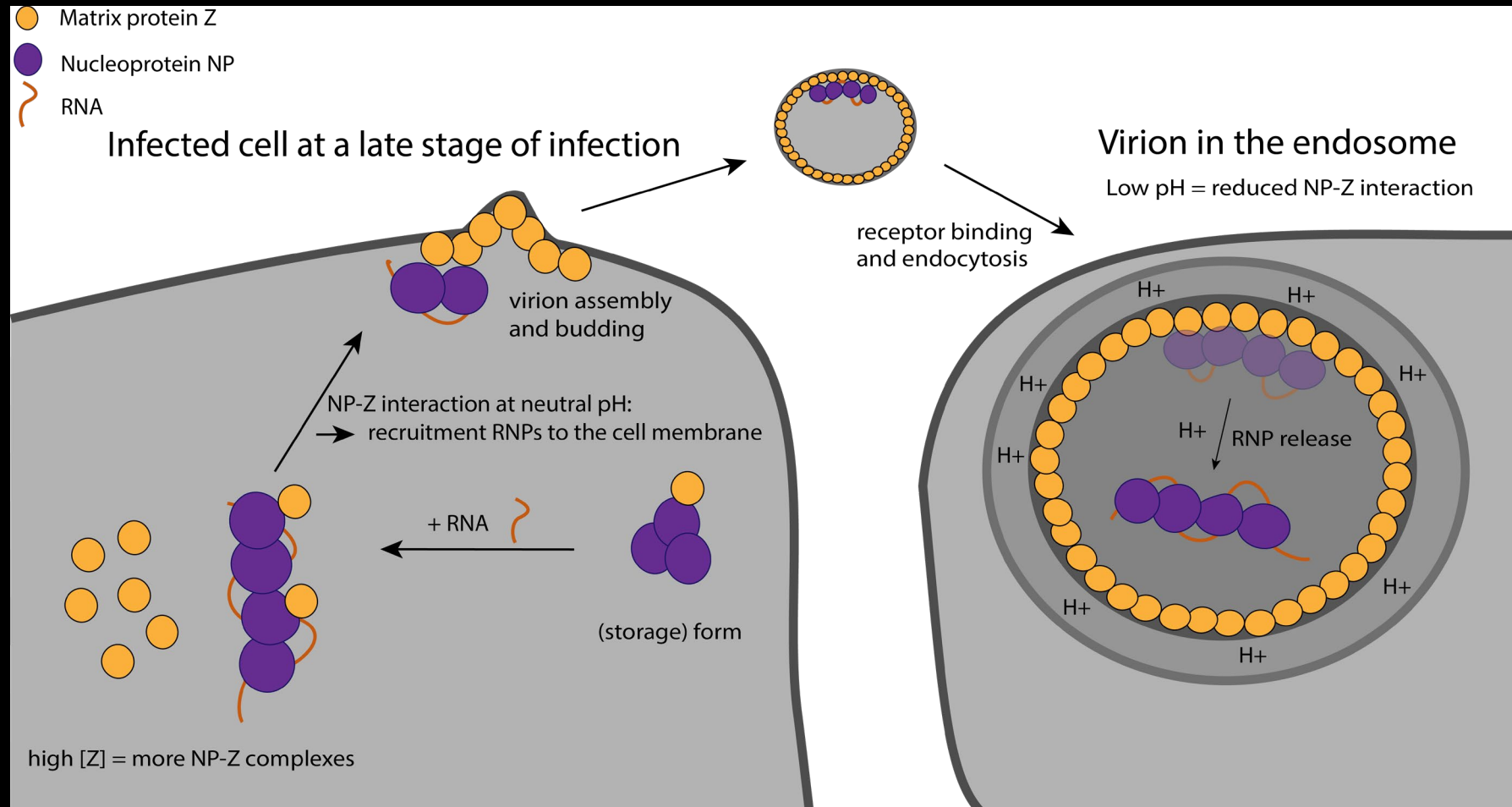
- RNA & Z bind simultaneously



- Z oligomerization & dissociation at low pH

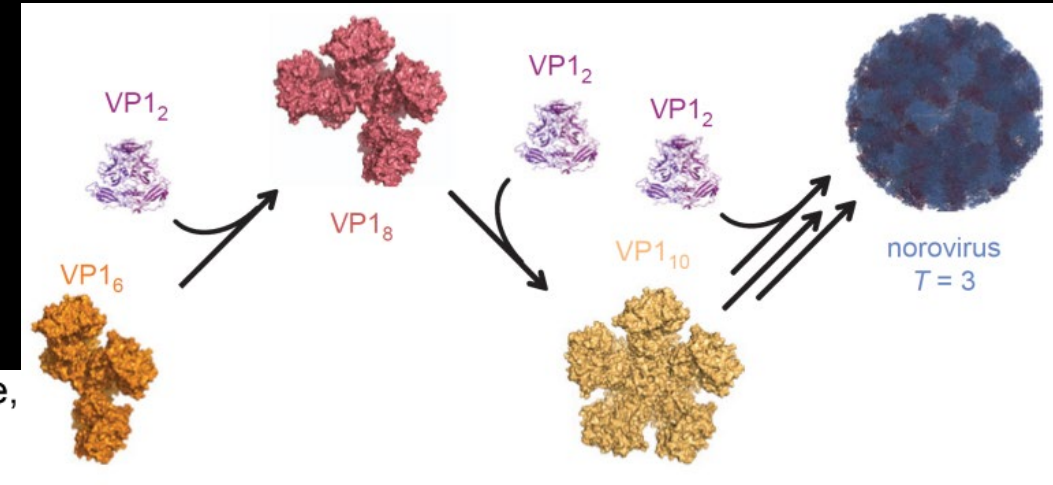


- RNP assembly, recruitment and uncoating

