

Hierarchical assembly of capsids is robust

S. Fraden

The Team



Gregory Grason



Douglas Hall



Hendrik Dietz



Chris Sigl



Fabian Kohler



Alba Monferrer



Daichi Hayakawa



Botond Tyukodi



Wei-Shao Wei



Rupam Saha



Anthony Trubiano



Thomas Videbæk

Brandeis
bioinspired
MRSEC



Seth Fraden



Mike Hagan



Ben Rogers



Tijana Ivanovic

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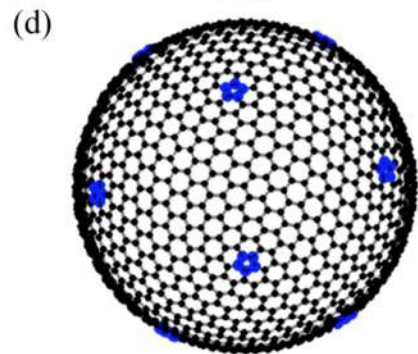
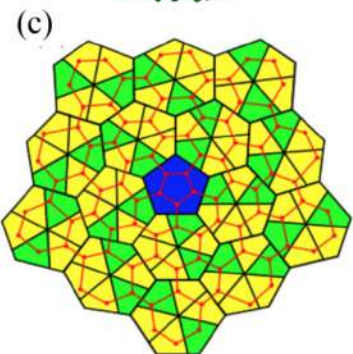
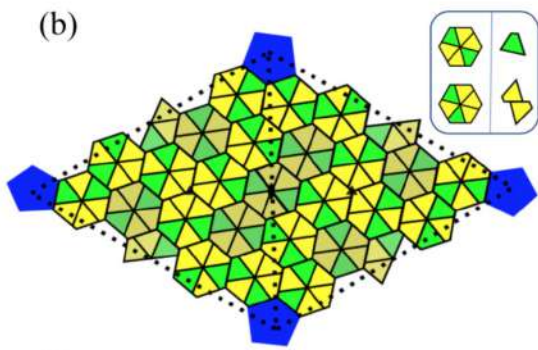
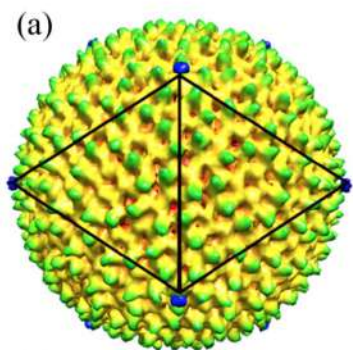
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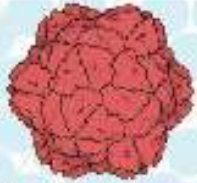
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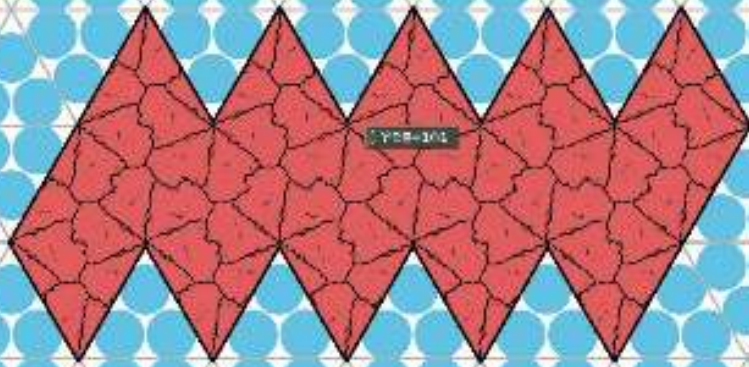
Caspar-Klug Reidun Twarock Geometrical tiling



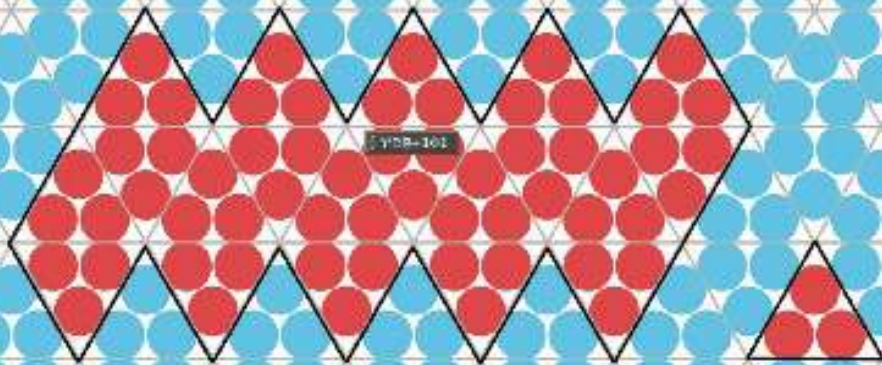


T=1
Satellite Tobacco Necrosis Virus

PDB entry **2buk**



PDB-101



PDB-101

Subunits are in perfect icosahedral symmetry.

Viral Quasisymmetry

Viruses are faced with a challenge: their genomes need to encode all of their proteins, but at the same time, these genomes need to fit into the tiny space of a viral capsid. In the 1960s, Donald Caspar and Aaron Klug discovered that viruses solve this problem using quasisymmetry.

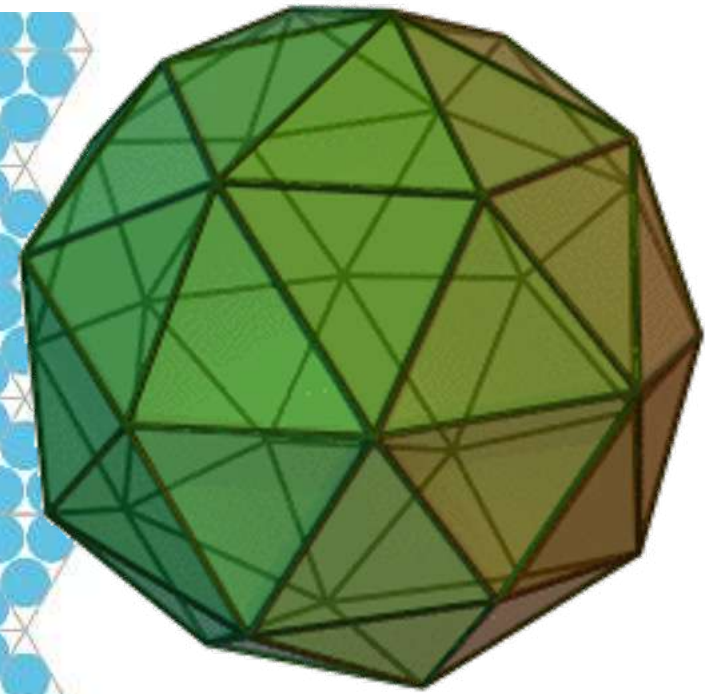
Viral capsids are built using many identical copies of one or a few capsid proteins, arranged to form a shell with icosahedral symmetry. Some viruses, such as Satellite Tobacco Necrosis Virus, build a tiny capsid with perfect symmetry. Other viruses need more room, so they build larger capsids, but still only using one type of building block. With small changes in shape, the subunits form pentamers and hexamers, and these pack into larger, quasisymmetrical capsids.

These paper models show a few examples of how quasisymmetry is used to build viruses of different sizes. The subunits are represented as circles, with ones that form pentamers in red and ones that form hexamers in shades of yellow and orange. For each virus, a model of the atomic structure is also included.

Cut out the models and tape the edges together to form the icosahedral virus.

Learn more at pdb101.rcsb.org

200 YEARS PDB-101

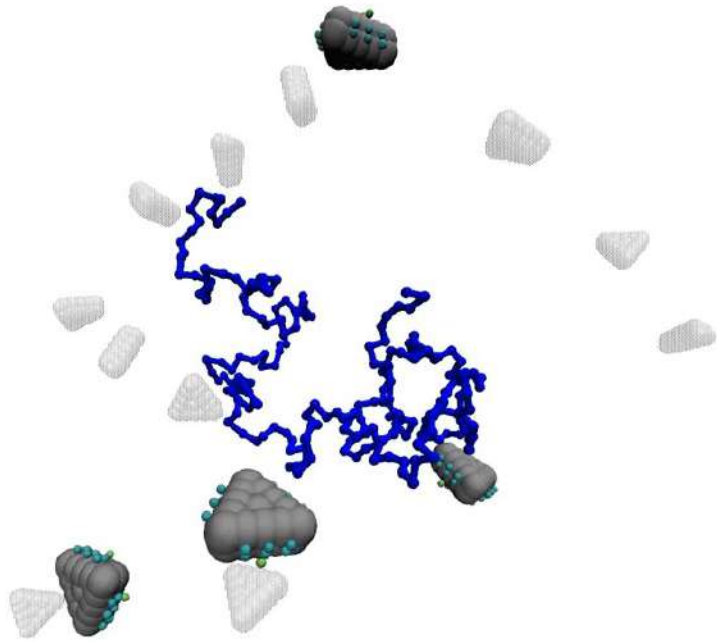


Icosahedra
20 identical triangles

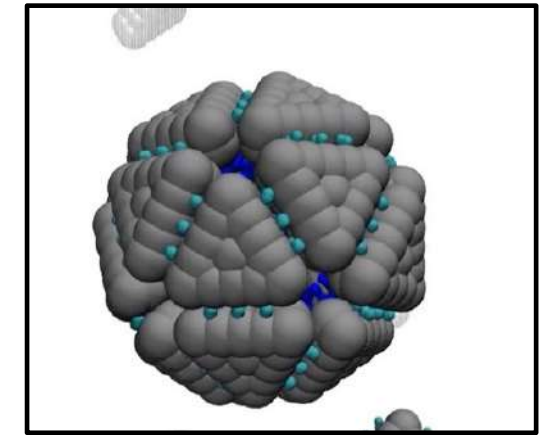
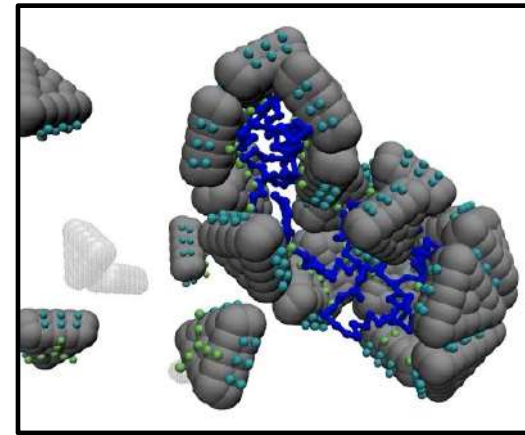
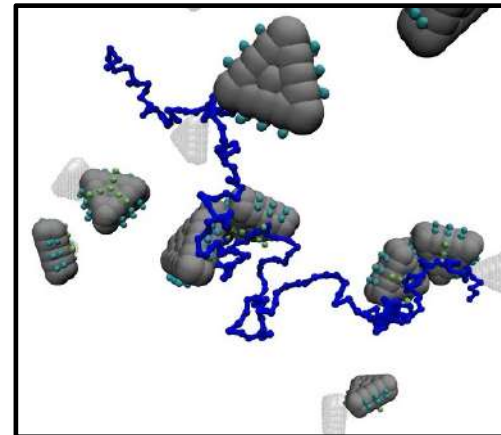
T=1
All sides equivalent

Icosahedron: largest possible structure built out of **identical** subunits

Icosahedral capsid assembly around a flexible polymer

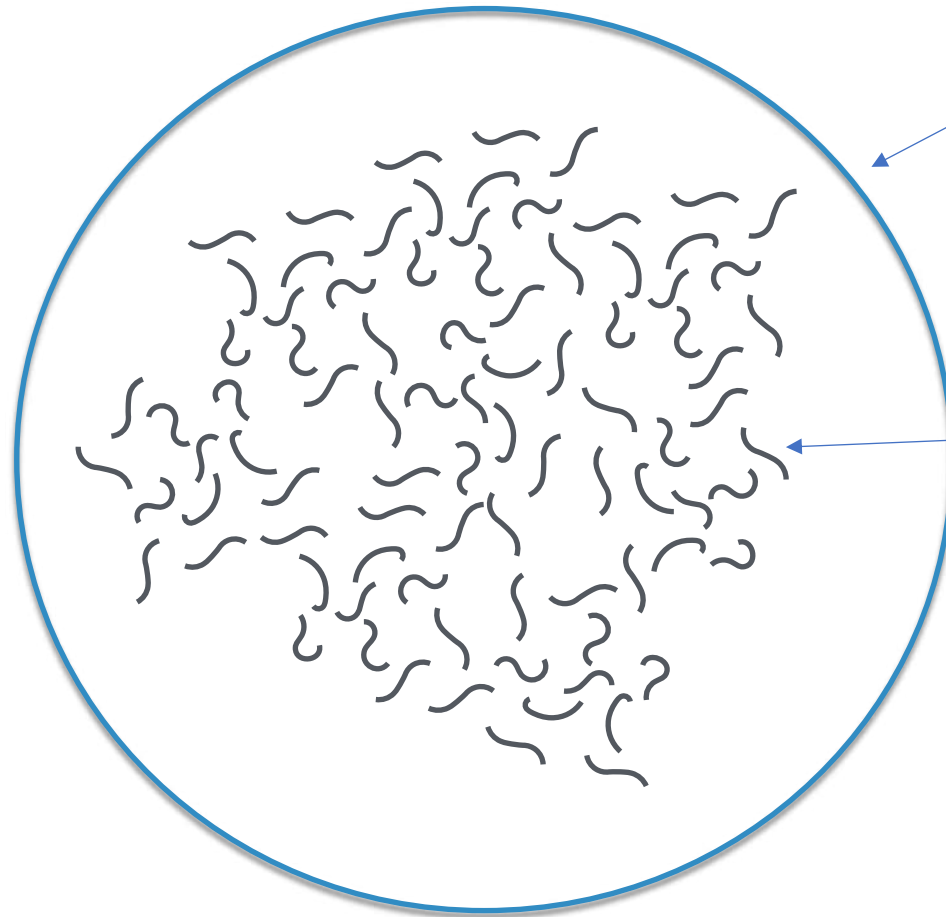
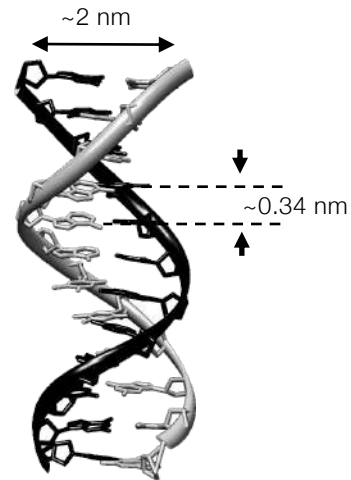
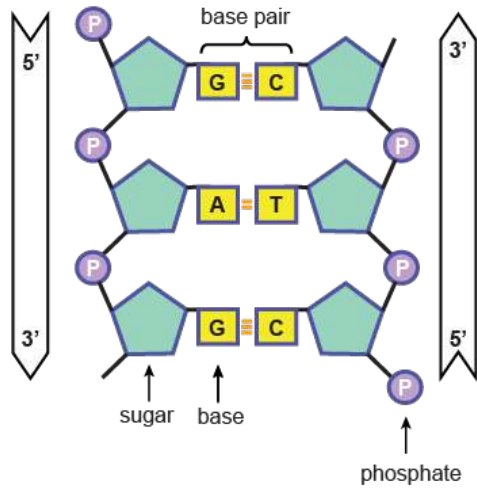


- Colloidal triangles
- Control rigidity
- Control dihedral angle
- Control valency
- Control specificity



Elrad O.M., & Hagan M.F.
Physical biology (2010)

DNA origami



Scaffold

- M13mp18
- ~ 8000 bases
- 1 circular

Staples

- chemical synthesis
- ~ 50 bases
- ~ 200 linear



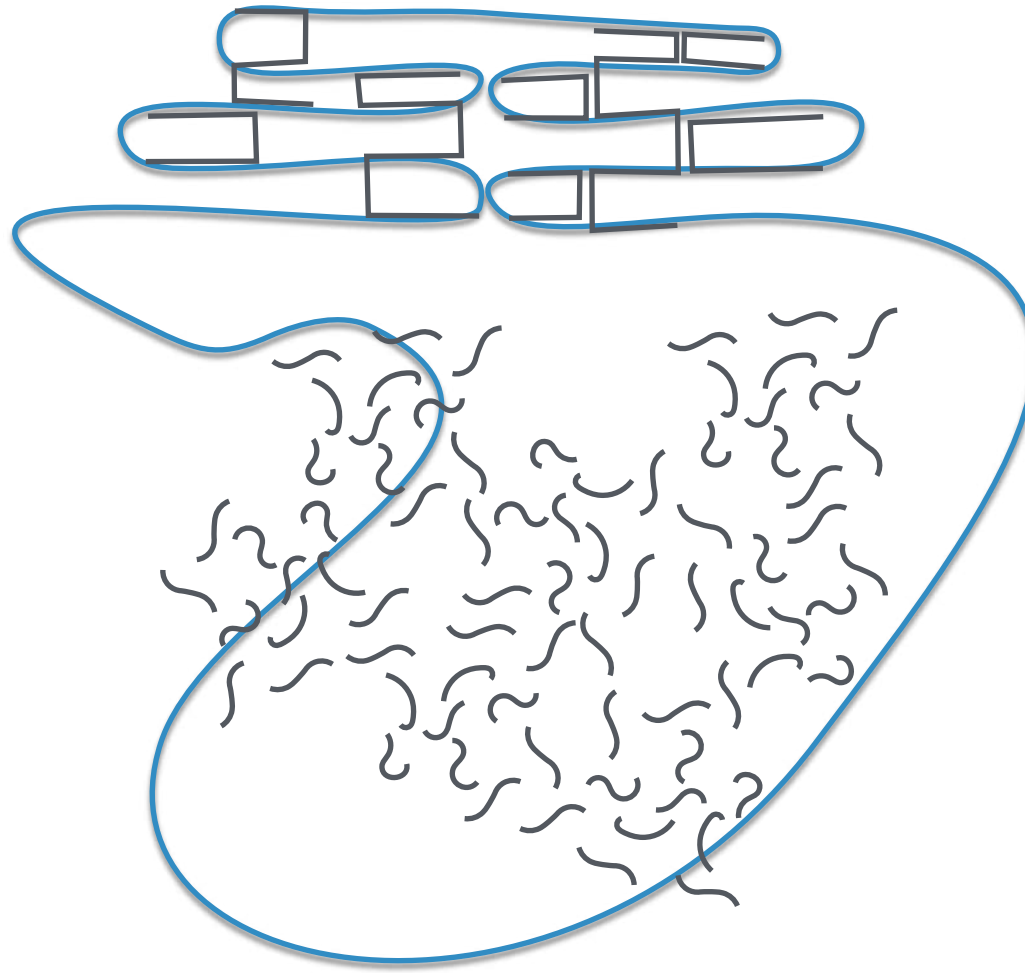
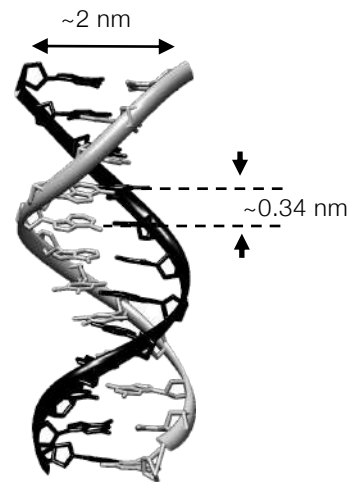
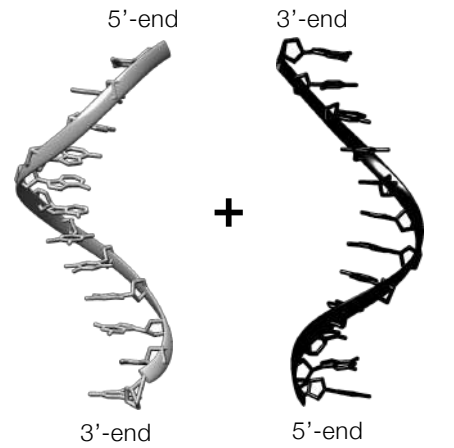
Hendrik Dietz



Chris Sigl



DNA origami



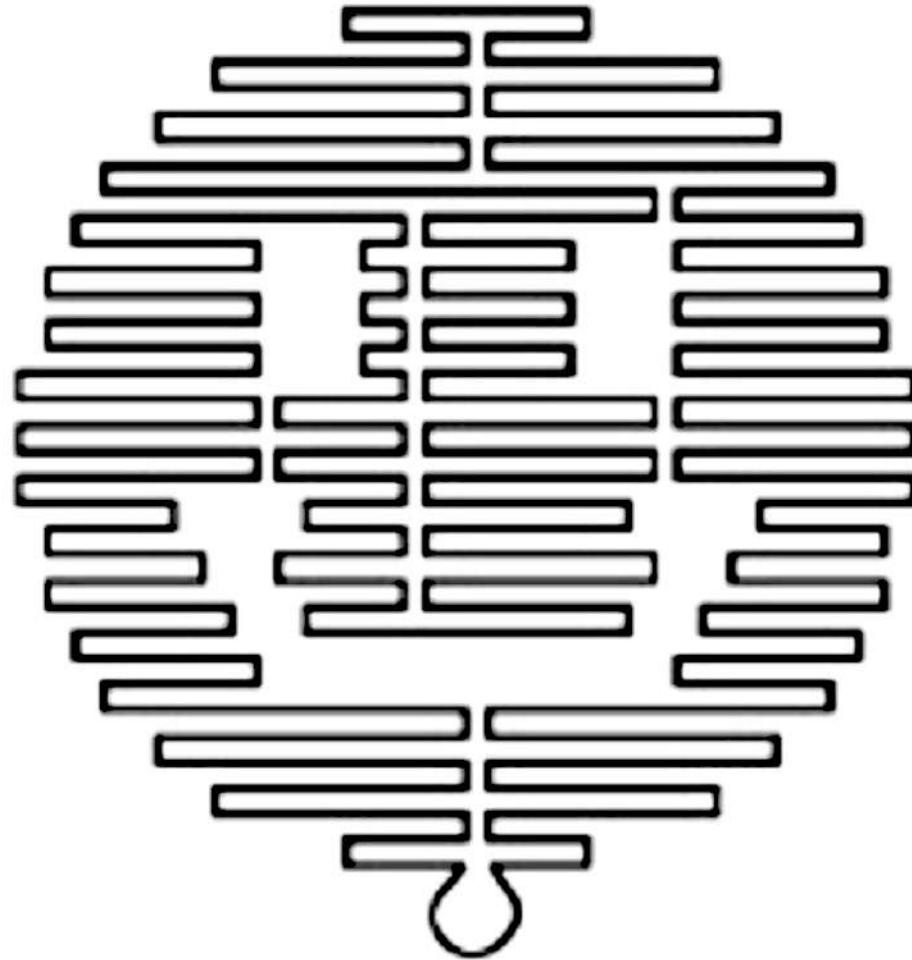
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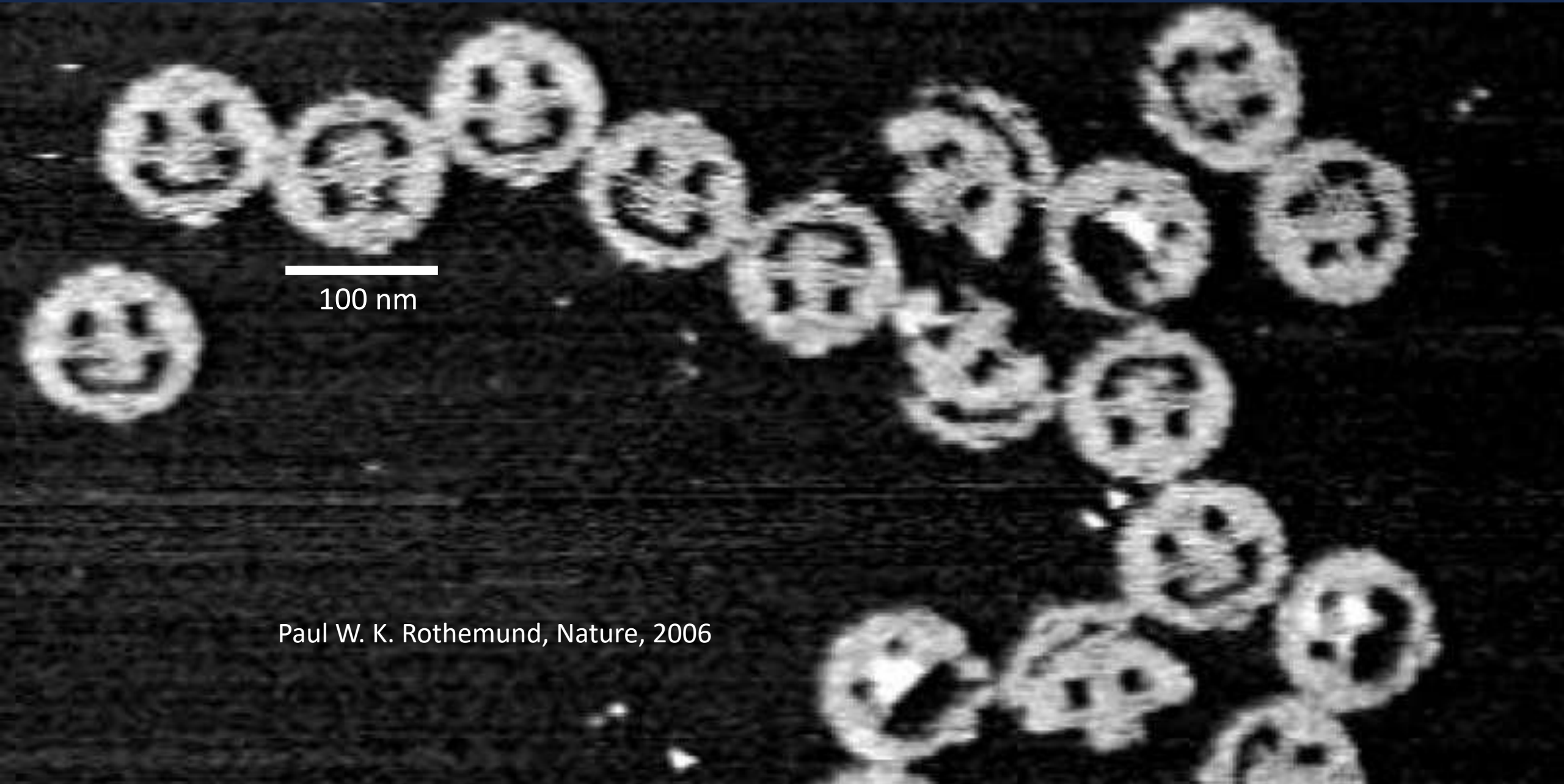
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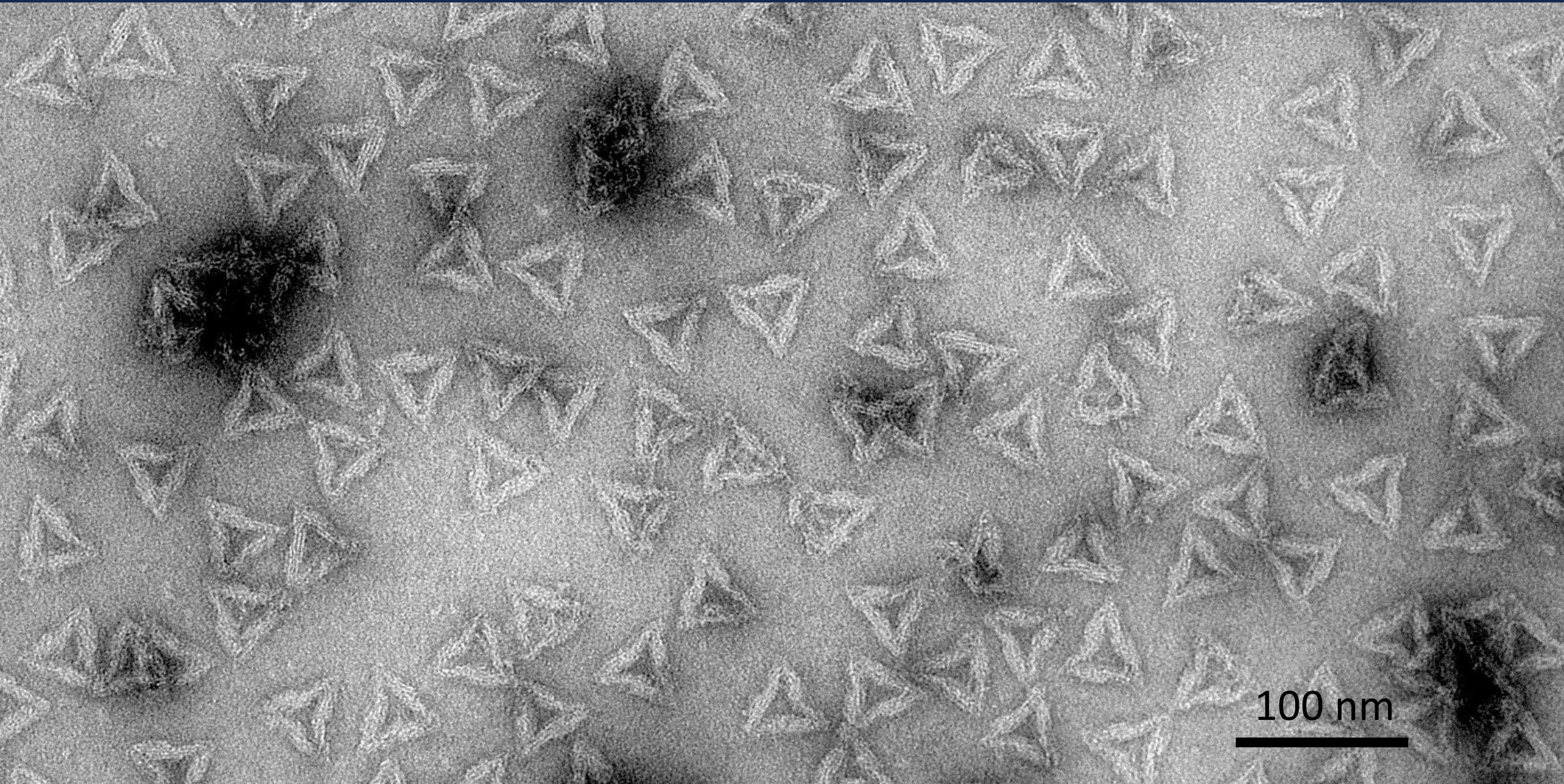
Paul W. K. Rothemund, Nature, 2006

DNA origami

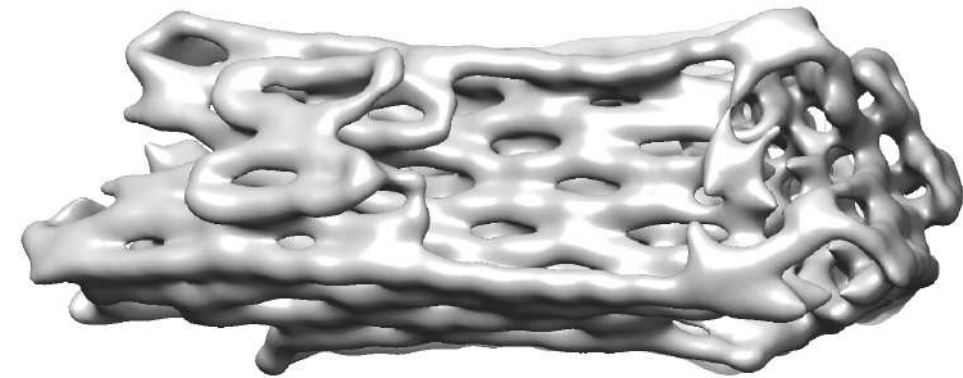
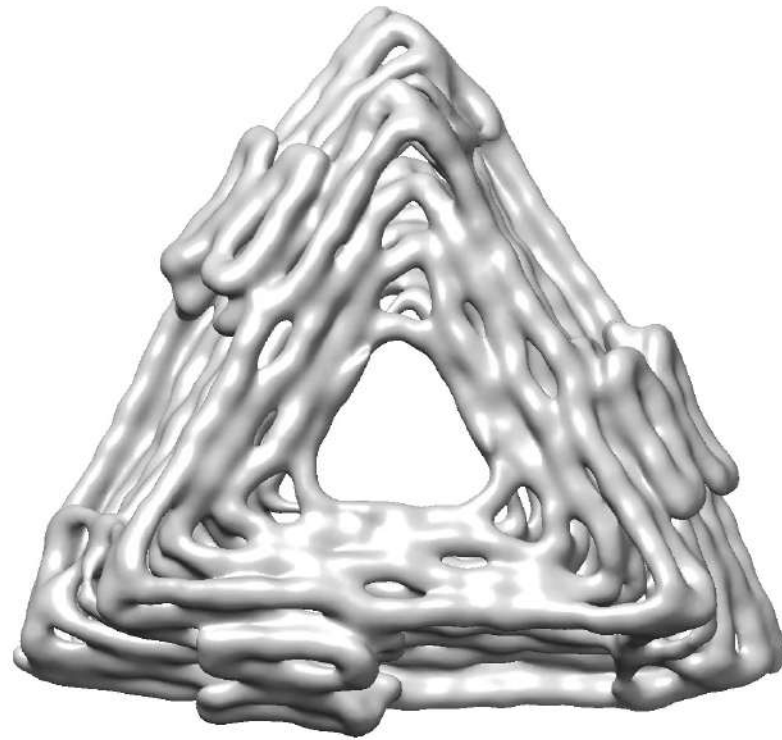
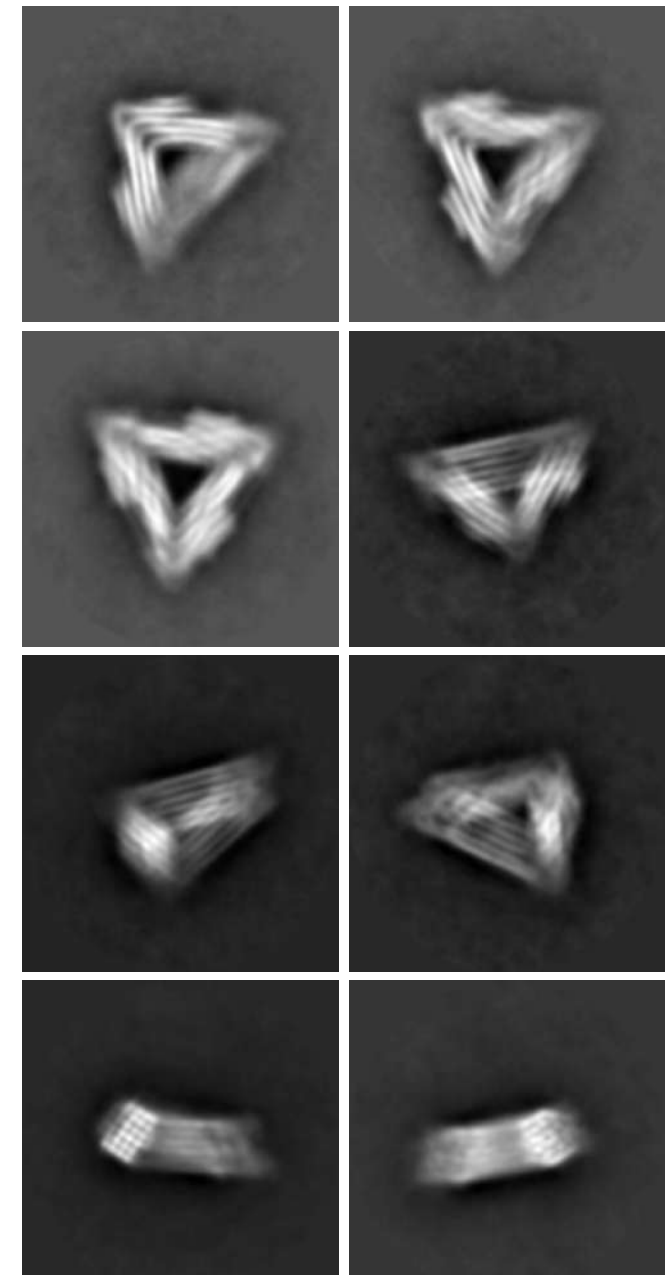