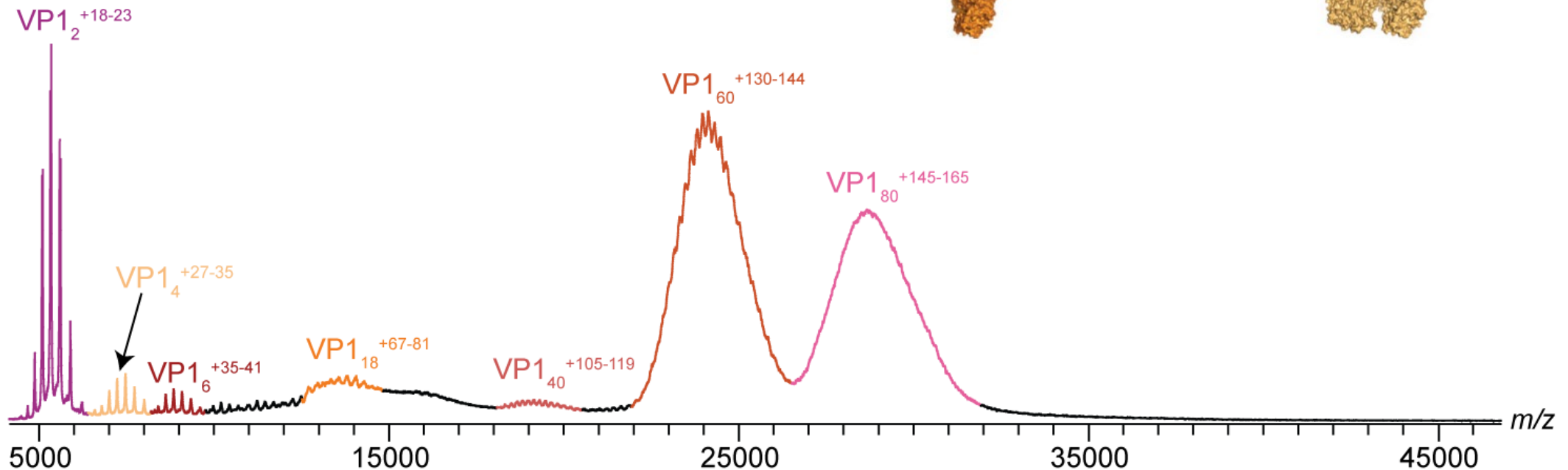


Structure of assembly intermediates?

- Assembly model from ion mobility data
 - Nucleation – fine-tuned process
 - Low abundant intermediates
- no detailed structures