Paul W. K. Rothemund, Nature, 2006

Origami triangles as building blocks



Origami triangles as building blocks



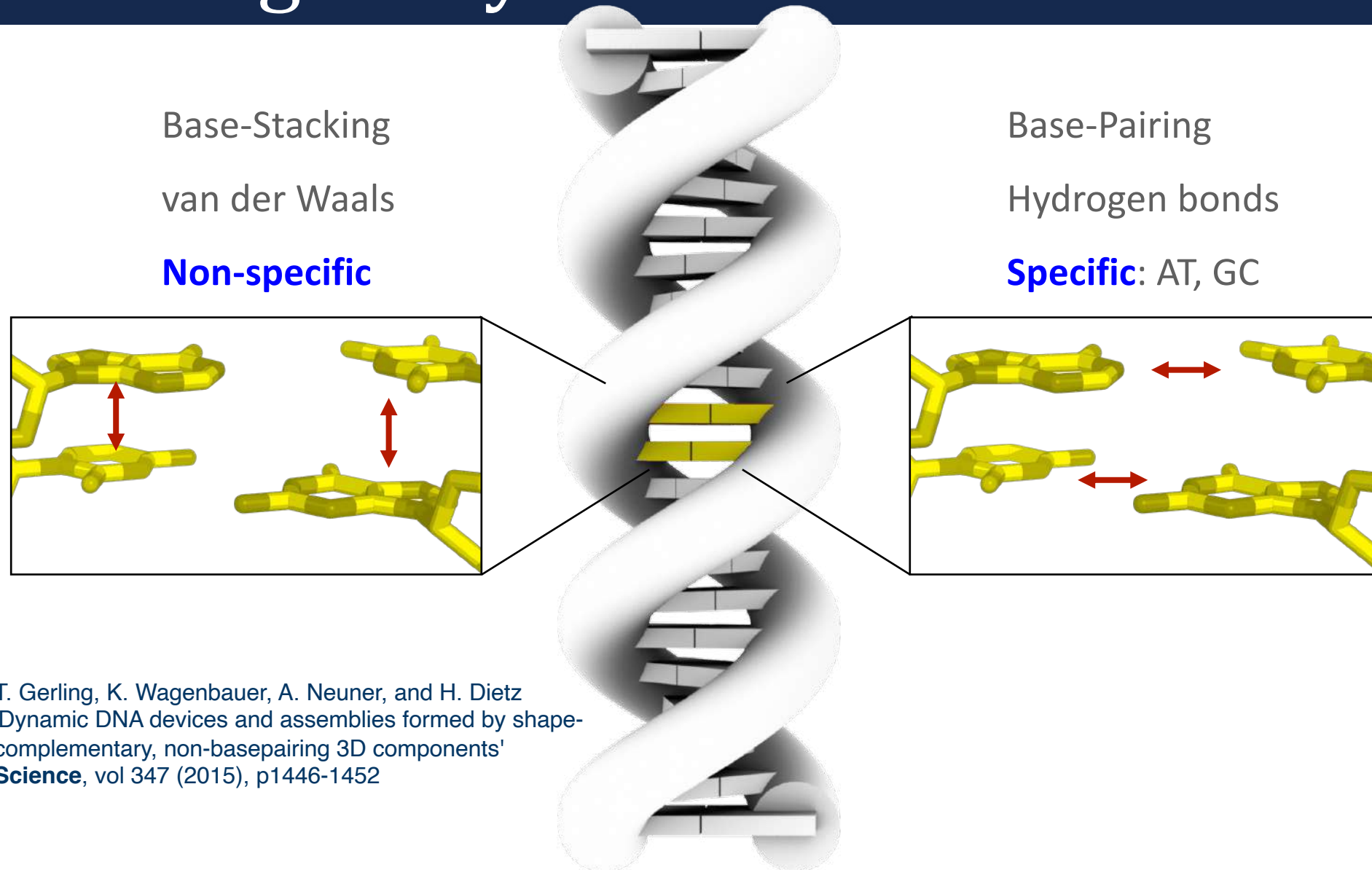
60 nm

Cryo electron microscopy, 2 nm resolution

Octahedral capsids from origami



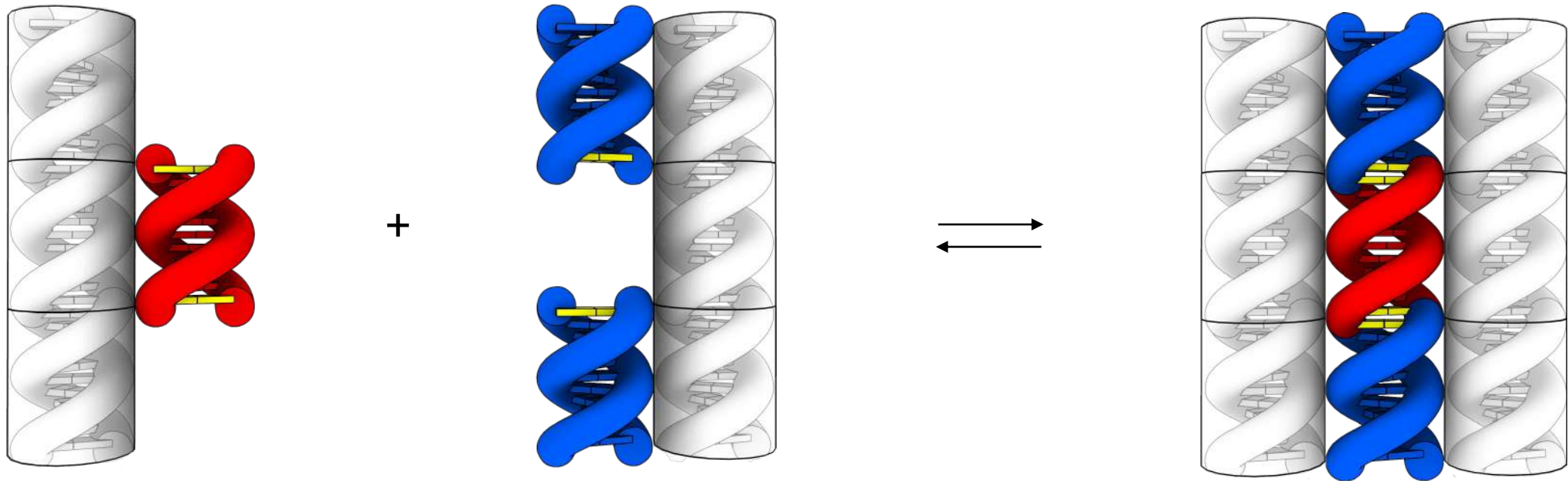
Base Stacking vs Hybridization



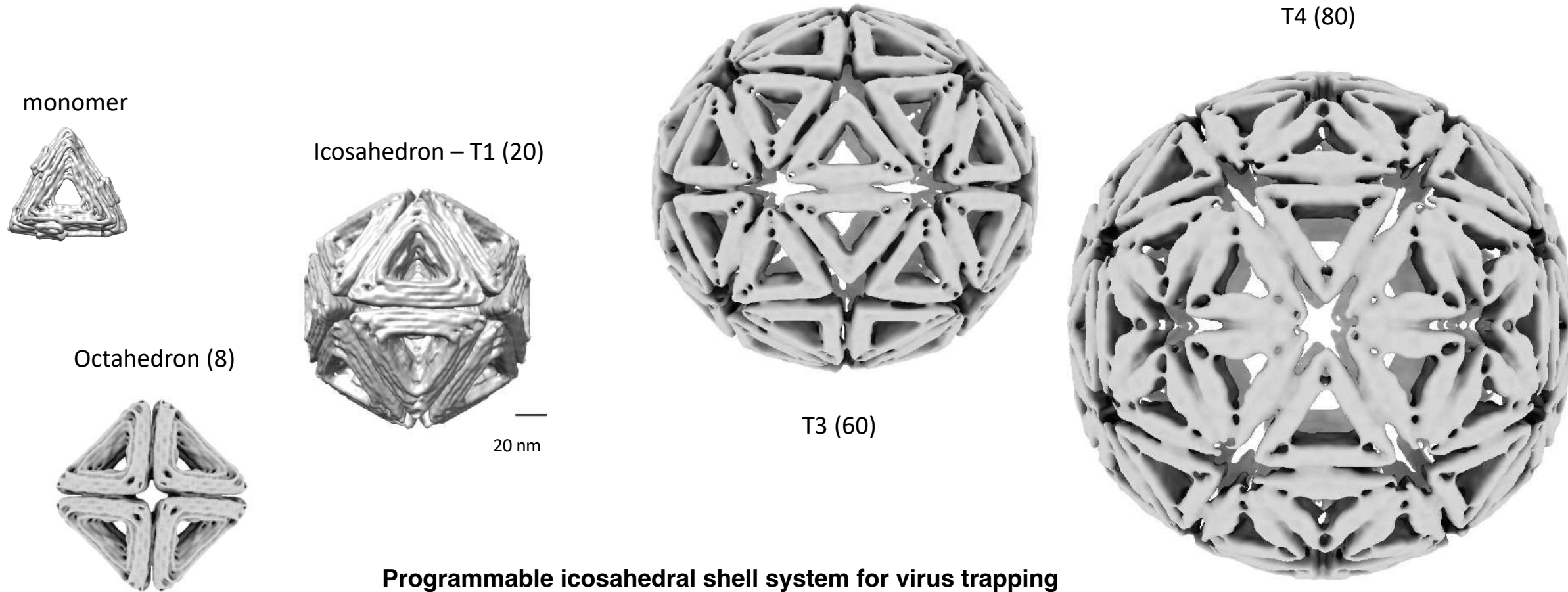
T. Gerling, K. Wagenbauer, A. Neuner, and H. Dietz
'Dynamic DNA devices and assemblies formed by shape-complementary, non-basepairing 3D components'
Science, vol 347 (2015), p1446-1452

Shape-Complementarity & Base-Stacking

Shape complementarity (lock and key)
restores specificity



Origami capsids



Programmable icosahedral shell system for virus trapping

Sigl, ..., Hagan, Fraden and Dietz, *Nature Materials* (2021)

DOI: [10.1038/s41563-021-01020-4](https://doi.org/10.1038/s41563-021-01020-4)

Geometrically programmed self-limited assembly of tubules using DNA origami colloids

Hayakawa, Videbæk, Hall, Fang, Sigl, Feigl, Dietz, Fraden, Hagan, Grason, Rogers, *PNAS* (2022)

<https://doi.org/10.1073/pnas.2207902119>



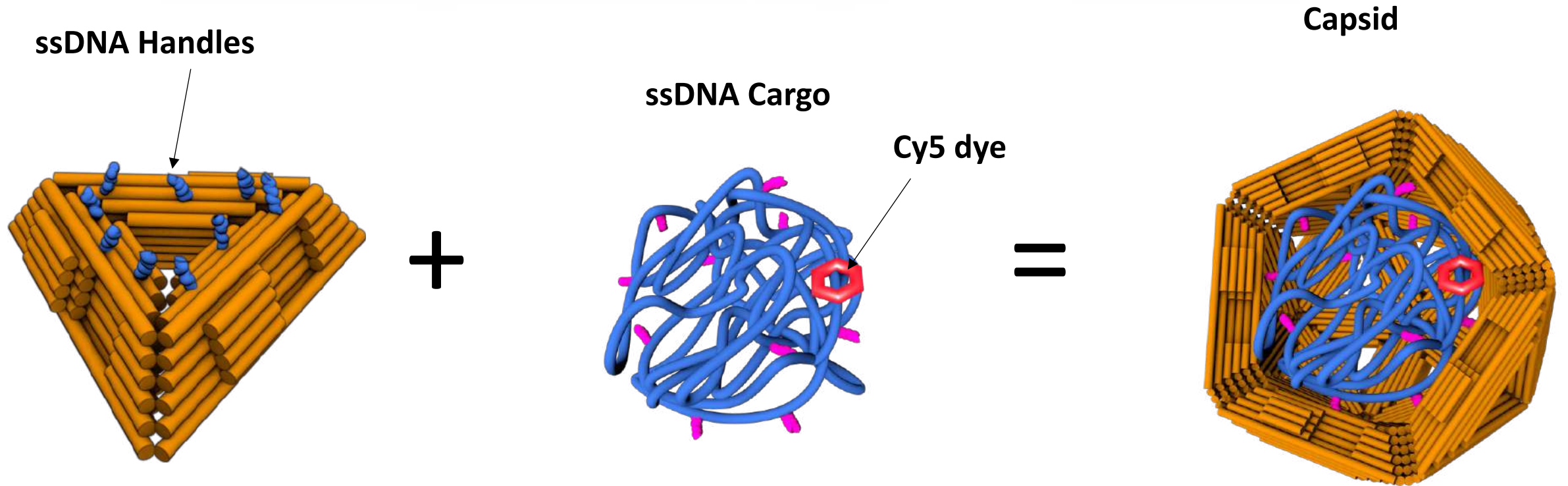
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encapsulation scaffolding

Cargo

Encapsulating a scaffold as cargo in a T1 icosahedron



Ali Aghvami

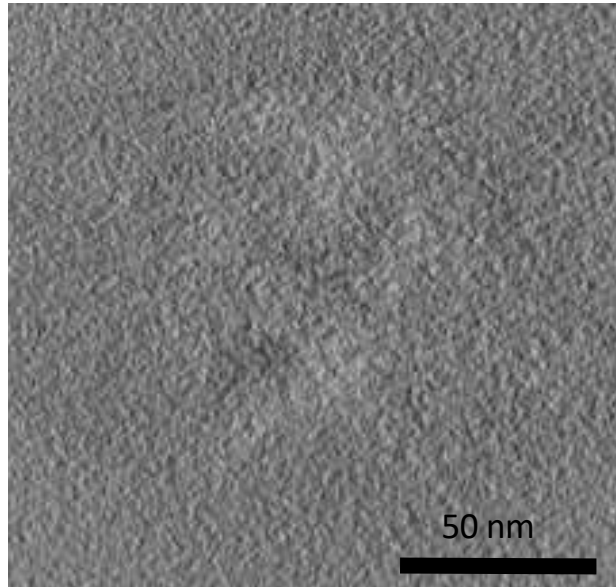
Elrad O.M., & Hagan M.F.
Physical biology (2010)

8064 bp scaffold with 10 handles

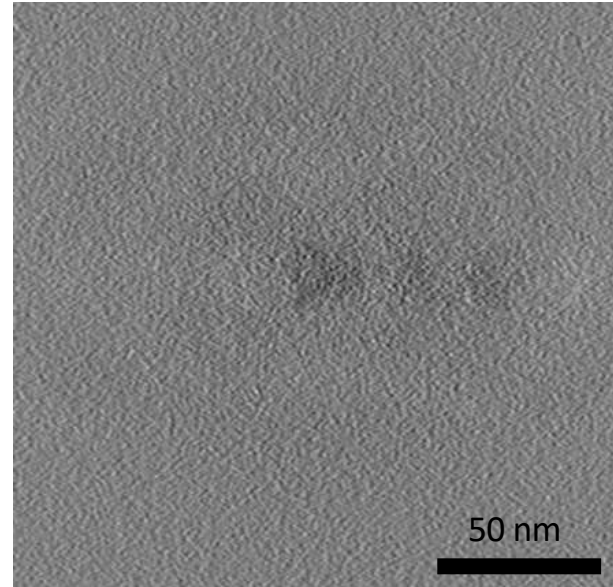
Complementary handles on scaffold and monomers

Cargo: TEM tomograms of encapsulated cargo in T1 capsids

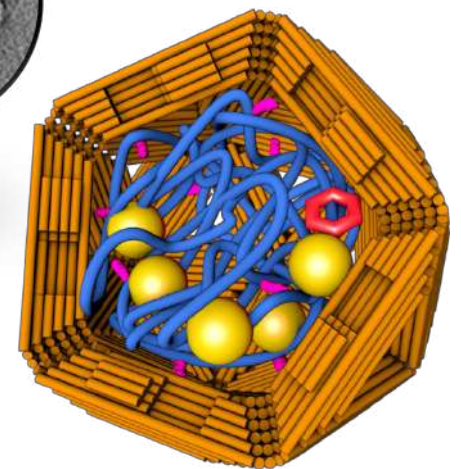
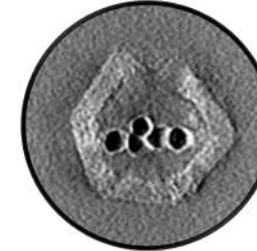
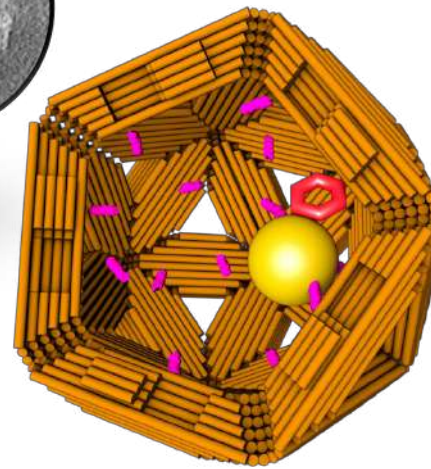
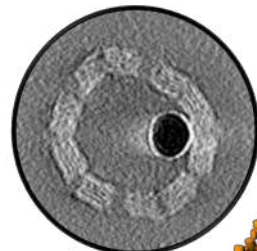
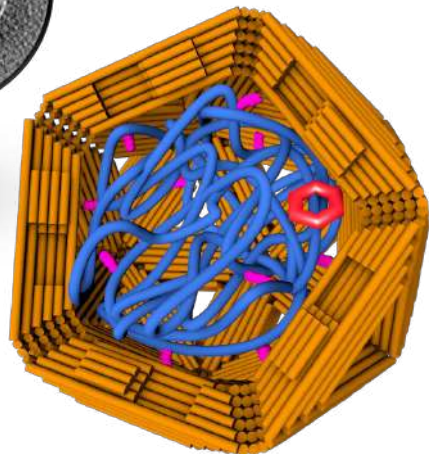
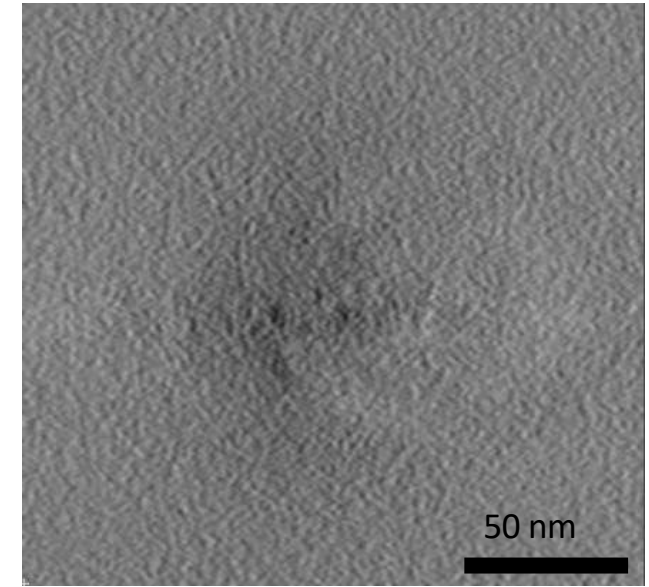
Scaffold



Gold Nano-particles

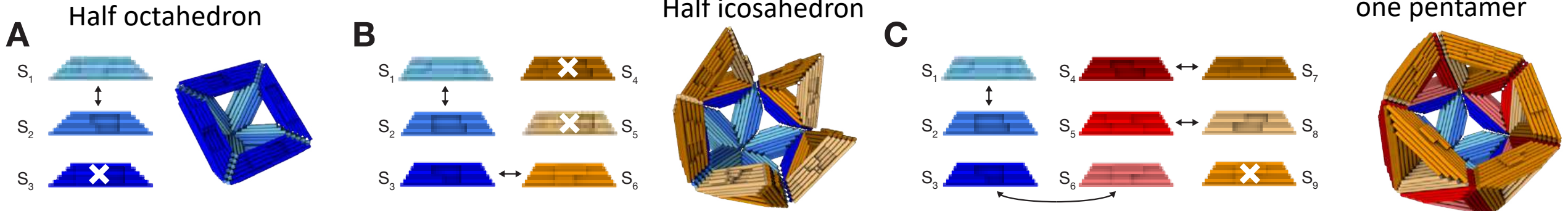


Scaffold & Gold n.p.



Virus deactivation

DNA origami capsids for virus deactivation: hepatitis B virus core



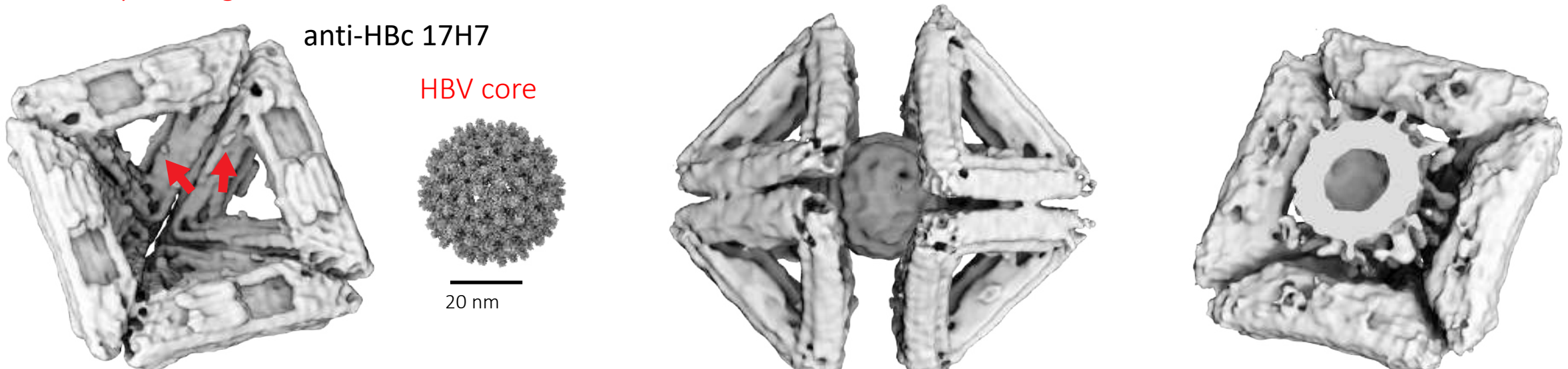
DNA origami capsids **capture** hepatitis B virus core

antibody binding site

anti-HBc 17H7

HBV core

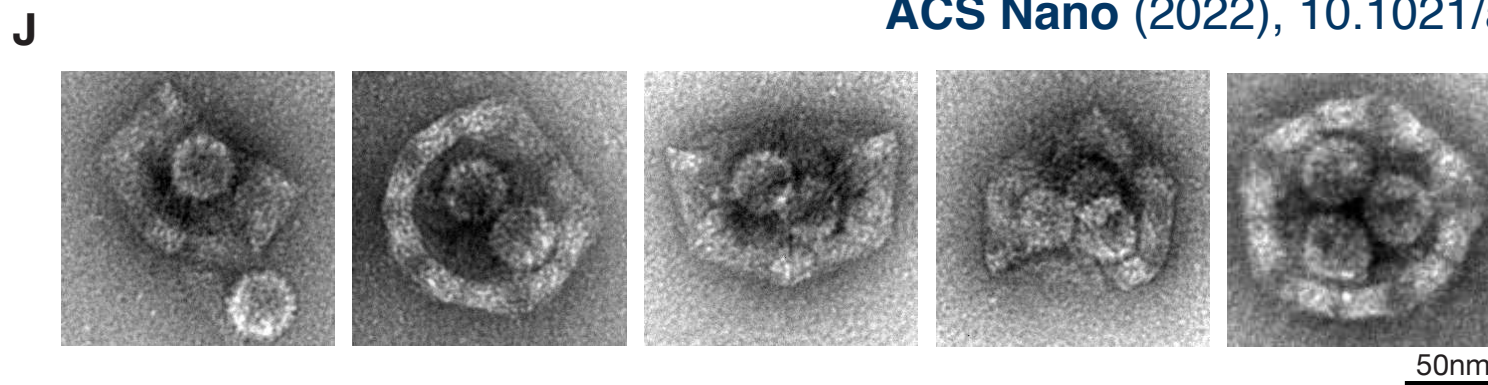
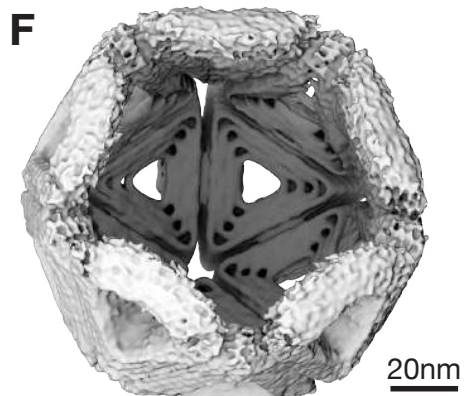
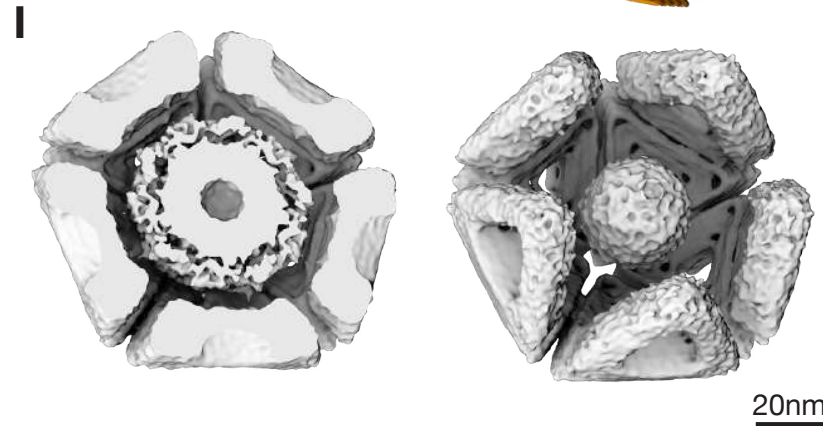
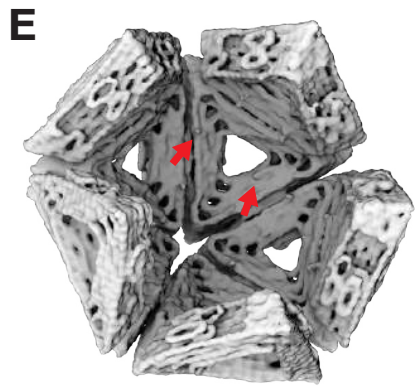
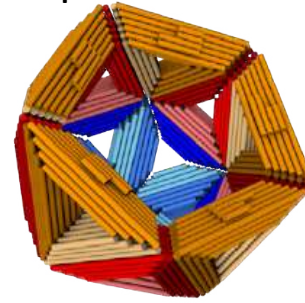
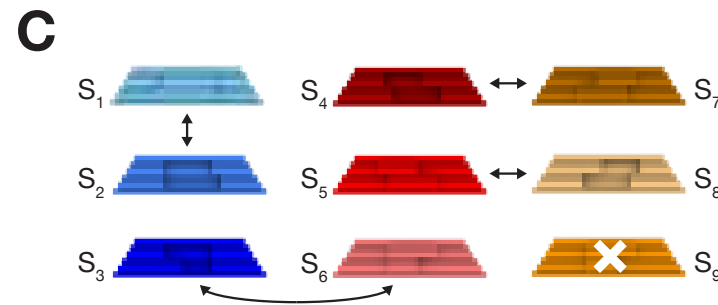
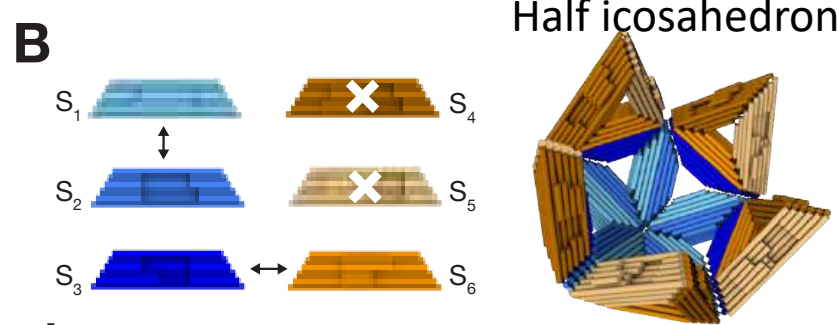
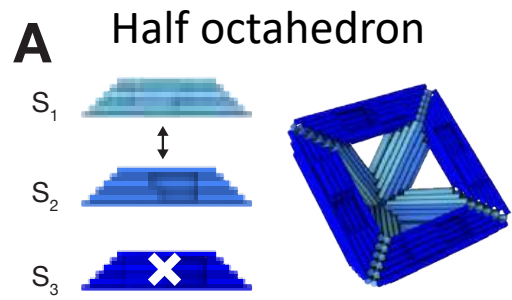
20 nm



Exploit viral assembly principles to deactivate virus

DNA origami capsids **capture** hepatitis B virus core

Icosahedron missing one pentamer

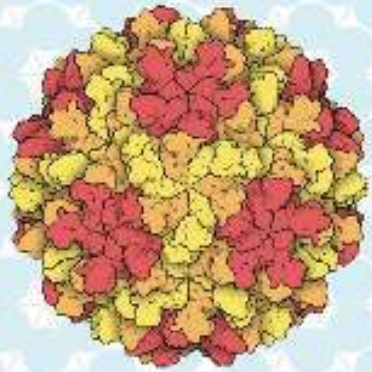


Programmable icosahedral shell system for virus trapping
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Hendrik Dietz 
virofight.eu

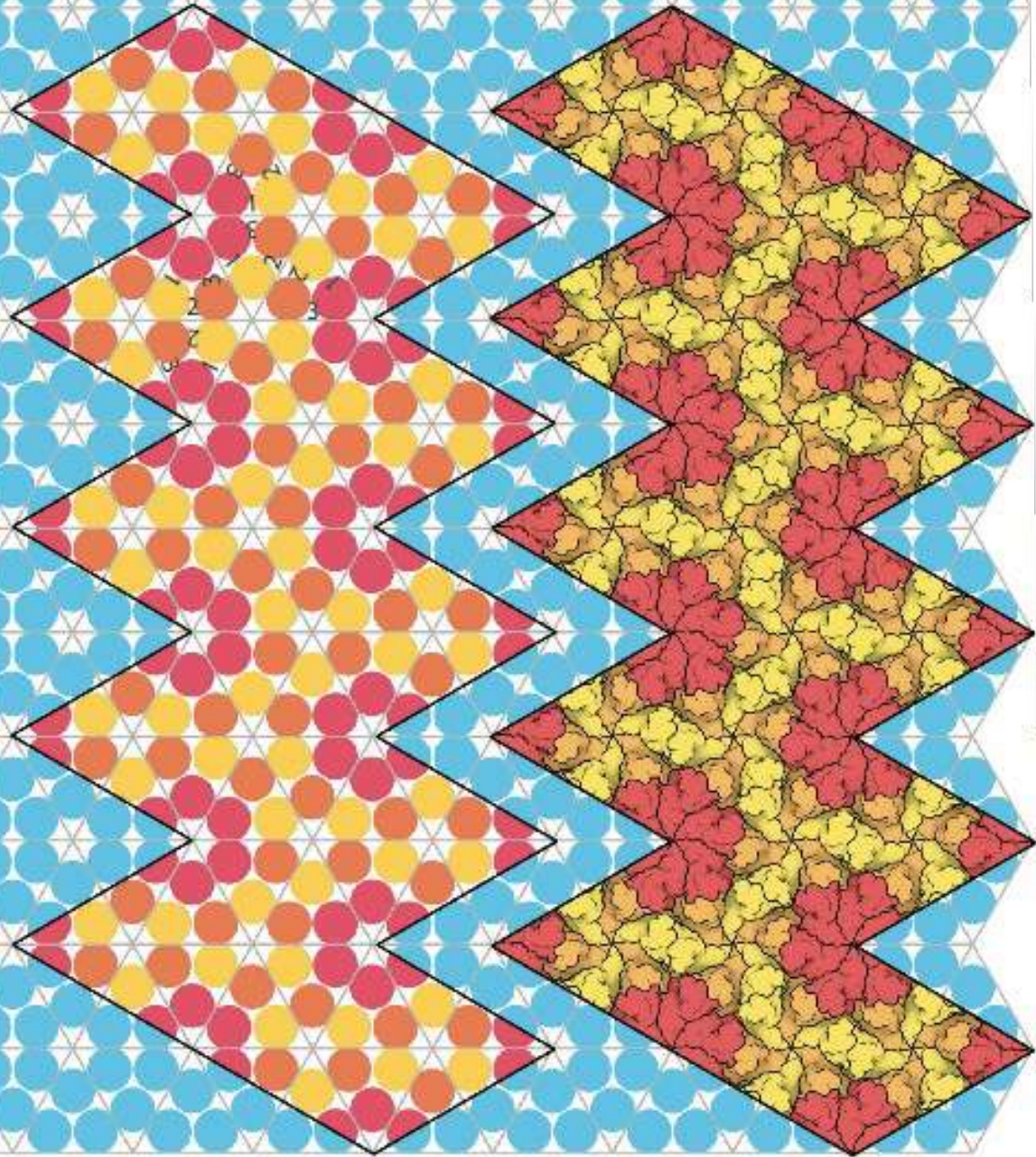
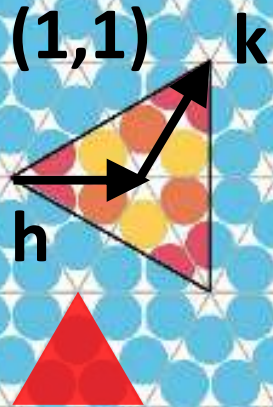
ACS Nano (2022), [10.1021/acsnano.1c11328](https://doi.org/10.1021/acsnano.1c11328)

T=3

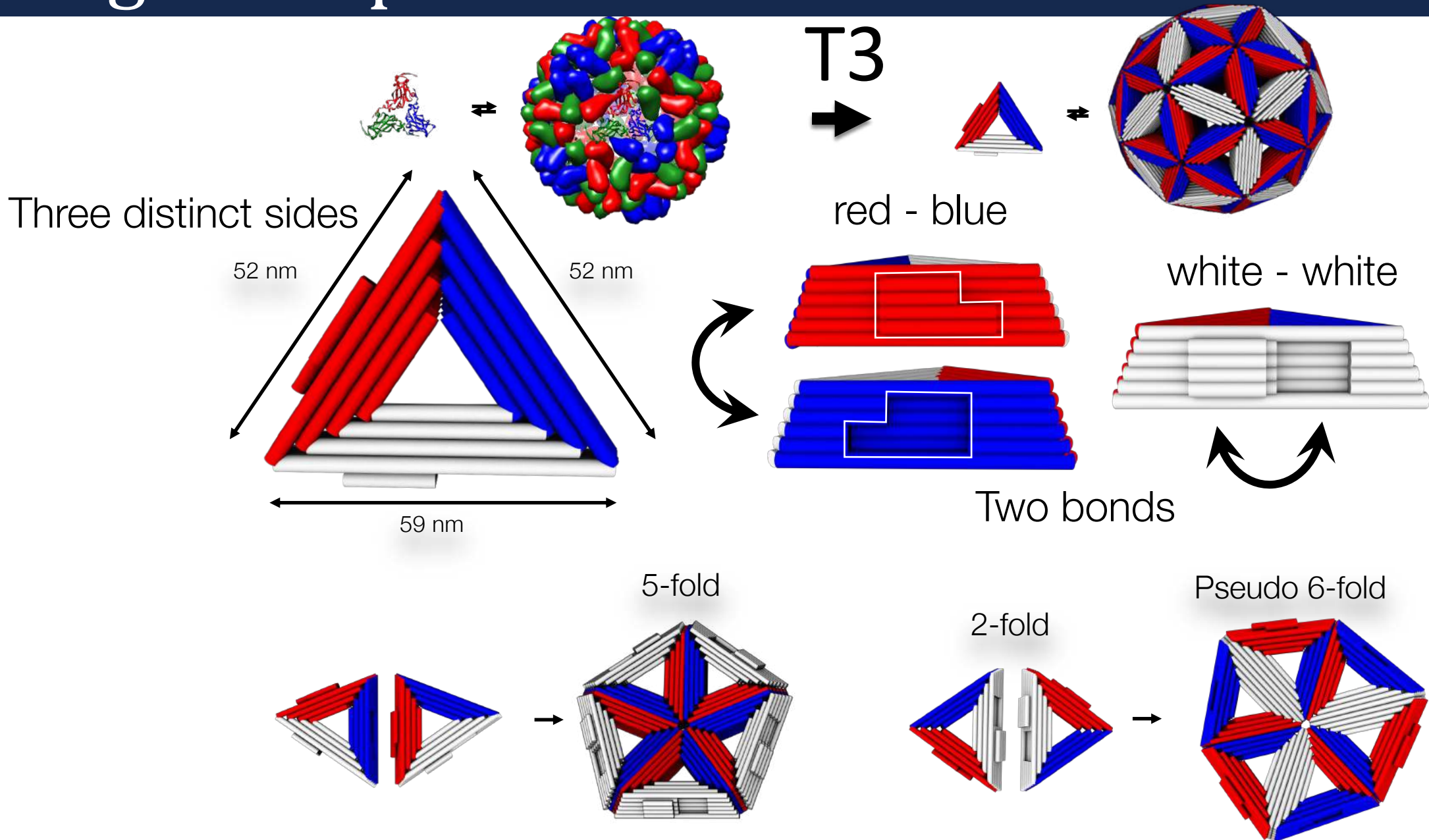


T=3
Tomato Bushy Stunt Virus
PDB entry: 2tbv

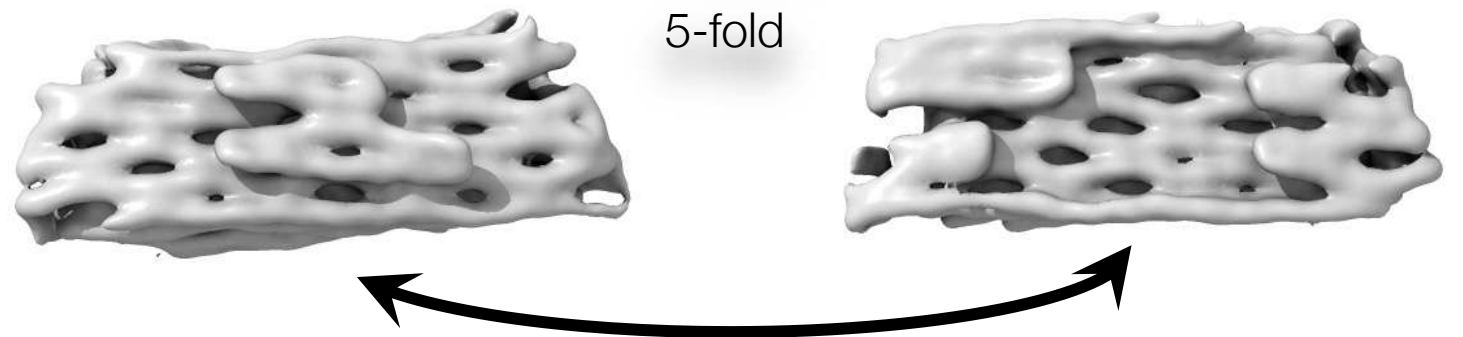
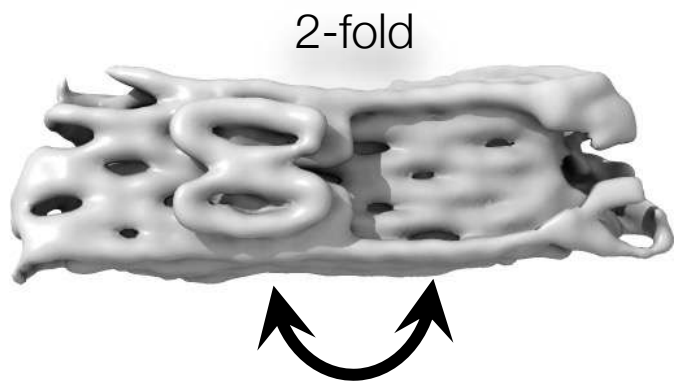
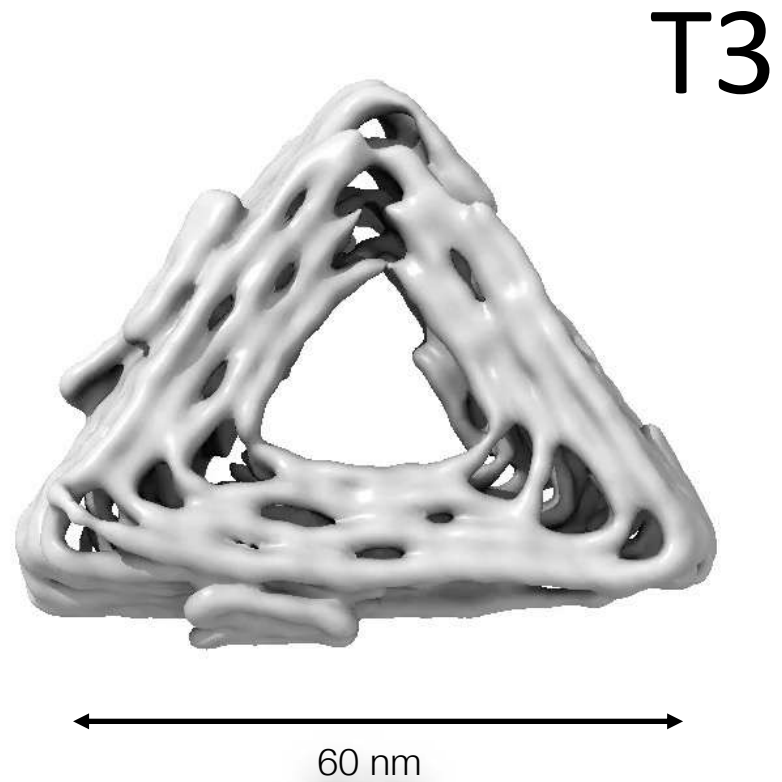
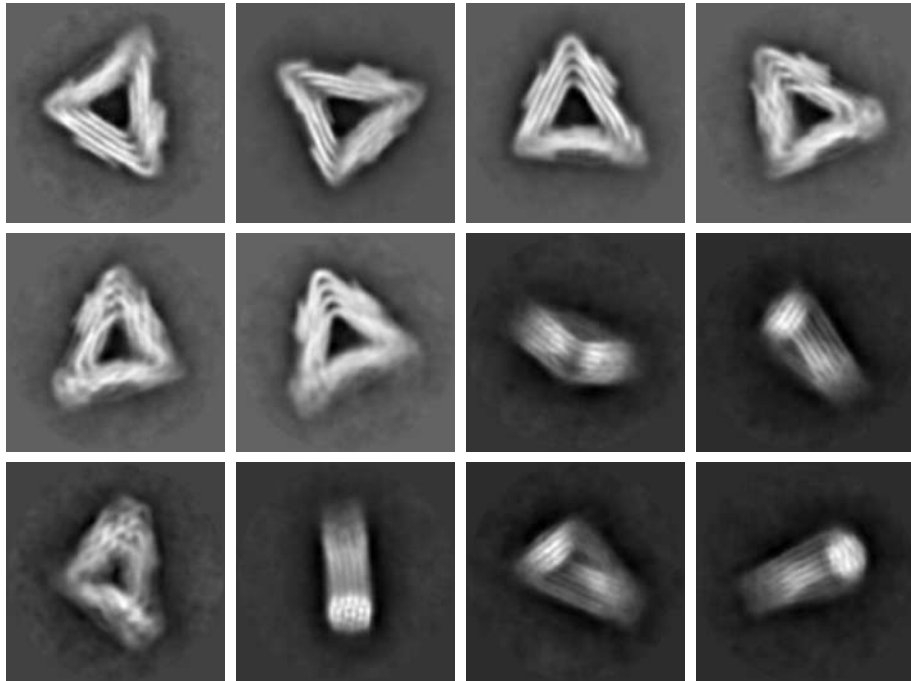
Subunits occupy three different positions: one type forms a five-fold interaction at the vertices of the icosahedron (shown in red), and two other types form a pseudo-six-fold interaction in the center of each icosahedral face (shown in yellow and orange).



Origami capsids

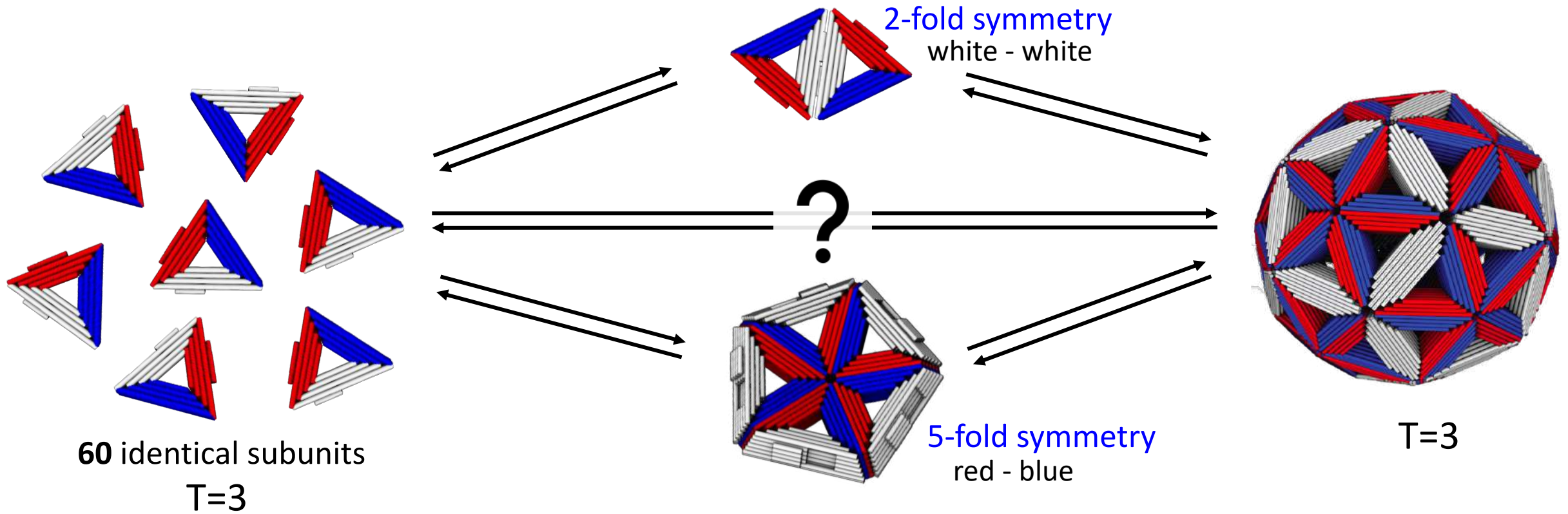


Origami capsids



What are the T=3 assembly pathways?