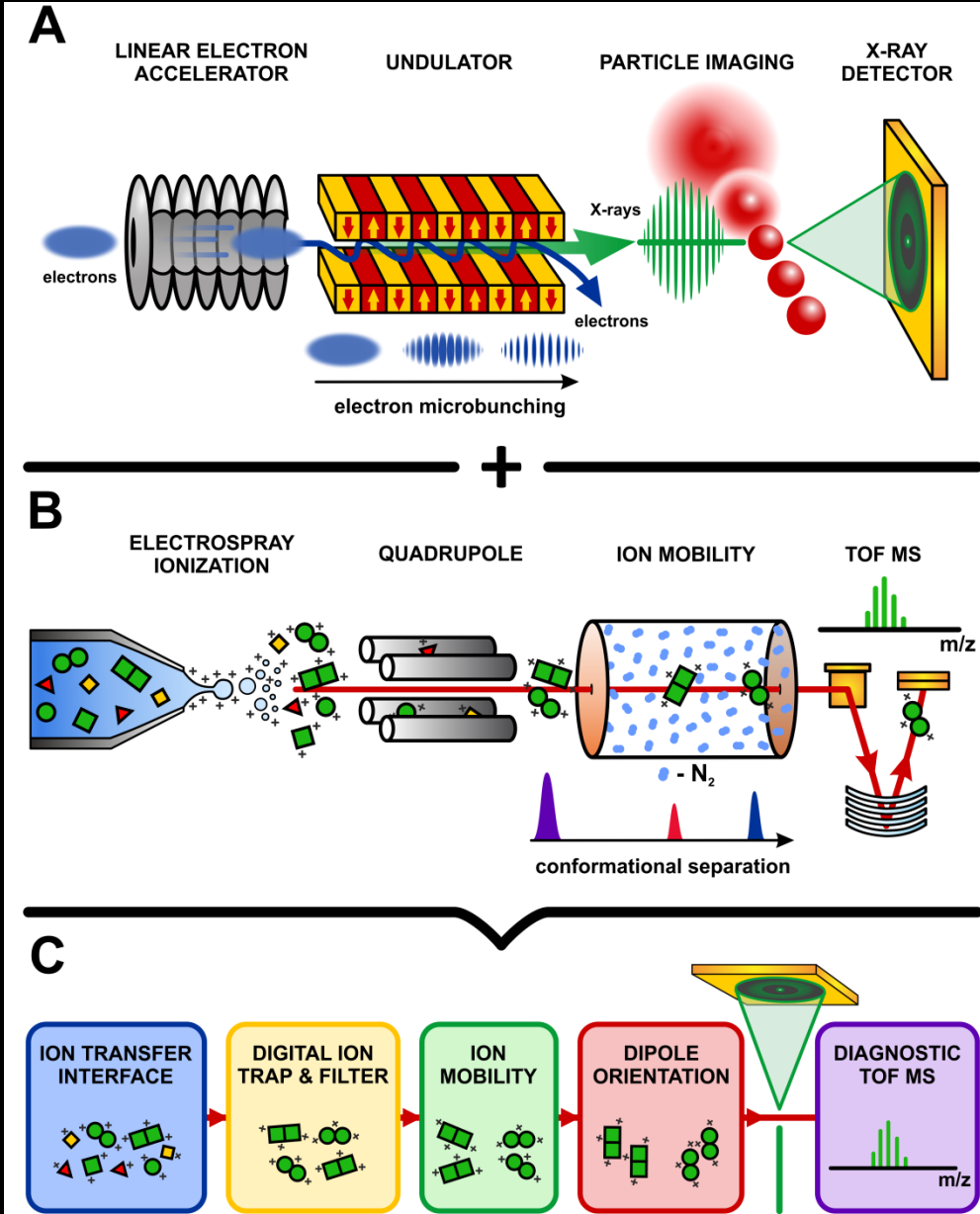


250 mM ammonium acetate,



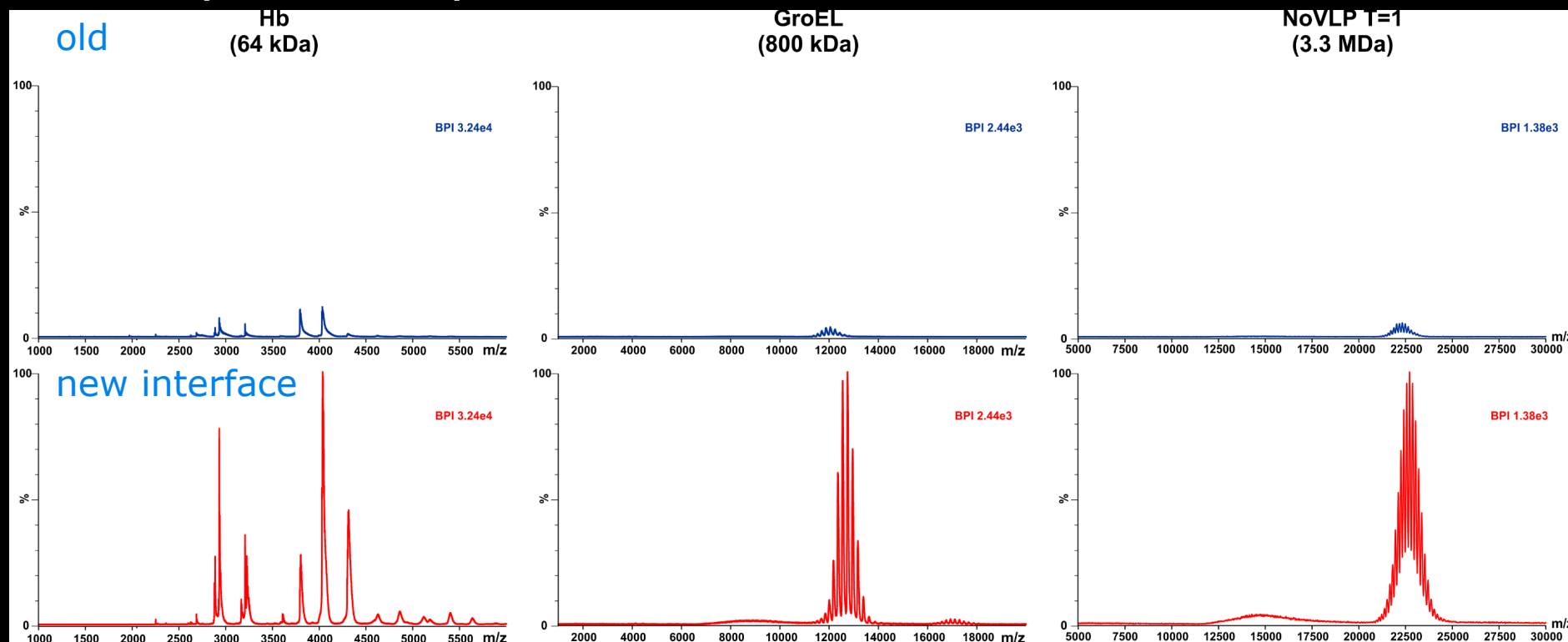
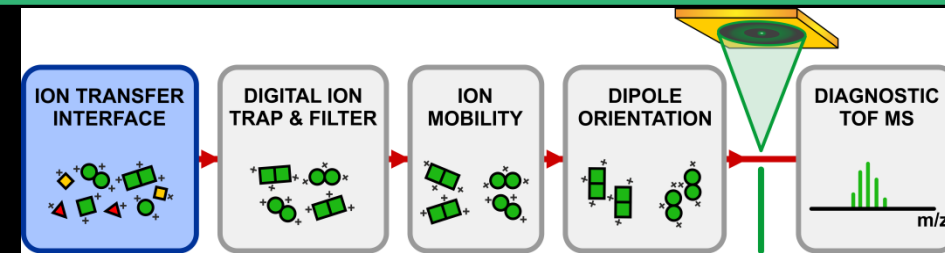
Why use native MS at European XFEL?