Symmetry is only one piece of the puzzle.



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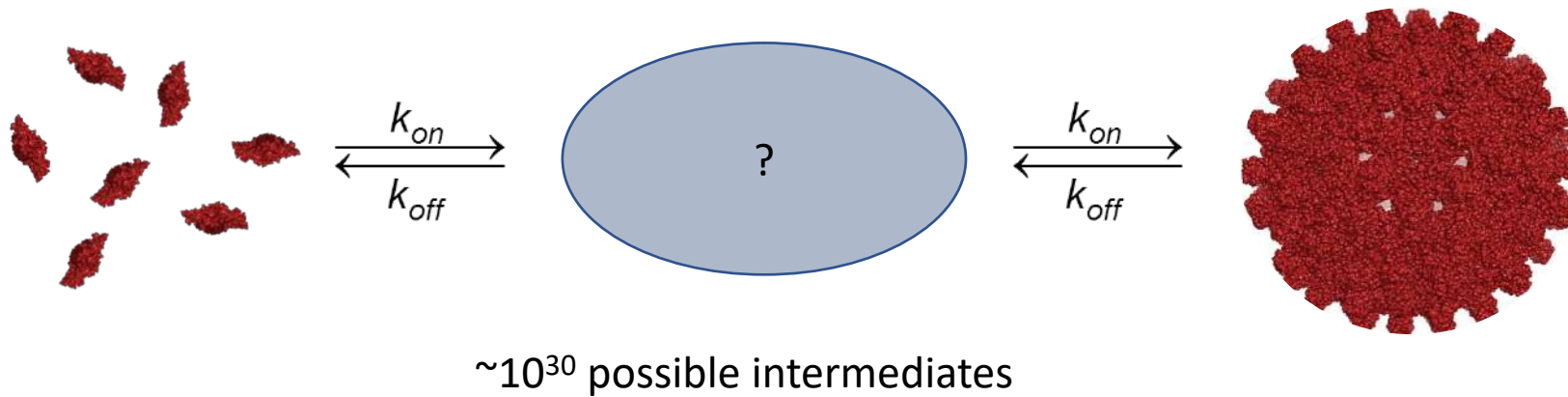
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Assembly pathways?

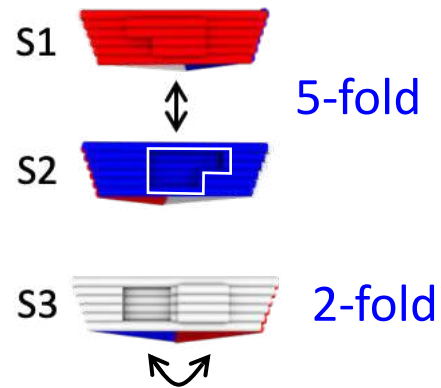
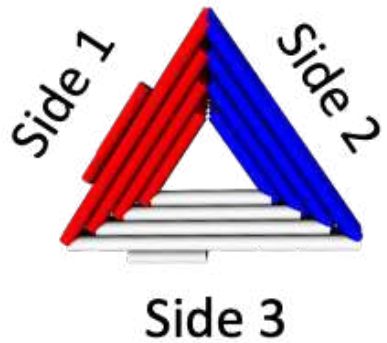
- alleviate errors
 - follow the most efficient path
 - avoid kinetic traps
- Weak protein-protein interactions, slow nucleation, reversible association



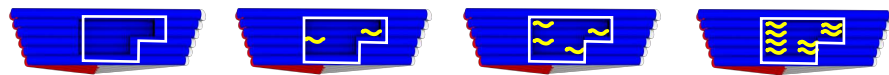
Symmetry is only one piece of the puzzle

Question: What are the optimal kinetic pathways?

From symmetry

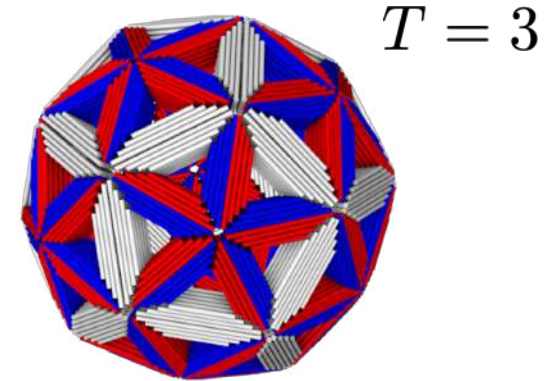


But what about the relative strength of the two interactions?



Increase # complementary ssDNA

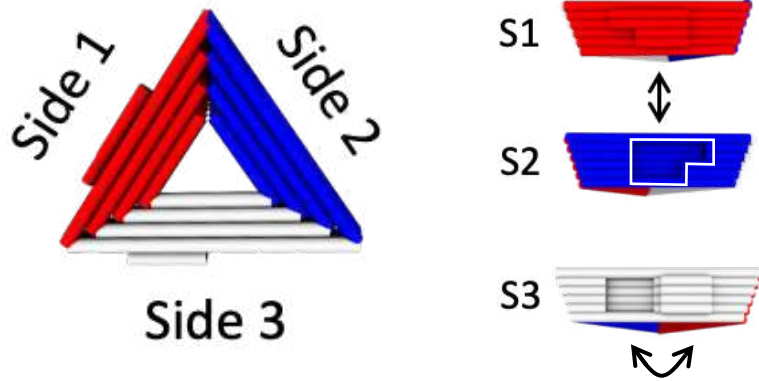
Binding strength ↑



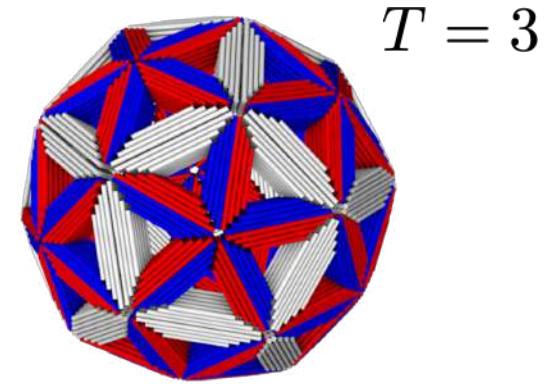
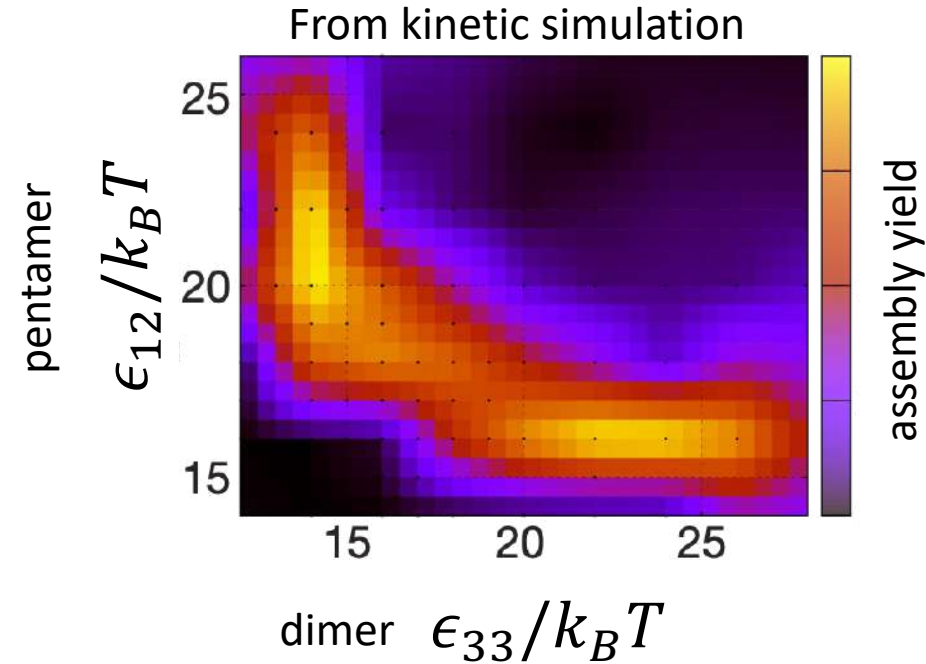
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From symmetry



But what about the relative strength of the two interactions?



**MONDAY 13 NOVEMBER 2023**

09.00 - 09.50 **Registration and Refreshments**

09.50 - 10.00 **Welcome and Housekeeping**

10.00 - 10.30 **William Gelbart, UCLA** IN CELLULO versus IN VITRO RECONSTITUTION OF RNA-SPECIFIC VIRUS-LIKE PARTICLES [Abstract](#)

10.30 - 11.00 **Dek Woolfson, University of Bristol** Rationally and computational design of functional protein barrels [Abstract](#)

11.00 - 11.30 **Farzad Fatehi, University of York** Geometry as a key to the virosphere – from viral infections to virus nanotechnology [Abstract](#)

11.30 - 12.00 **Refreshments**

12.00 - 12.30 **Roya Zandi, University of CA, Riverside** The role of flexibility of capsid proteins in the symmetry of viral shells [Abstract](#)

12.30 - 13.00 **Danielle Tullman-Ercek, Northwestern University** Designing with nanoscale building blocks: engineering self-assembling protein superstructures for applications in vaccines, drug delivery and biochemical production [Abstract](#)

13.00 - 14.30 **Lunch**

14.30 - 15.00 **David Bhella, MRC - University of Glasgow Centre for Virus Research** Capsid stability and receptor mediated priming in calicivirus entry

15.00 - 15.30 **Alexander Borodavka, University of Cambridge** How do Viruses Use RNA Granules and RNA Chaperones to Assemble Their Segmented Genomes? [Abstract](#)

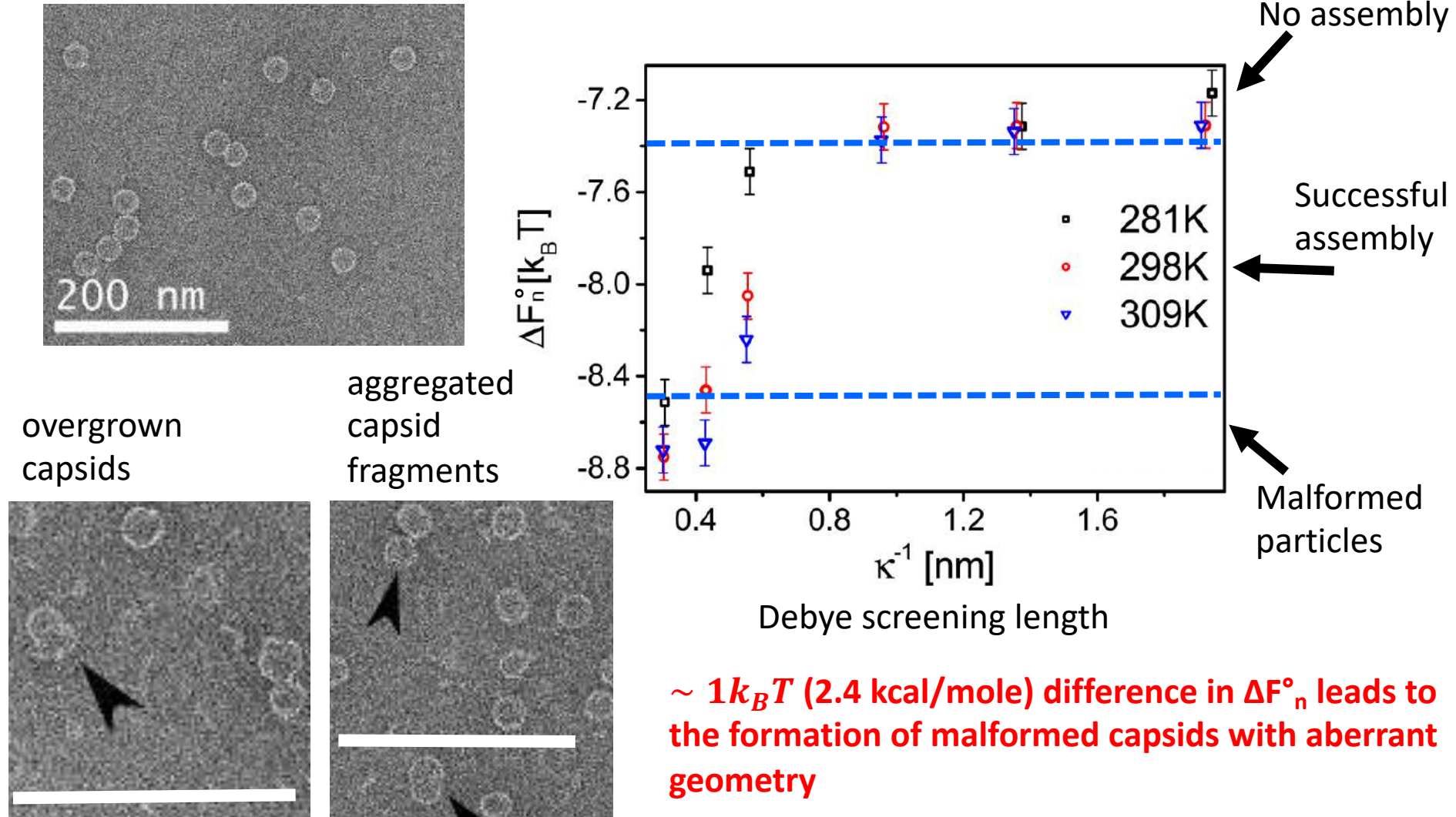
15.30 - 16.00 **Refreshments**

16.00 - 16.30 **Uri Raviv, The Hebrew University of Jerusalem** Mechanism of Virus Capsid Assembly [Abstract](#)

16.30 - 17.00 **Erwin Frey, Ludwig-Maximilians-Universitaet Muenchen** The time complexity of self-assembly [Abstract](#)

Thermodynamic analysis of assembly products

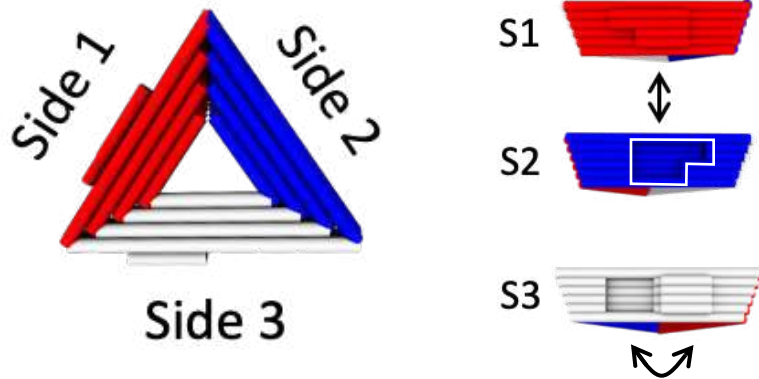
At $20\mu\text{M}$ protein successful assembly can be realized within a narrow range of association free energies ($7.4 - 8.5k_B T$)



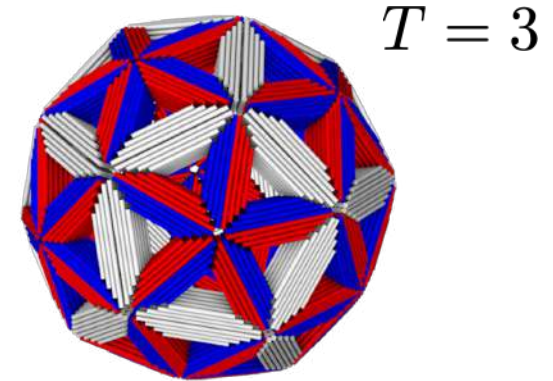
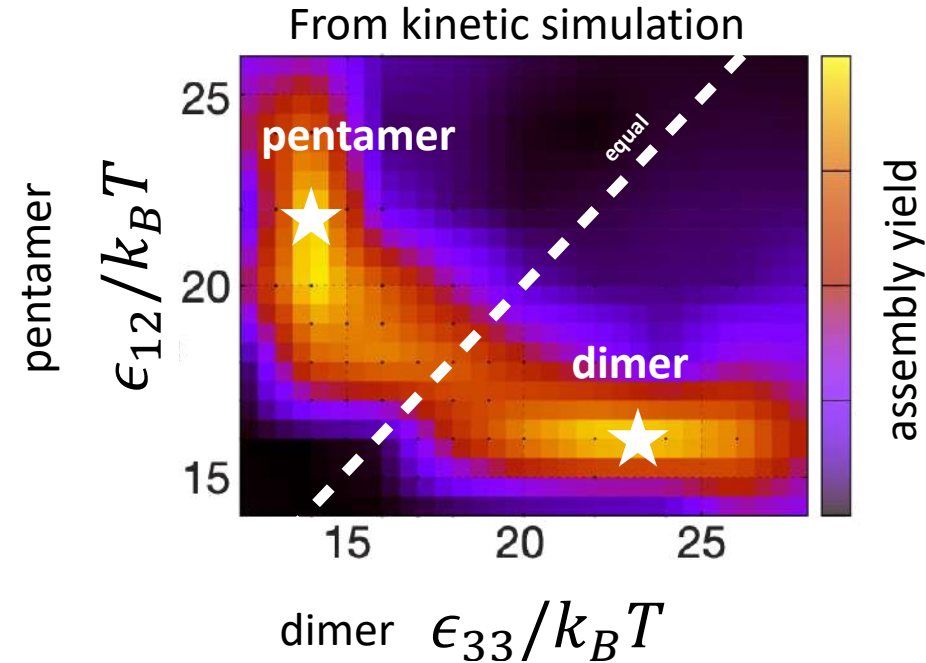
Symmetry is only one piece of the puzzle

Question: What are the optimal kinetic pathways?

From symmetry



But what about the relative strength of the two interactions?



Takeaway: Hierarchical pathways predicted* to improve assembly yield

predicted*: Before experiment!

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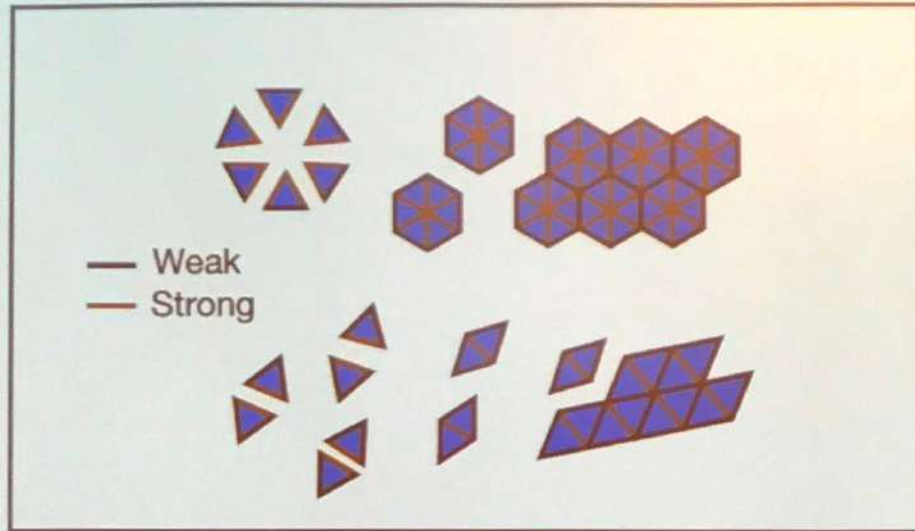
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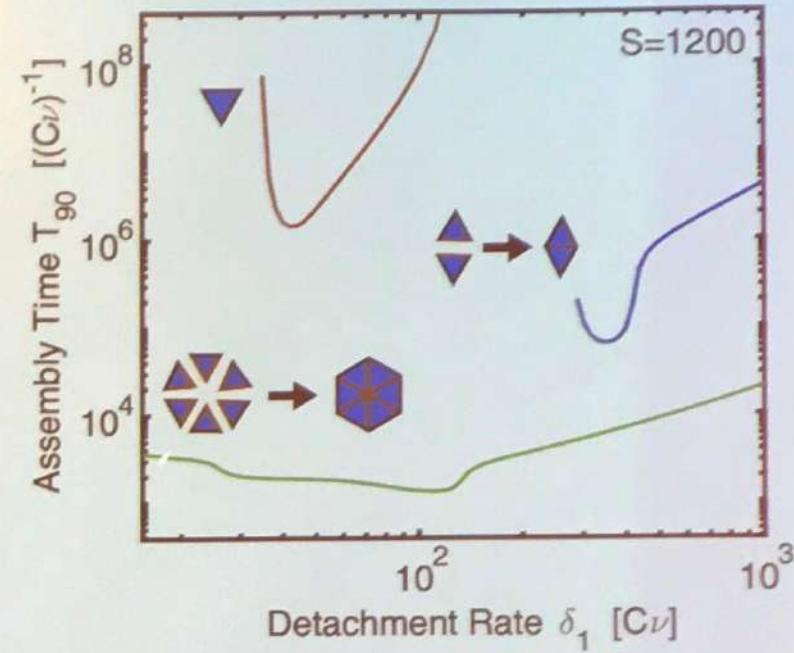
16.30 - 17.00 **Erwin Frey, Ludwig-Maximilians-Universitaet Muenchen** The time complexity of self-assembly [Abstract](#)

How can we make assembly robust?