- *Imaging before destruction*
 - Potential for transient / low abundant states - sorting problem
 - Successful scattering down to GroEL (sqS, F Maia et al)
 - Use EuXFEL for SPI
- Native MS for selectivity and low background
 - Gas phase structural integrity (Esser et al PNAS 2022)
- *Imaging on the fly*



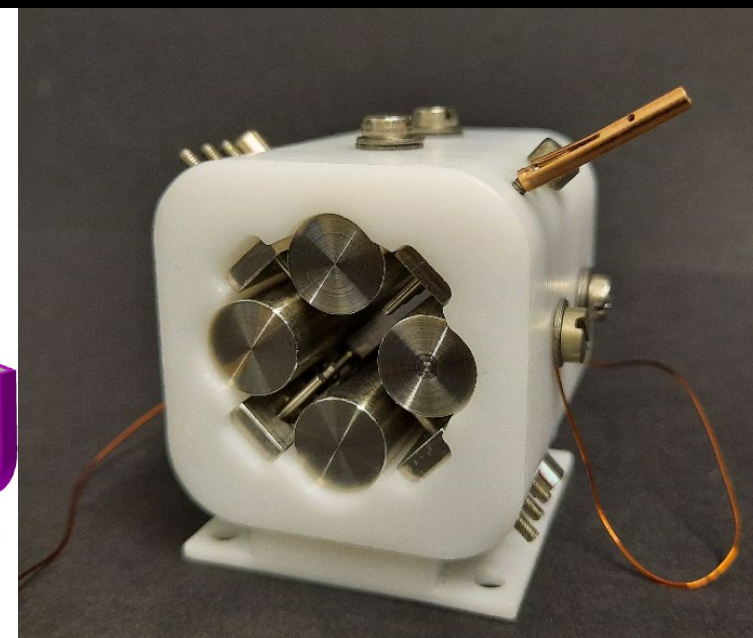
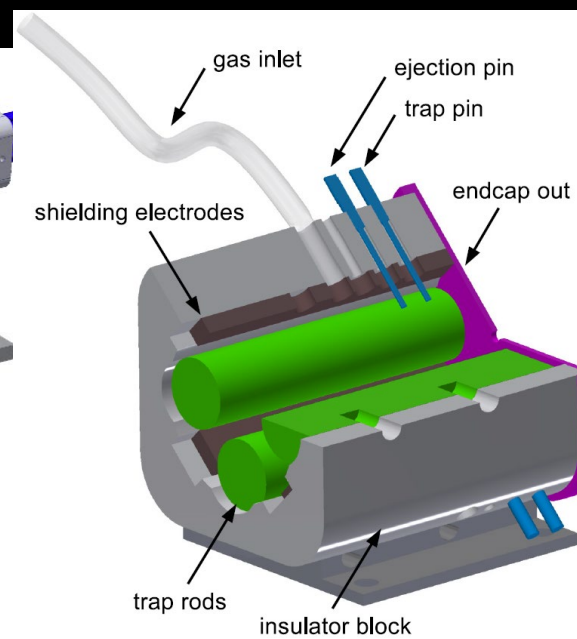
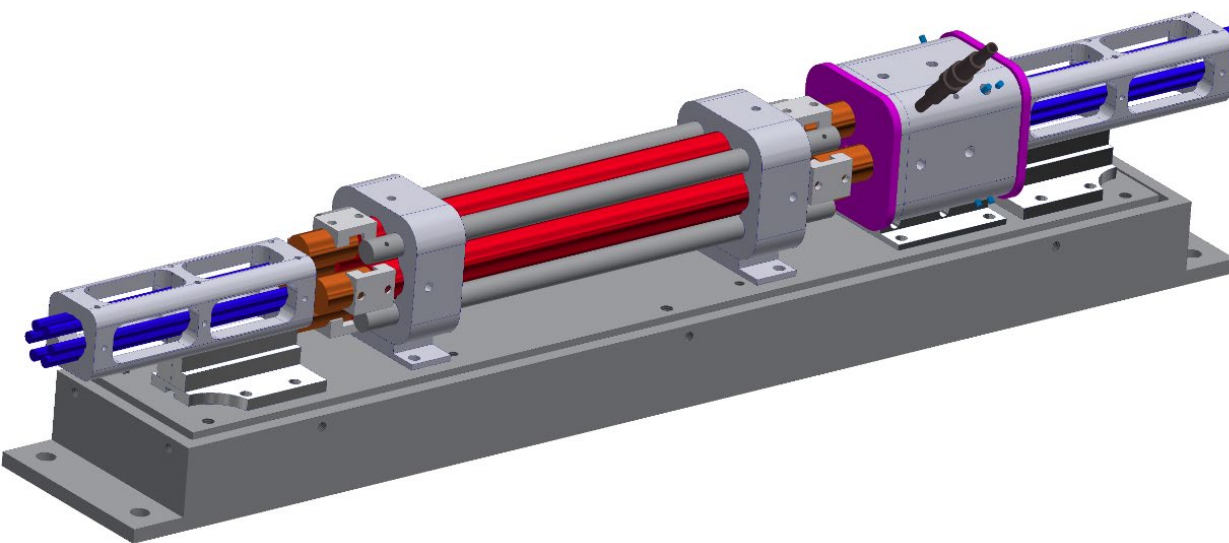
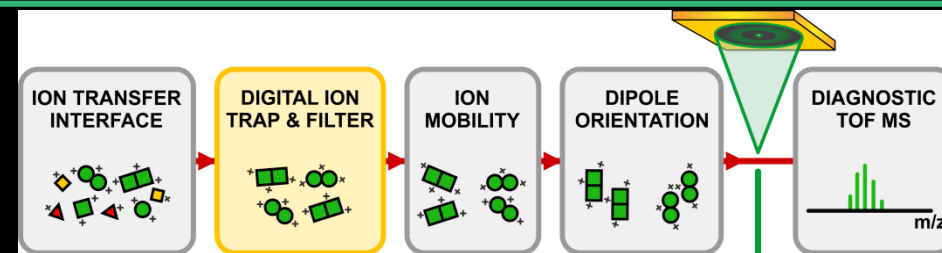
Why use native MS at European XFEL?