Hierarchical self-assembly schemes

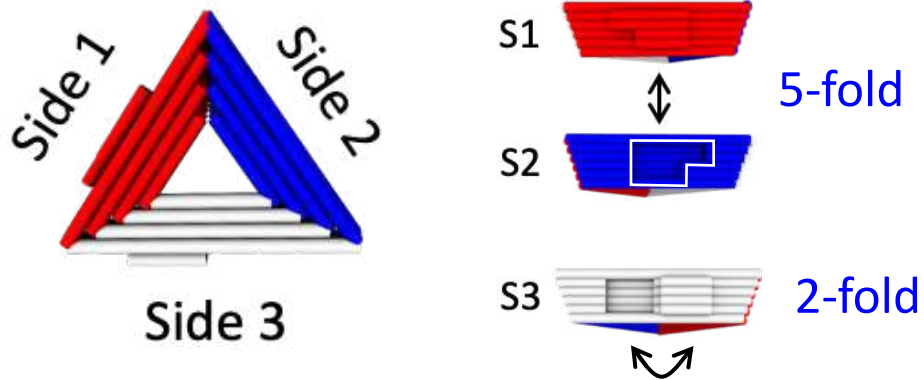
Make triangle-shaped building blocks form higher-order constituents



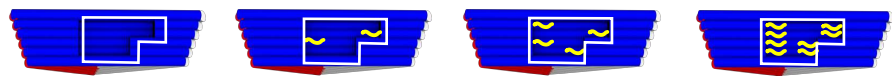
Tuned block-block interactions

What are the optimal kinetic pathways?

From symmetry



But what about the relative strength of the two interactions?

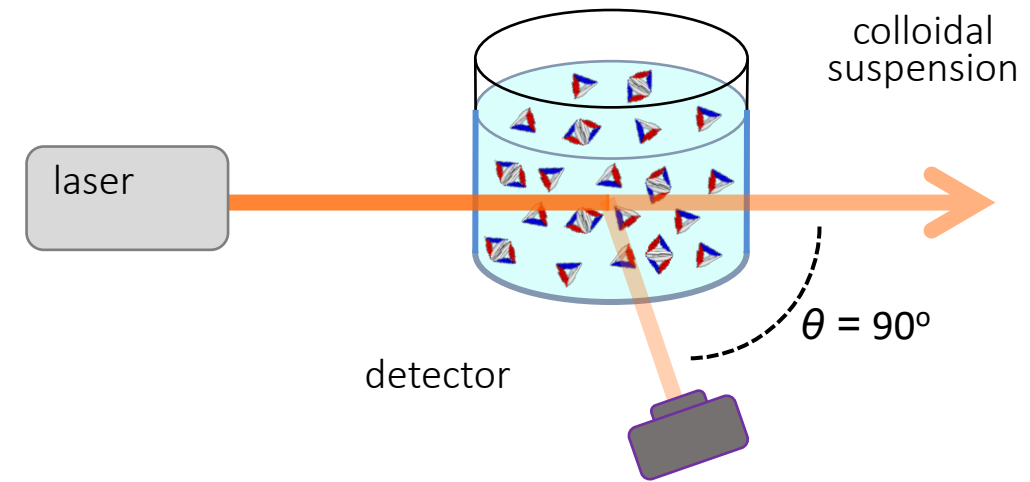


Increase # complementary ssDNA

Binding strength ↑

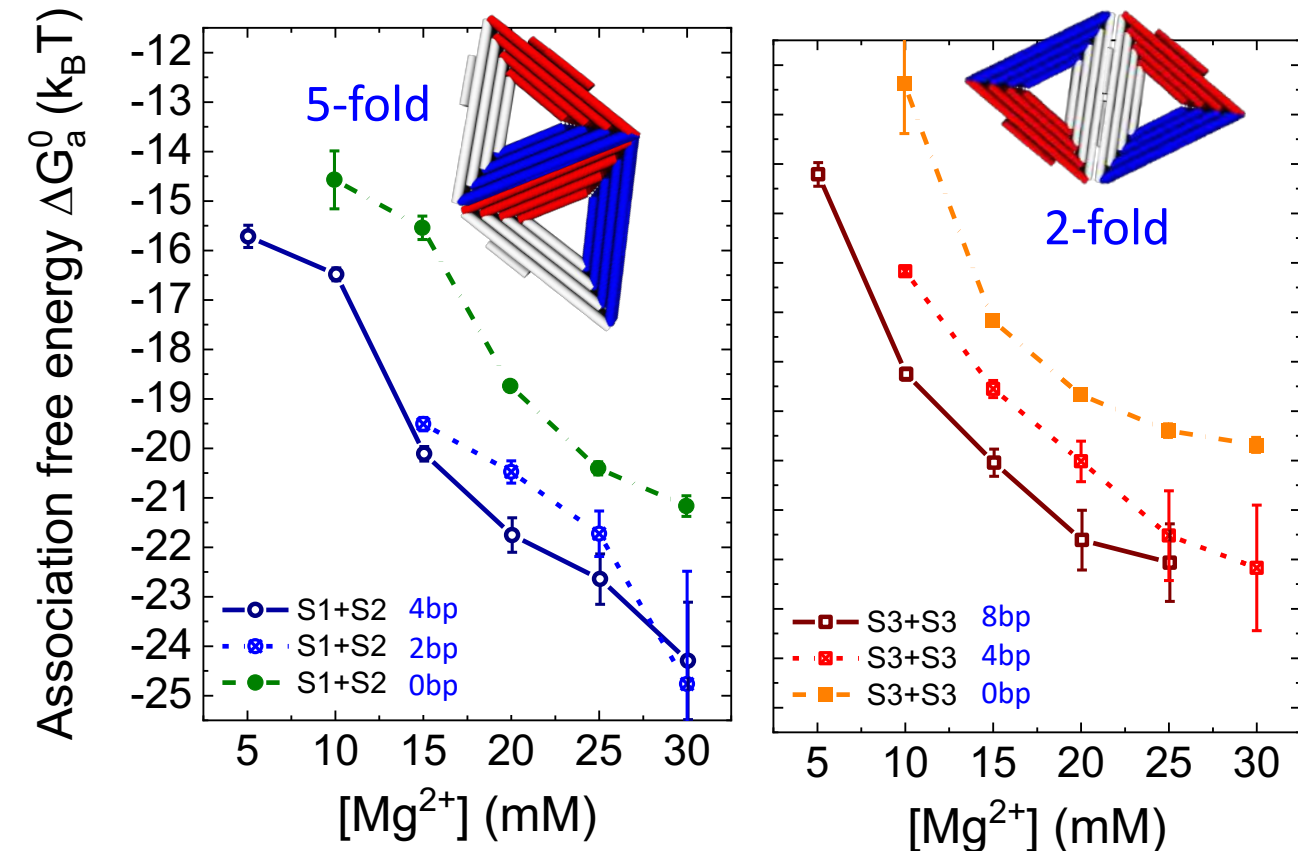
Free energy measurement

Static light scattering - **Gibbsometer**
& real time **assembly monitor**

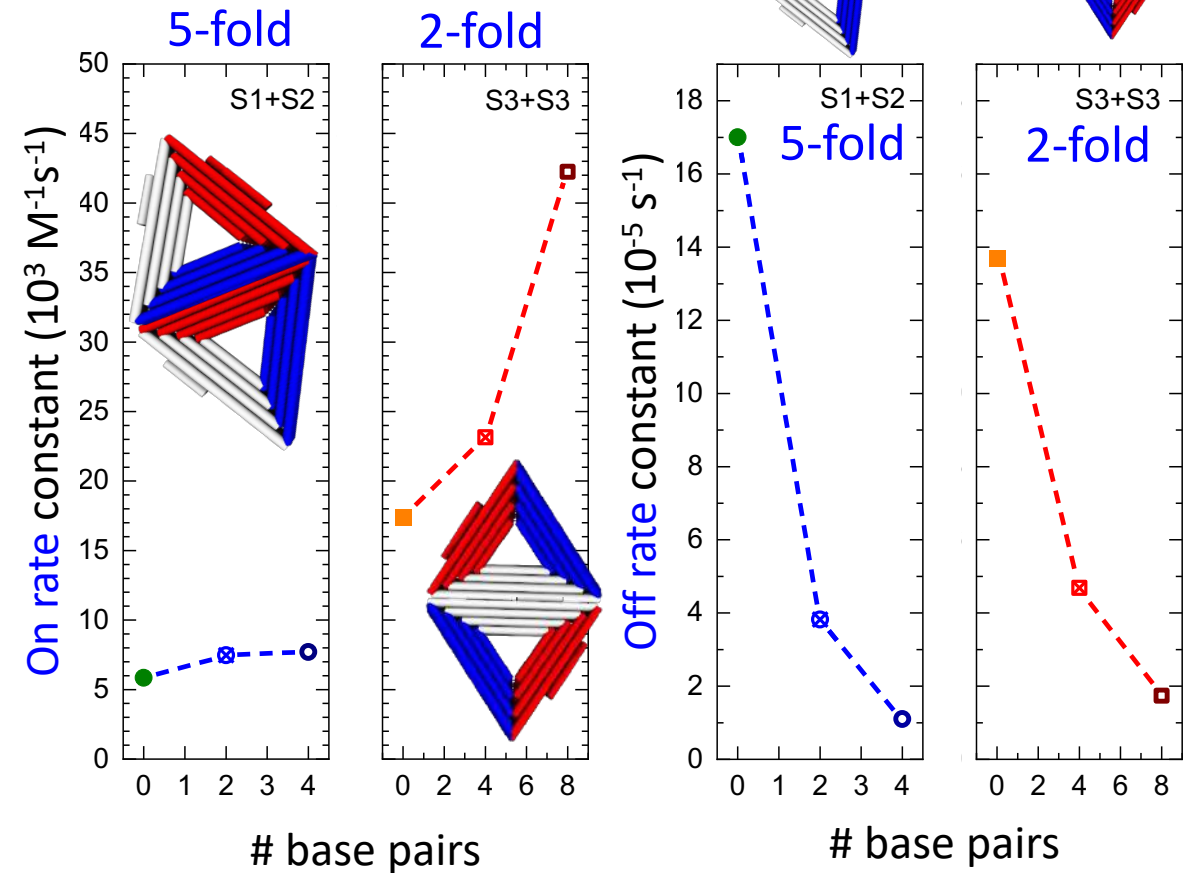


Monomer – dimer free energy

- (1) Bond strength **increases** with the number of added base pairs & the magnesium ion concentration.
- (2) Off rate constant **strongly** decreases with number of added base pairs.
- (3) On rate constant shows a **weak** dependence with number of added base pairs.
- (4) On rate 5 orders of magnitude slower than diffusion limited rate.



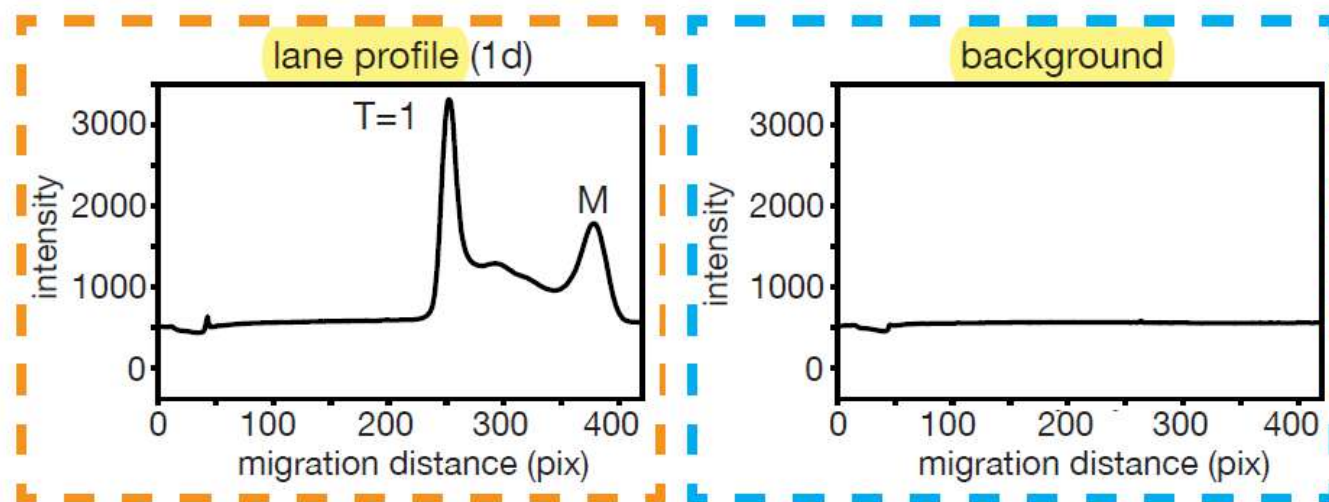
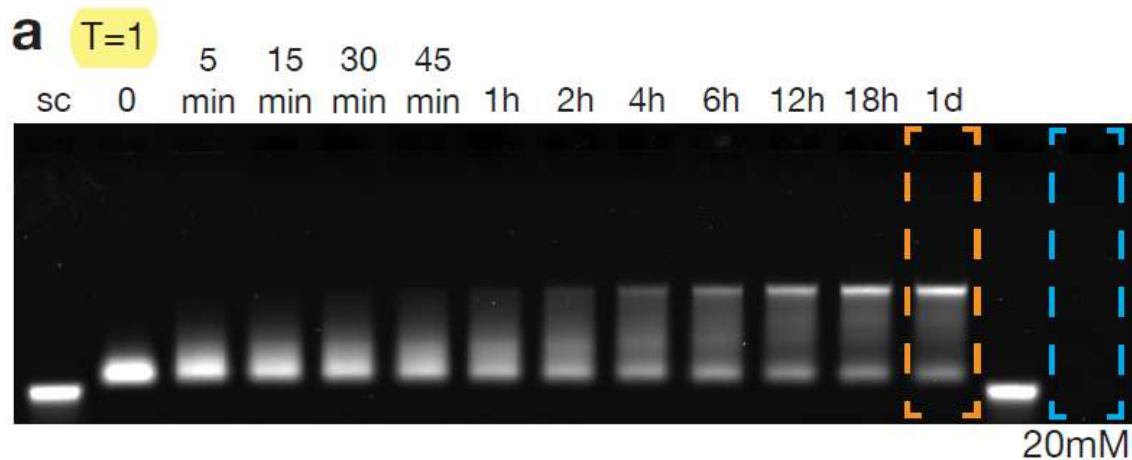
Gibbs free energy vs Mg^{2+} concentration



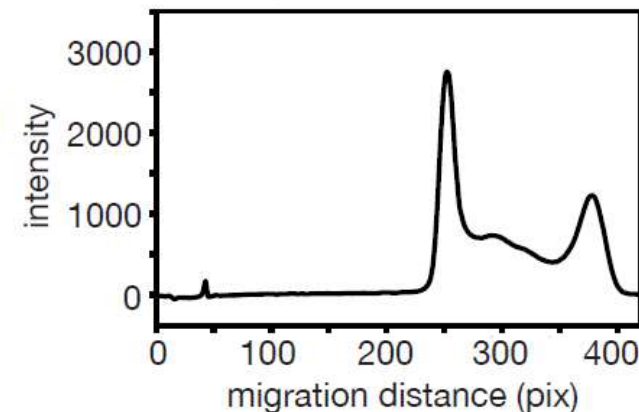
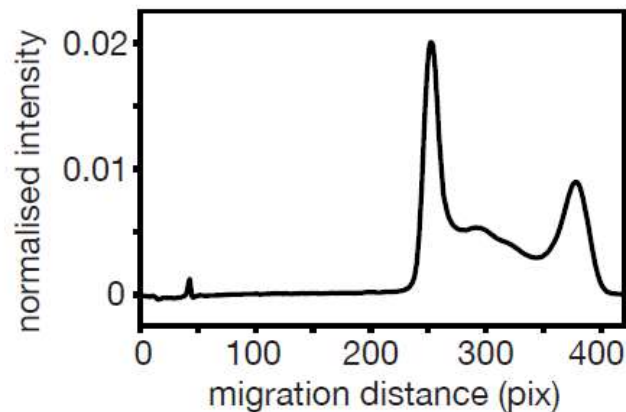
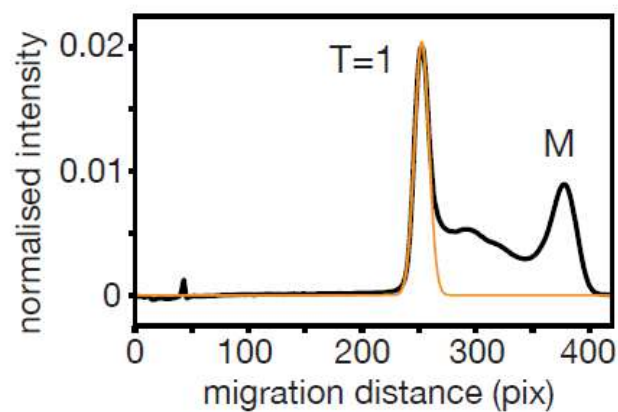
On and off rate constants vs. # base pairs (20 mM Mg^{2+})

Kinetics: Experiment

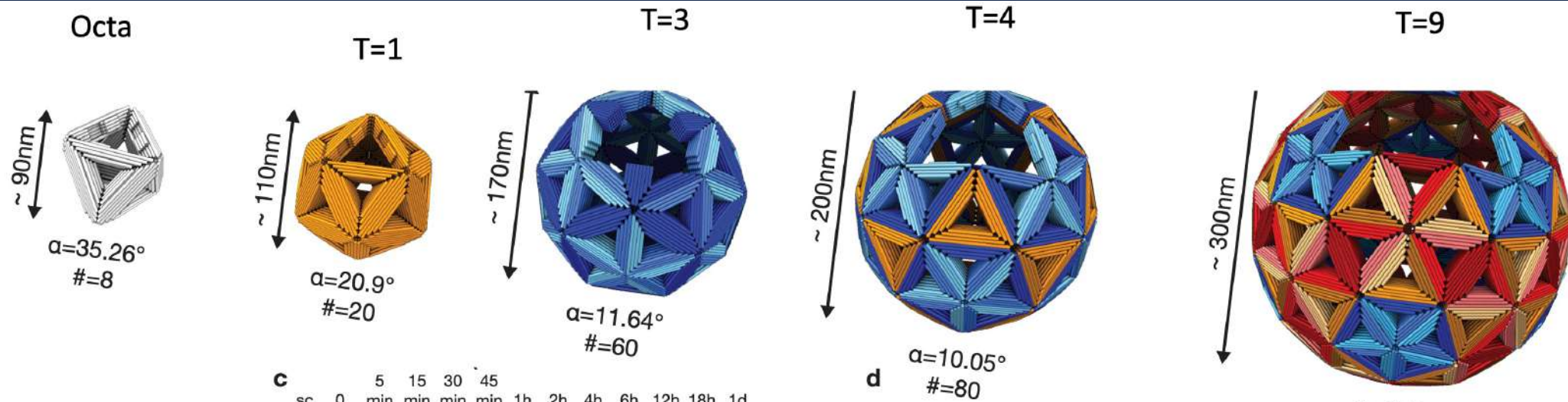
Gel Electrophoresis



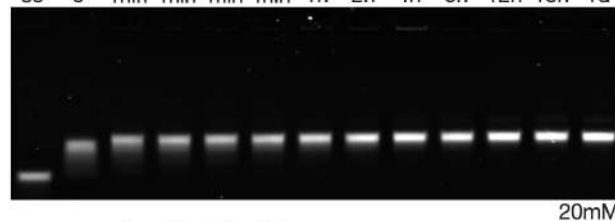
Capsid yield



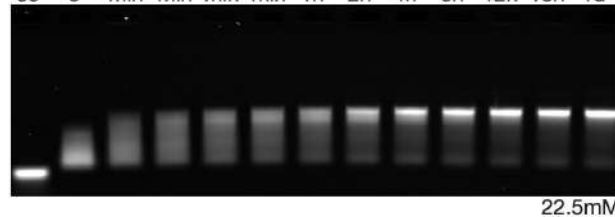
Kinetics: Experiment



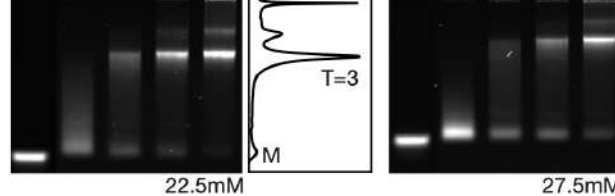
c 5 15 30 45
sc 0 min min min min 1h 2h 4h 6h 12h 18h 1d