- nanoESI
 - low background & sample consumption
 - 10,000 patterns in 16 min with 1 μm focus
 - Aerolens (Fasmatech)



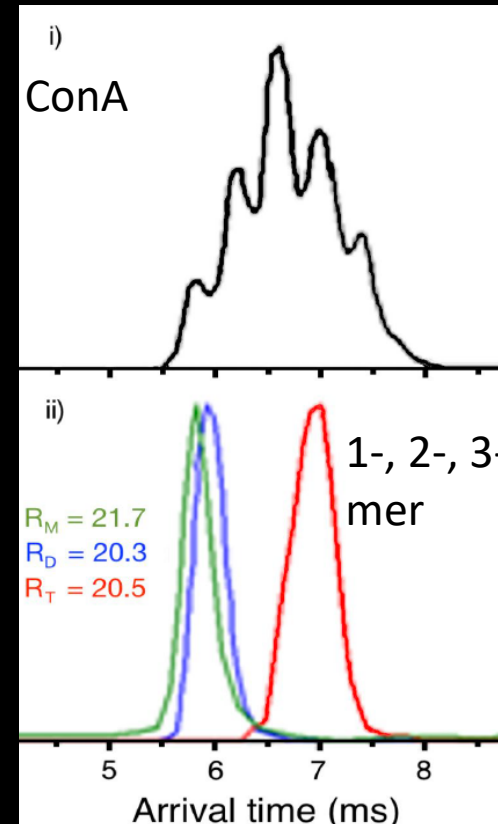
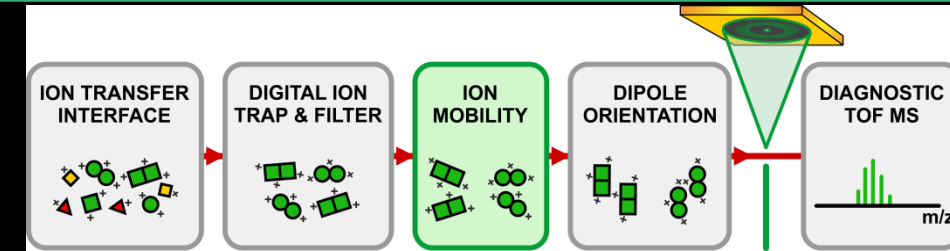
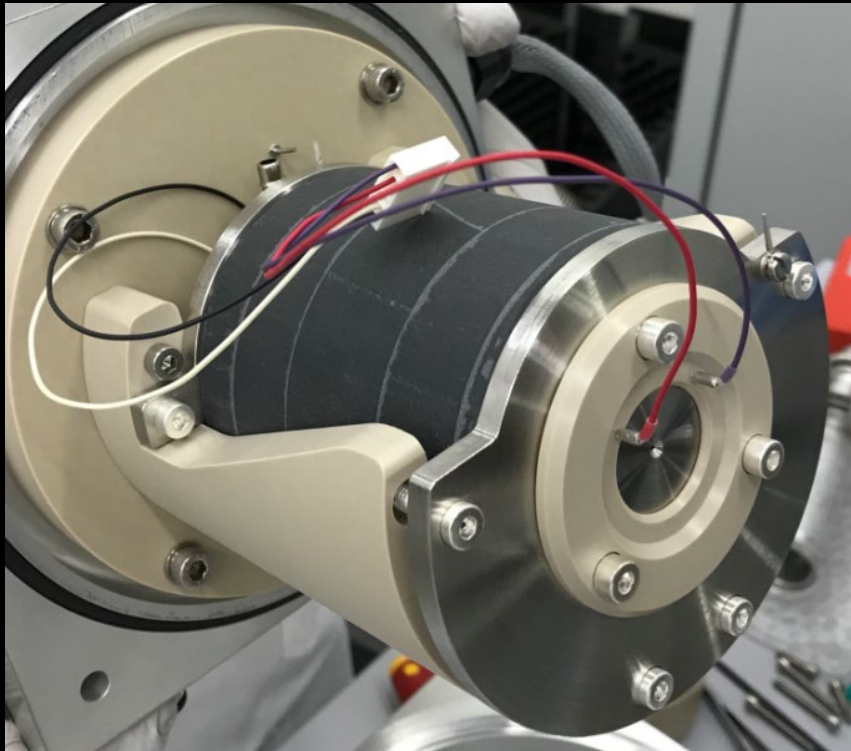
Why use native MS at European XFEL?