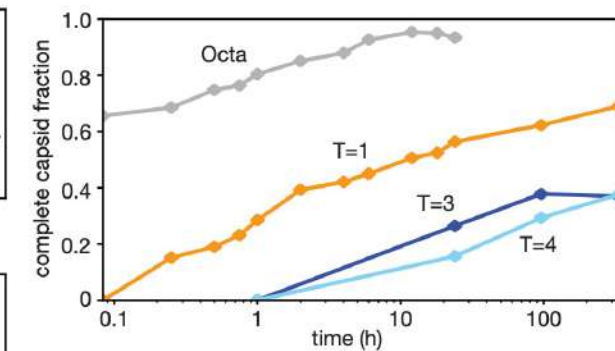
5 15 30 45
sc 0 min min min min 1h 2h 4h 6h 12h 18h 1d



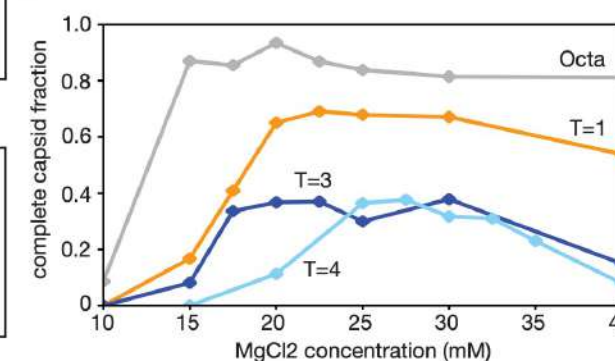
sc 1h 1d 4d 14d



d $\alpha=10.05^\circ$
 $\#=80$



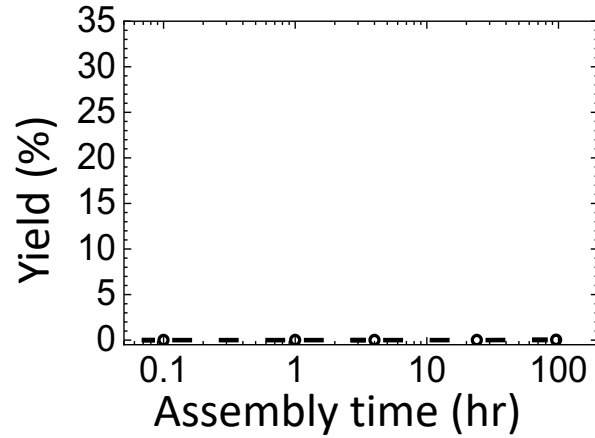
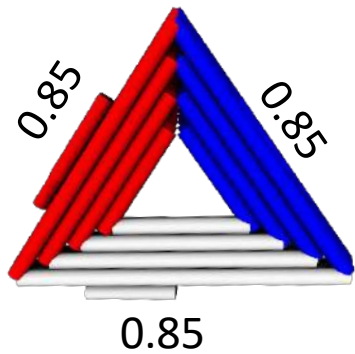
e



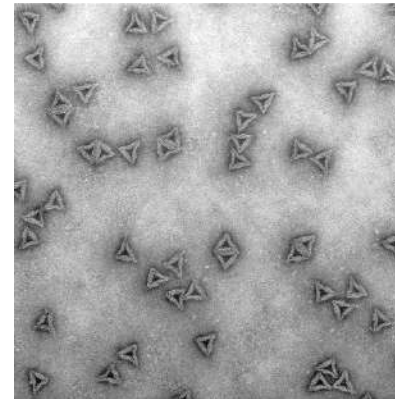
Equalitarian assembly (T=3)

Weak binding – no assembly

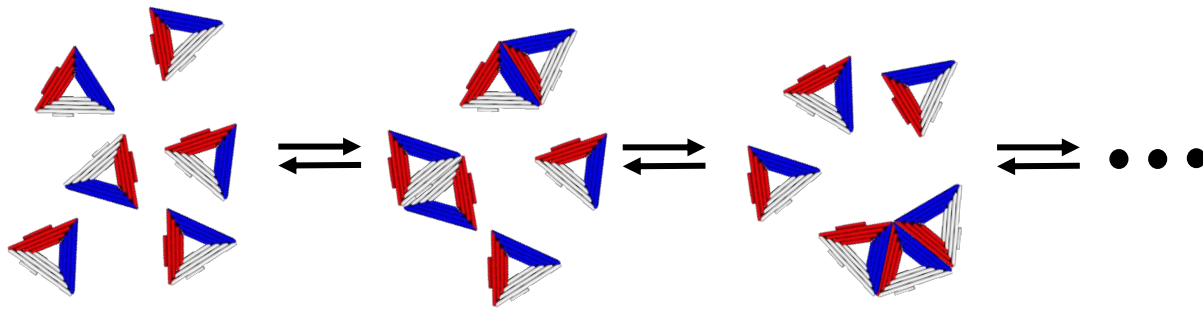
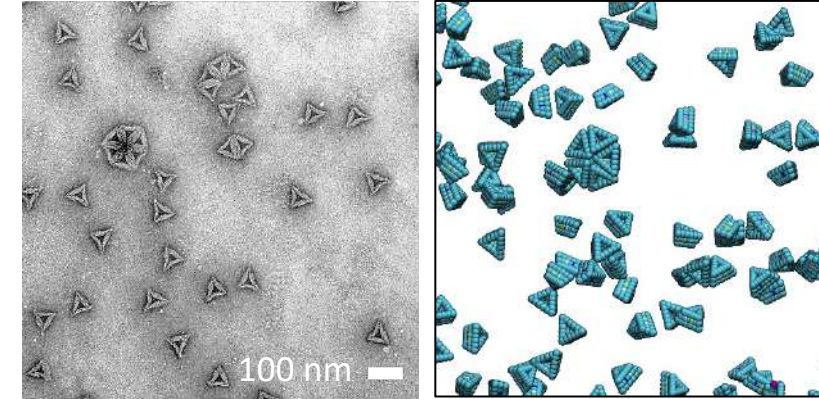
Equal-binding
($-15.0 k_B T$)



early assembly stage



final assembly stage

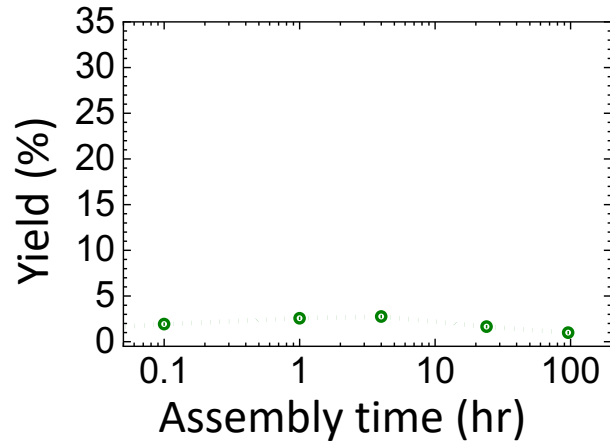
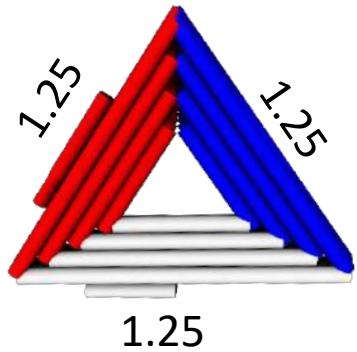


Bonds are too weak.
Nuclei don't form.
Majority subunits remain as monomers.

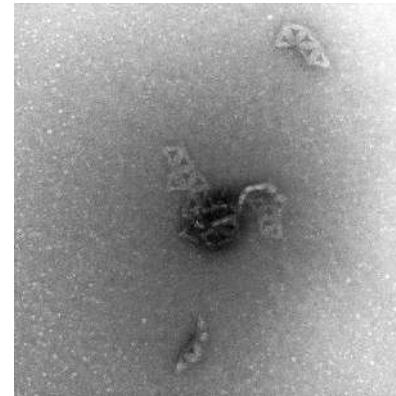
Equalitarian assembly (T=3)

Strong binding – kinetic traps

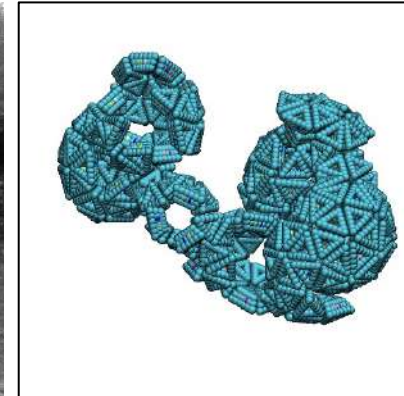
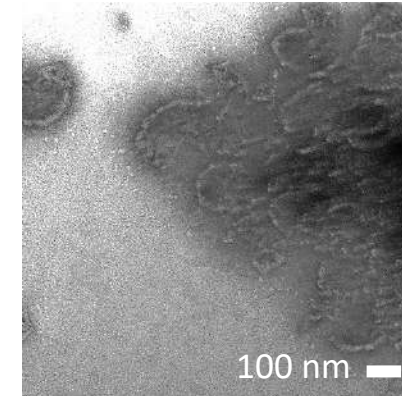
Equal-binding
($-21.7 k_B T$)



early assembly stage



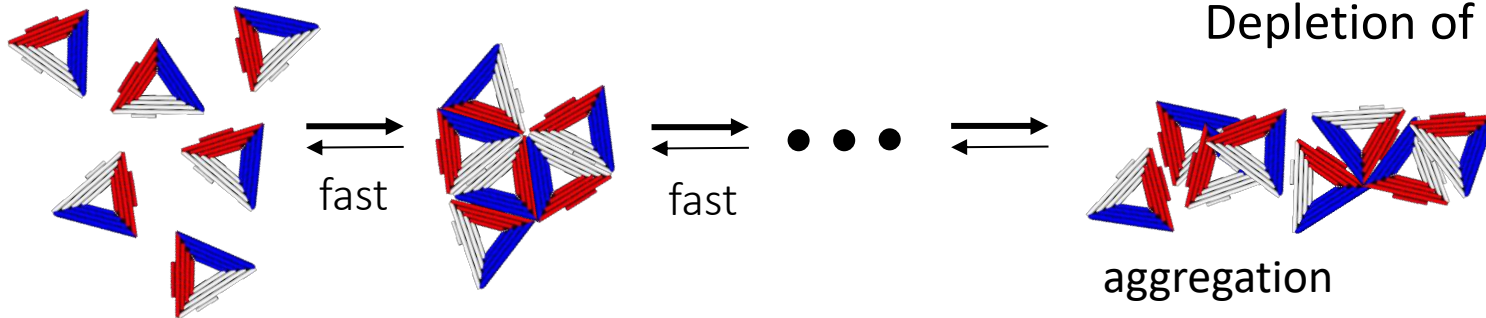
final assembly stage



Bonds are too strong.

Subunits rapidly assemble into intermediate species.

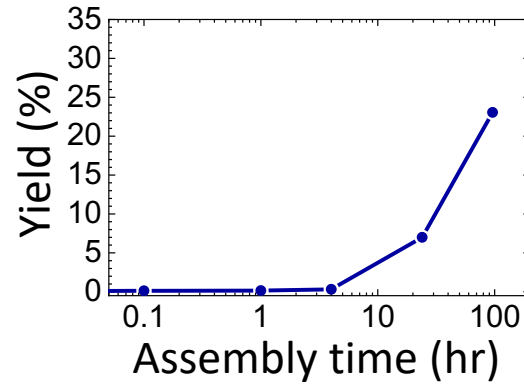
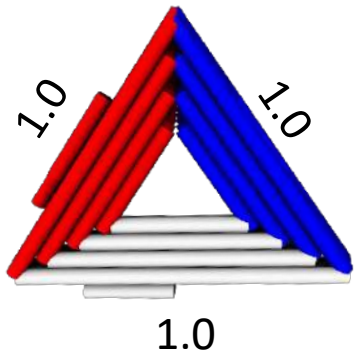
Depletion of monomers and low off-rate forms **kinetic traps**.



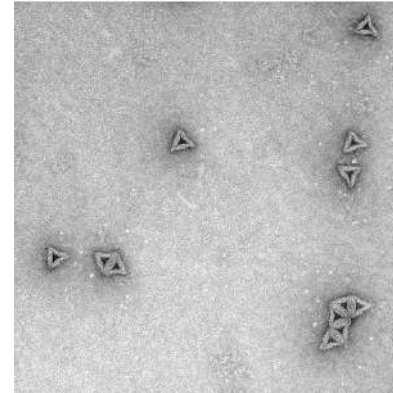
Equalitarian assembly (T=3)

Intermediate binding

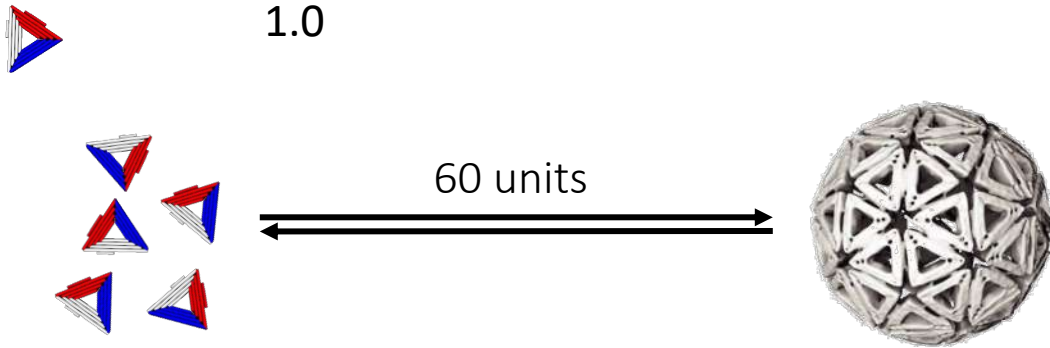
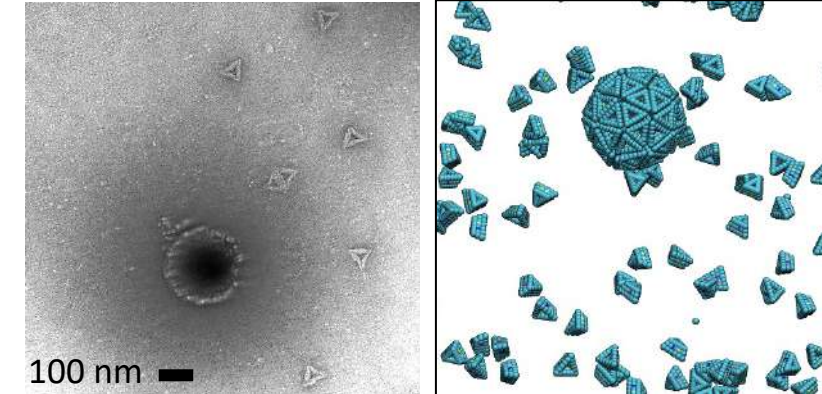
Equal-binding
($-17.4 k_B T$)



early assembly stage



final assembly stage

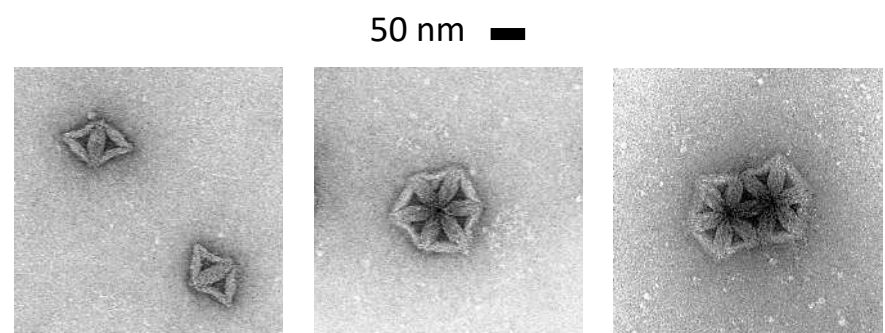


Intermediate species form after a few-hour-long lag phase. Quick consumption of free subunits. Monomer – capsid equilibrium is consistent with a nucleation and growth mechanism.

Goldilocks!

Dimer-bias ($T=3$)

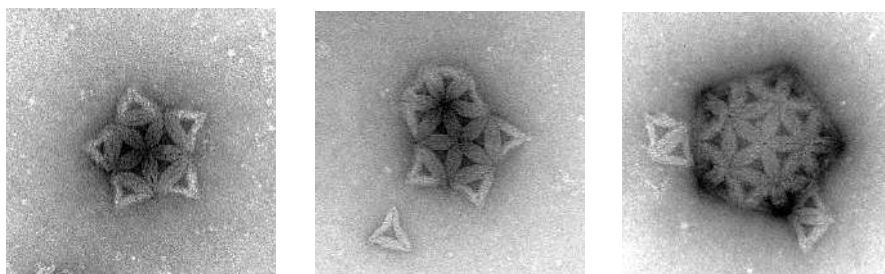
Hierarchical assembly pathways



$N = 2$
1x dimer

$N = 6$
3x dimers

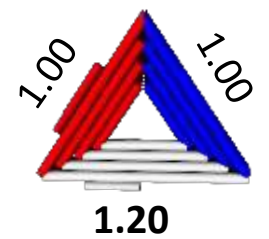
$N = 10$
5x dimers



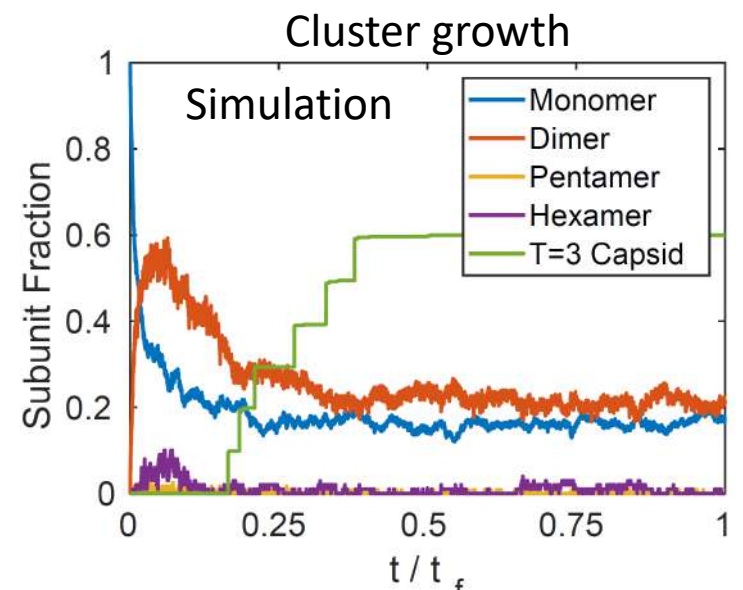
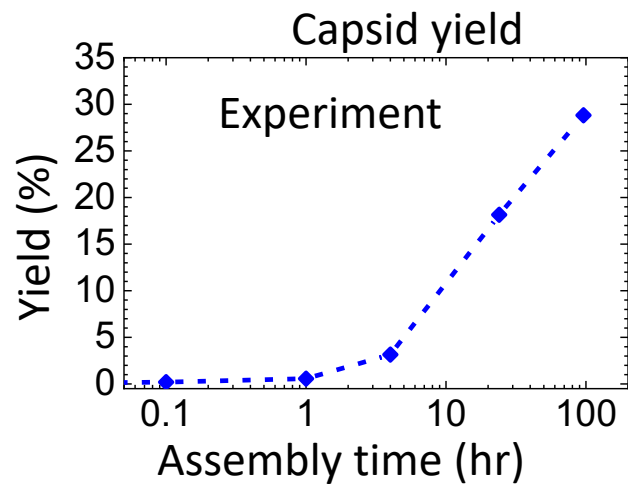
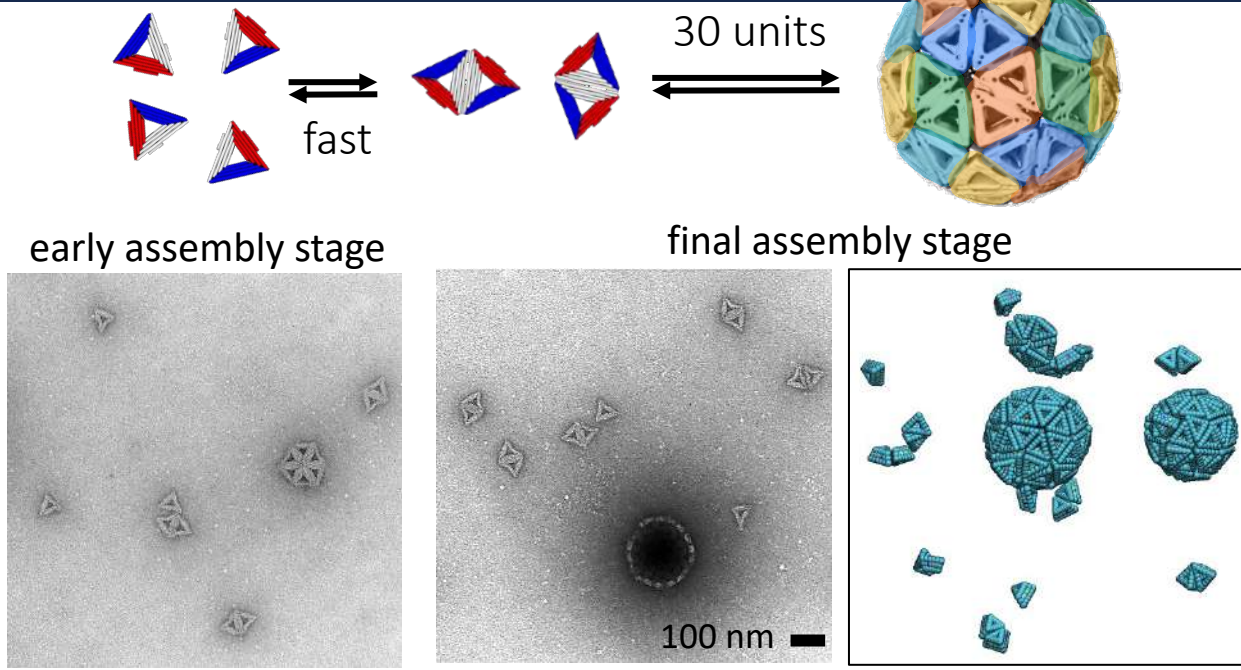
$N = 10$
5x dimers