- Digital quadrupole & trap (Greifswald)
 - Mass selection → purify low abundant species
 - Time particle release
 - Increase ion density
 - Trapping capacity for 100 ms
 - No structural damage



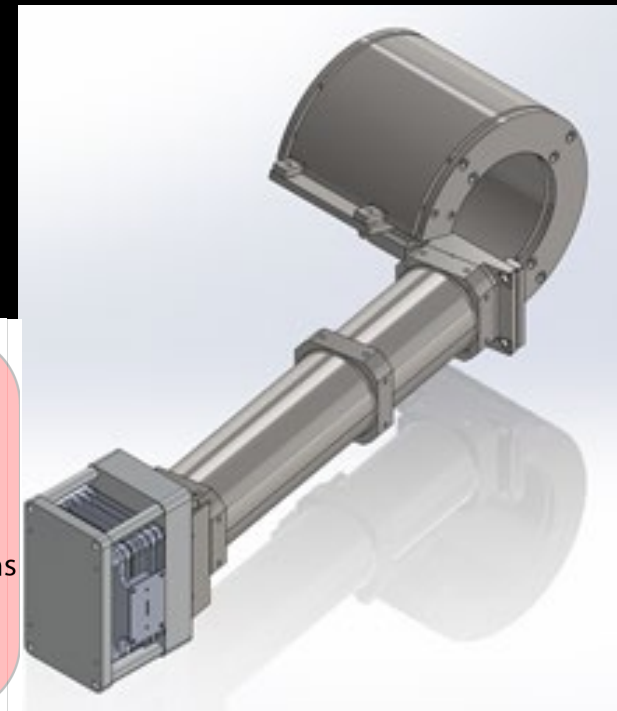
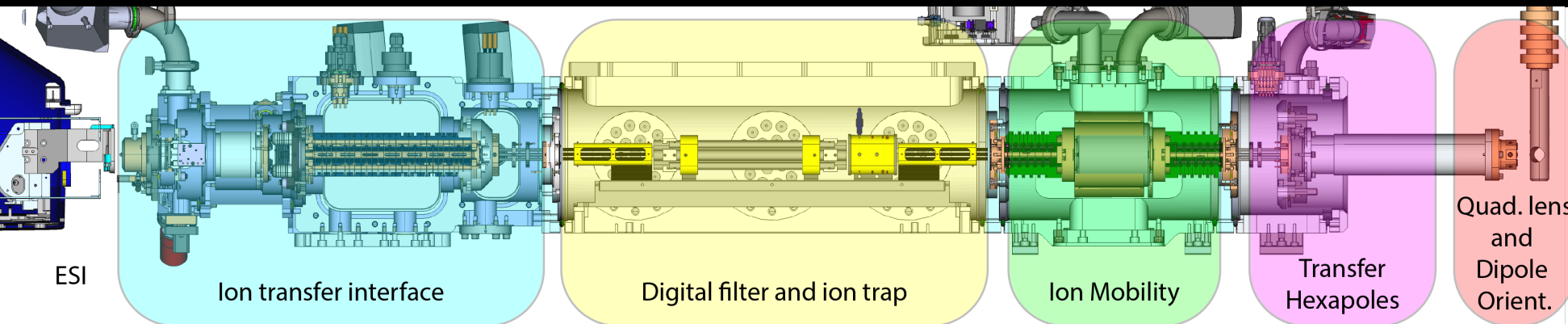
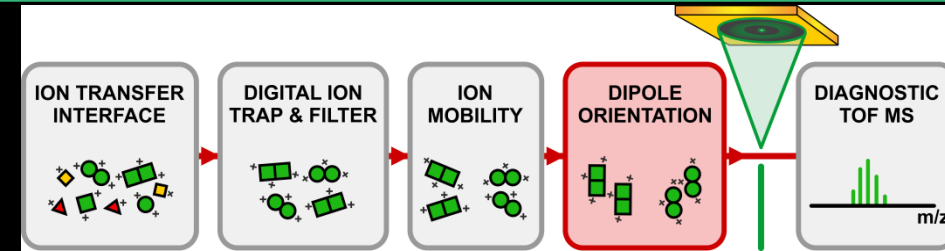
Why use native MS at European XFEL?

- Ion mobility separation (Manchester/ MS Vision)
 - Conformational separation



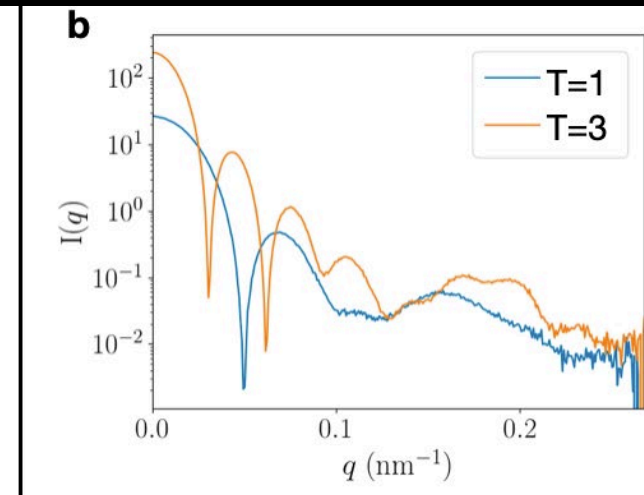
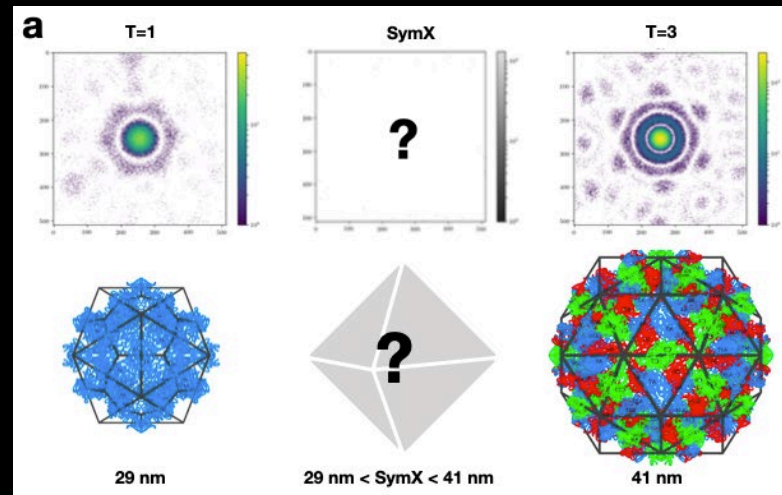
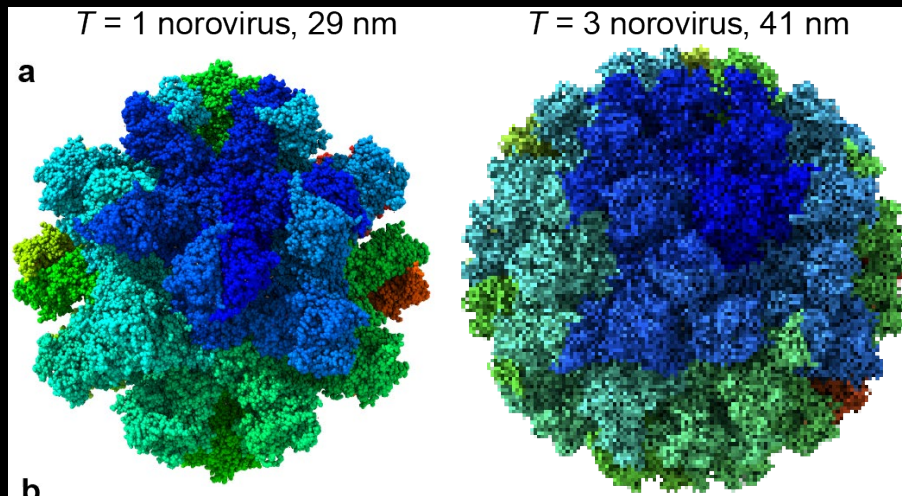
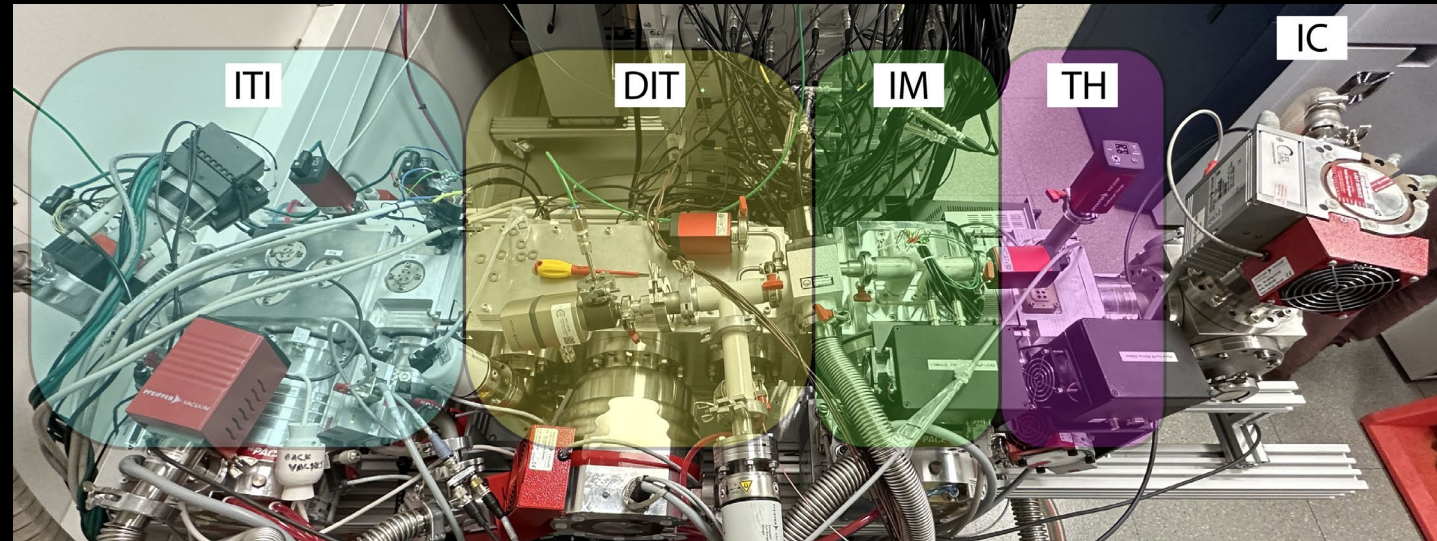
Why use native MS at European XFEL?

- Dipole orientation (Uppsala, Fasmatech)
 - Simplify pattern assembly
 - Use patterns with scarce / missing data
- ToF – online diagnostics (MS Vision, John Hoyes)
 - Sample quality & influx
 - Proper selection