$N = 12$
6x dimers

$N = 20$
10x dimers

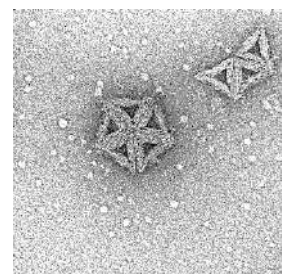


Dimer-bias
(by $3.7 k_B T$)

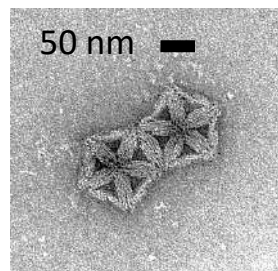


Pentamer bias T=3 assembly

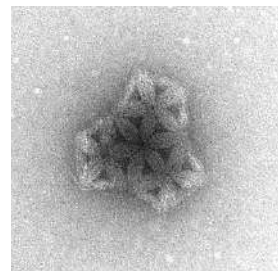
Hierarchical assembly pathways



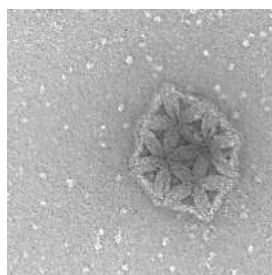
N = 5
1x pentamer



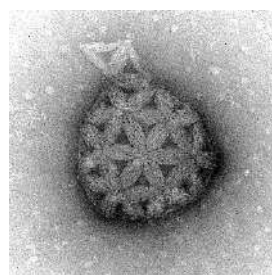
N = 10
2x pentamers



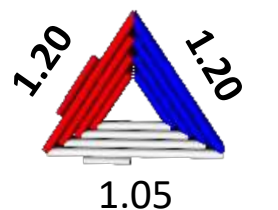
N = 15
3x pentamers



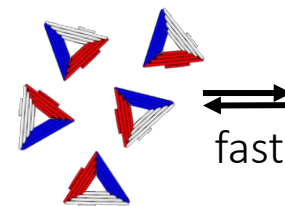
N = 14
2x P + 2x D



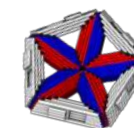
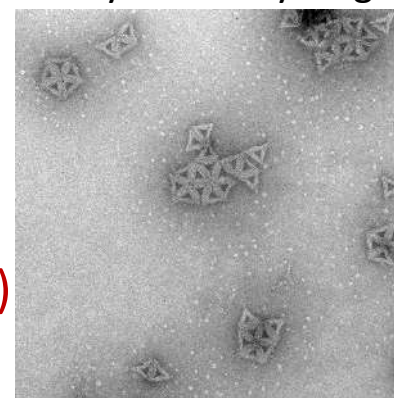
N = 21
3x P + 3x D



Pentamer bias
(by $3.0 k_B T$)



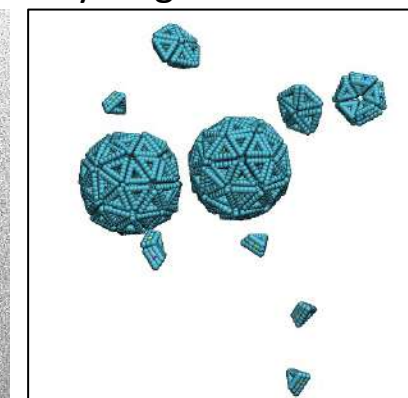
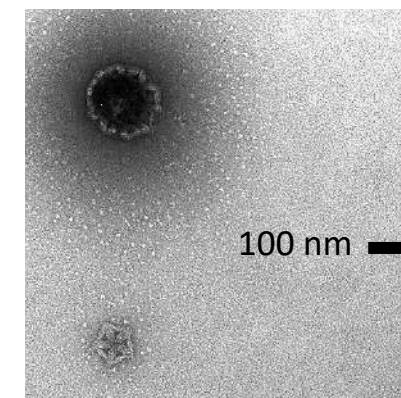
early assembly stage



12 units

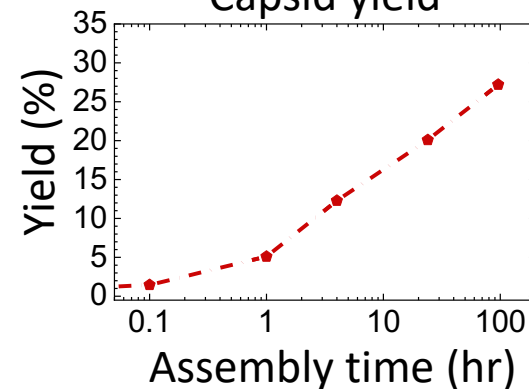


final assembly stage

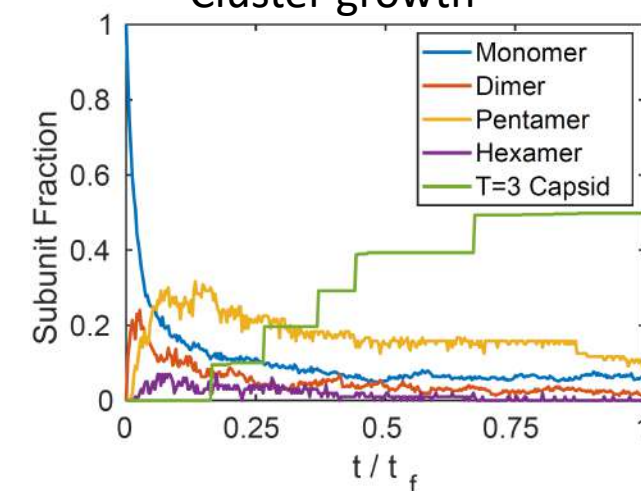


100 nm

Capsid yield



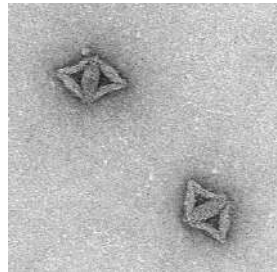
Cluster growth



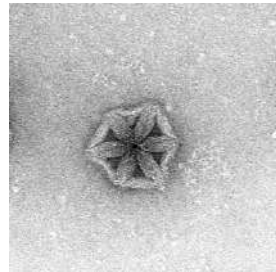
Funneling free energy landscape

Experiment

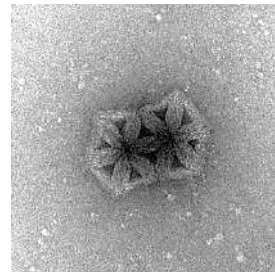
Dimer-bias (by $3.7 k_B T$)



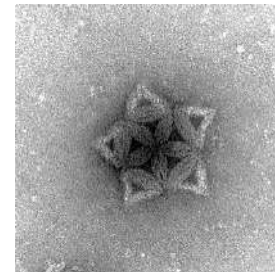
N = 2
1x dimer



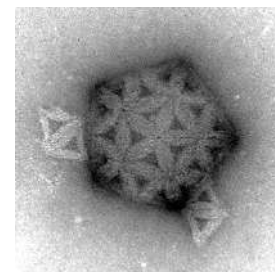
N = 6
3x dimers



N = 10
5x dimers

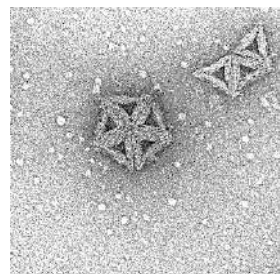


N = 10
5x dimers

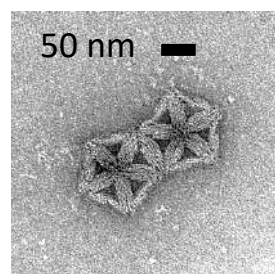


N = 20
10x dimers

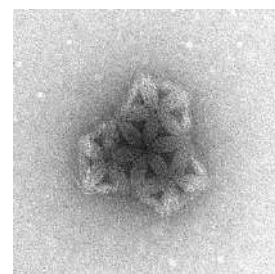
Pentamer-bias (by $3.0 k_B T$)



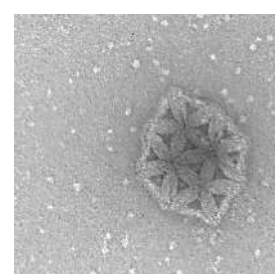
N = 5
1x pentamer



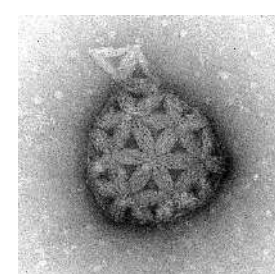
N = 10
2x pentamers



N = 15
3x pentamers

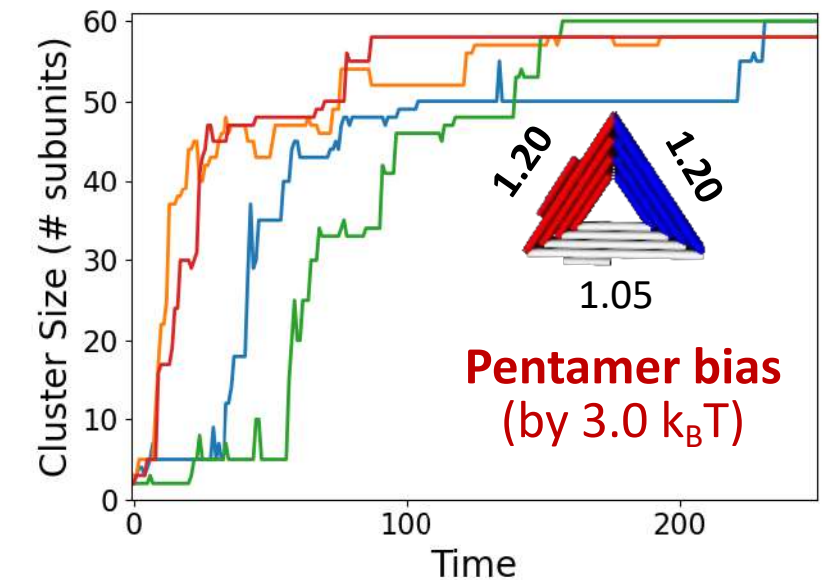
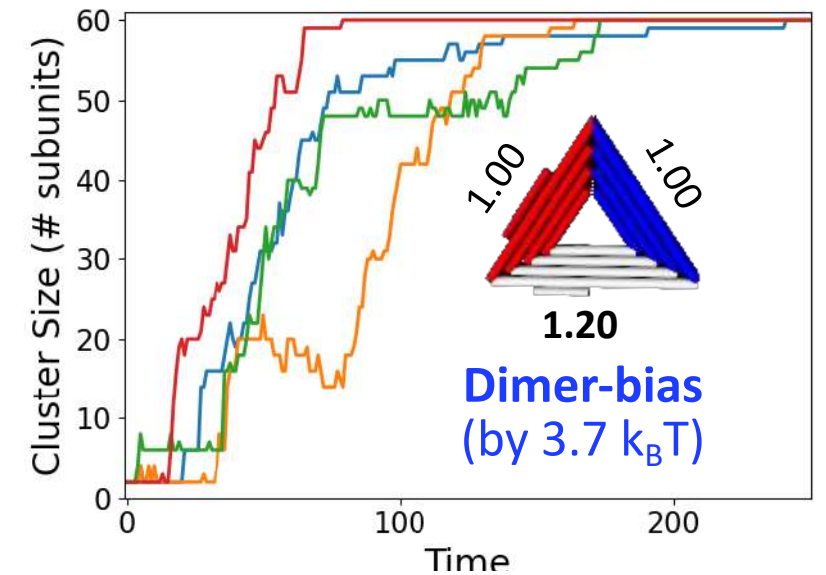


N = 14
2x P + 2x D



N = 21
3x P + 3x D

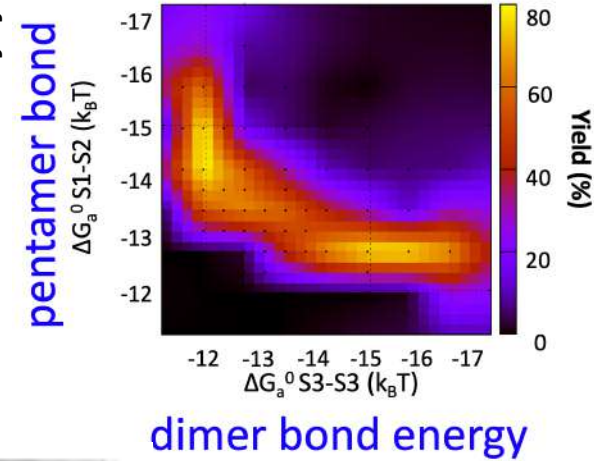
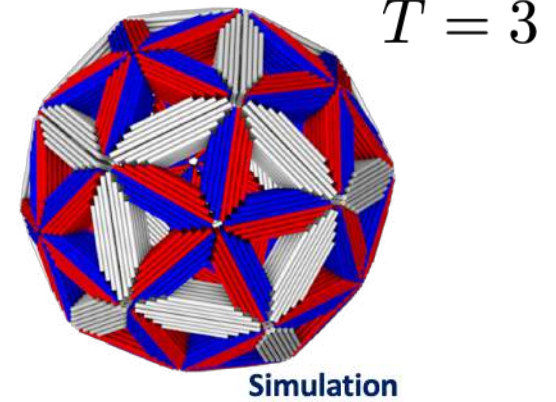
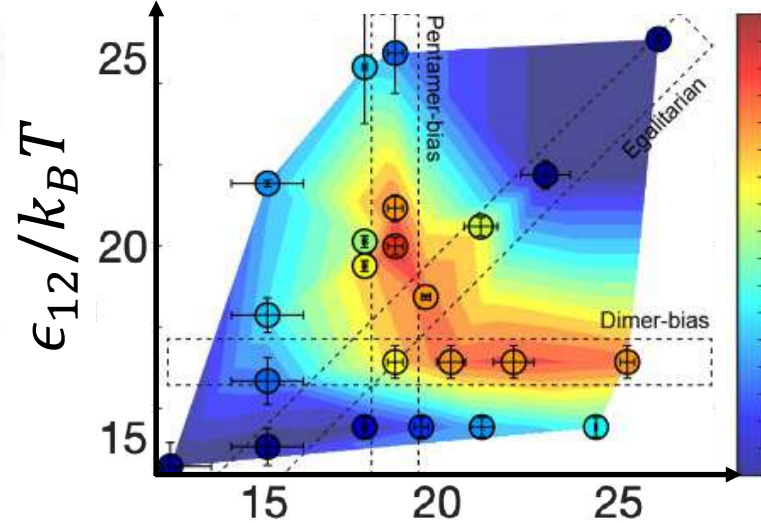
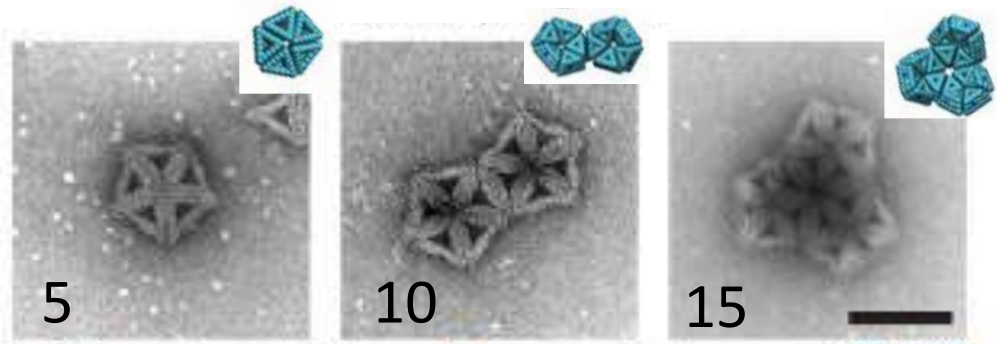
Simulation: Individual trajectories



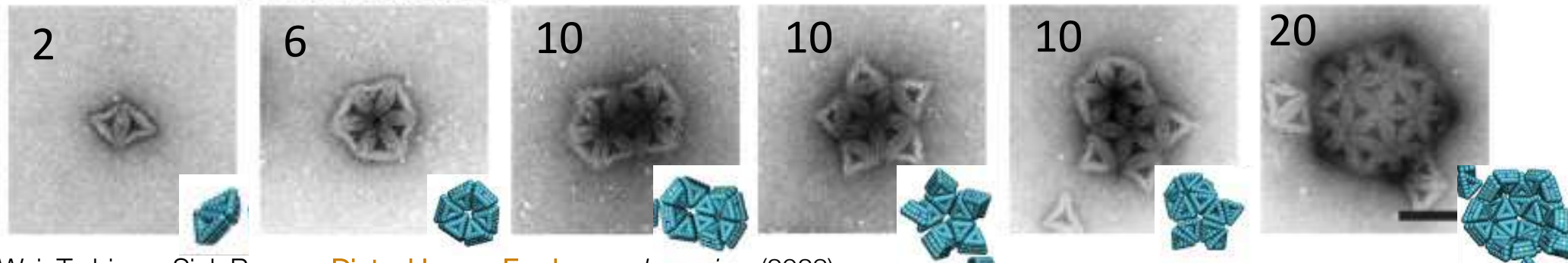
Hierarchical assembly is more robust than egalitarian assembly

Experiments confirm predictions

pentamer pathway intermediates



dimer pathway intermediates



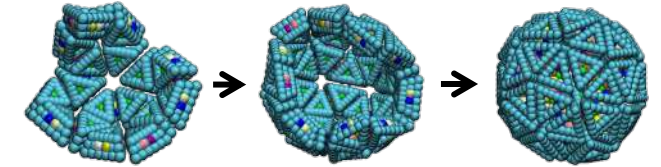
Summary

- ❖ **You can do anything with DNA origami.** Build subunits with programmable bond energy, direction, valency and addressability, for studying self-assembly pathways, kinetics, and yield.

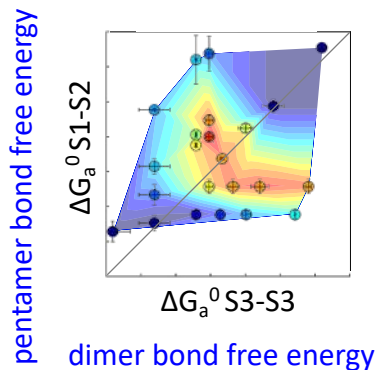


- Construct building blocks with **designed structures in 3D to sub-nm accuracy and bond strengths controlled to $k_B T$ precision**, quantified *in situ* by cryo-EM and static light scattering.

- Coarse-grained representation of subunits, short-ranged potentials, and Langevin dynamics.



- ❖ **Hierarchical assembly is robust and has high yield.**



- **Avoids kinetic traps by funneling intermediate assemblies** towards the designed final target by reducing number of intermediates and excluding off-path structures.



Hierarchical assembly is more robust than egalitarian assembly in synthetic capsids

<https://arxiv.org/abs/2310.18790>

Economical routes to size-specific assembly of self-closing structures

<https://arxiv.org/abs/2311.01383>