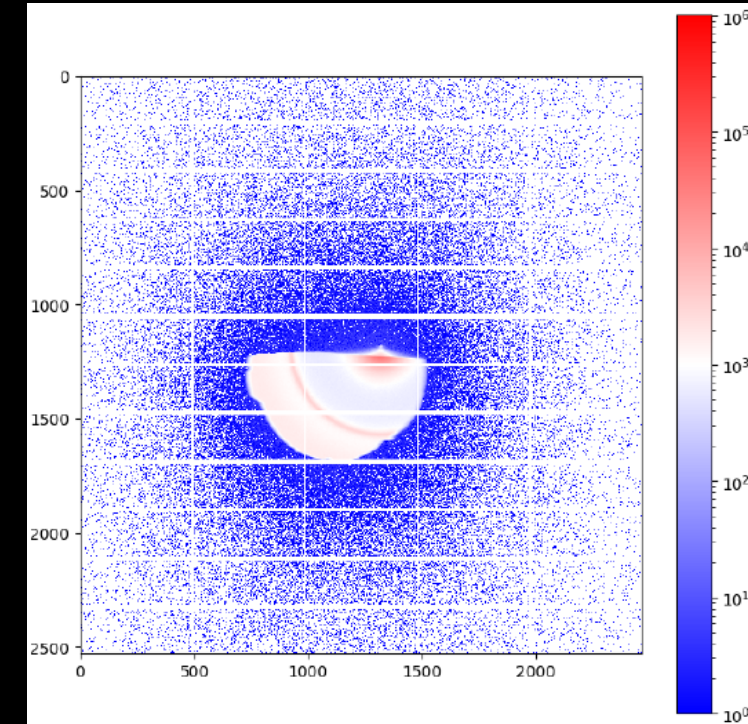
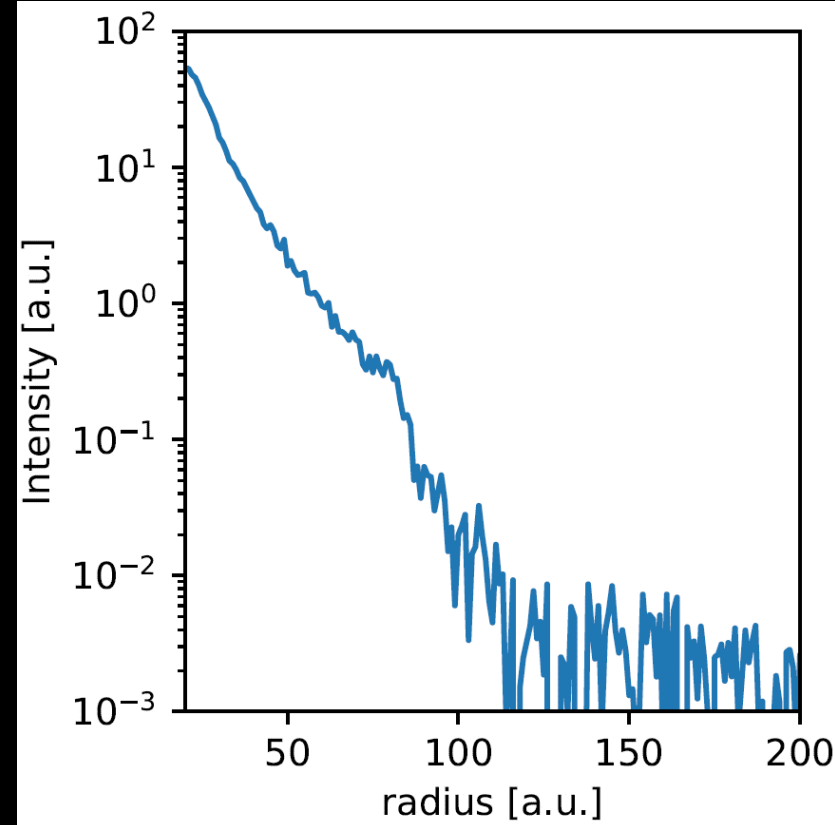


Why use native MS at European XFEL?

- Proof-of-principle on norovirus
- Identify symmetry



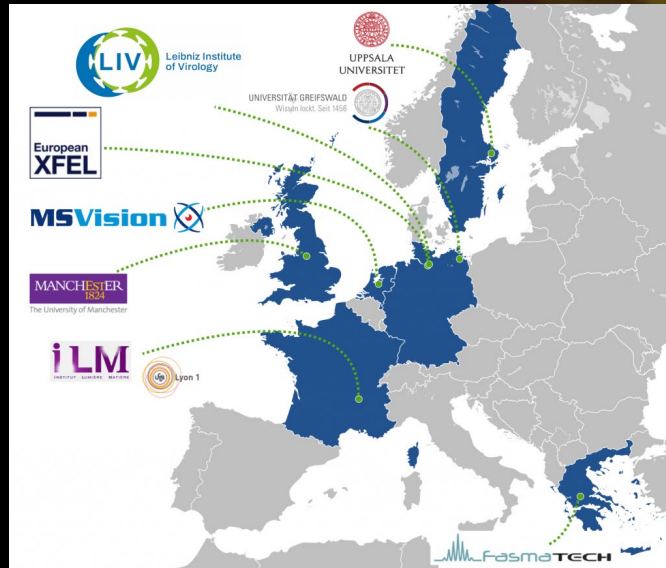
- Nov 2022 – dominated by beamline background
- Nov 2023 – signal after background reduction
 - 20 nm Au nanoparticles, 10 min acquisition
- Next stop proteins 😊
 - Heterogenous samples
 - Time resolved



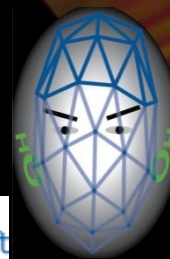
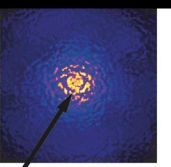
Silver behenate powder diffraction

Acknowledgements

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MS SPIDOC PRESENTS:

<https://xfel.tind.io/record/2561>

NOROVIRUS SUPERSTAR

VÉRO MISCHTIZ



Molecular Biophysics

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